



A package for the  
management of NMR data

*Author:*

Francesco Bruno

*Major contributors:*

Letizia Fiorucci

Version: 0.1a.1

Documentation release date:

February 21, 2023



# Contents

<b>1</b>	<b>Introduction</b>	<b>1</b>
<b>2</b>	<b>User guide</b>	<b>2</b>
2.1	Initialize the package . . . . .	2
2.1.1	Extra variables . . . . .	2
2.2	Processing of a 'raw'1D spectrum . . . . .	3
2.2.1	The class <code>pSpectrum_1D</code> . . . . .	5
2.3	Processing of a 'raw'2D spectrum . . . . .	5
2.3.1	Computing projections . . . . .	7
2.4	Simulating data . . . . .	8
2.4.1	Simulate 1D data . . . . .	8
2.4.2	Simulate 2D data . . . . .	9
2.5	The <code>Pseudo_2D</code> class . . . . .	11
<b>3</b>	<b>List of modules and functions</b>	<b>13</b>
3.1	<b>MISC package</b> . . . . .	13
3.1.1	<code>misc.SNR</code> . . . . .	13
3.1.2	<code>misc.SNR_2D</code> . . . . .	14
3.1.3	<code>misc.avg_antidiag</code> . . . . .	15
3.1.4	<code>misc.binomial_triangle</code> . . . . .	16
3.1.5	<code>misc.calcres</code> . . . . .	17
3.1.6	<code>misc.cmap2list</code> . . . . .	18
3.1.7	<code>misc.edit_checkboxes</code> . . . . .	19
3.1.8	<code>misc.find_nearest</code> . . . . .	20
3.1.9	<code>misc.freq2ppm</code> . . . . .	21
3.1.10	<code>misc.get_trace</code> . . . . .	22
3.1.11	<code>misc.get_ylim</code> . . . . .	23
3.1.12	<code>misc.hankel</code> . . . . .	24
3.1.13	<code>misc.hz2pt</code> . . . . .	25
3.1.14	<code>misc.makeacqus_1D</code> . . . . .	26
3.1.15	<code>misc.makeacqus_2D</code> . . . . .	27
3.1.16	<code>misc.mathformat</code> . . . . .	28
3.1.17	<code>misc.molfrac</code> . . . . .	29
3.1.18	<code>misc.noise_std</code> . . . . .	30
3.1.19	<code>misc.nuc_format</code> . . . . .	31
3.1.20	<code>misc.polyn</code> . . . . .	32
3.1.21	<code>misc.ppm2freq</code> . . . . .	33
3.1.22	<code>misc.ppmfind</code> . . . . .	34
3.1.23	<code>misc.pretty_scale</code> . . . . .	35
3.1.24	<code>misc.print_dict</code> . . . . .	36
3.1.25	<code>misc.print_list</code> . . . . .	37

3.1.26	misc.procpar . . . . .	38
3.1.27	misc.readlistfile . . . . .	39
3.1.28	misc.select_for_integration . . . . .	40
3.1.29	misc.select_traces . . . . .	41
3.1.30	misc.set_fontsizes . . . . .	42
3.1.31	misc.set_ylim . . . . .	43
3.1.32	misc.show_cmap . . . . .	44
3.1.33	misc.split_acqus_2D . . . . .	45
3.1.34	misc.split_procs_2D . . . . .	46
3.1.35	misc.trim_data . . . . .	47
3.1.36	misc.unhankel . . . . .	48
3.1.37	misc.write_acqus_1D . . . . .	49
3.1.38	misc.write_acqus_2D . . . . .	50
3.1.39	misc.write_help . . . . .	51
3.1.40	misc.write_ser . . . . .	52
3.2	<b>PROCESSING package . . . . .</b>	<b>53</b>
3.2.1	processing.Cadzow . . . . .	53
3.2.2	processing.Cadzow_2D . . . . .	54
3.2.3	processing.EAE . . . . .	55
3.2.4	processing.LRD . . . . .	56
3.2.5	processing.MCR . . . . .	57
3.2.6	processing.MCR_ALS . . . . .	58
3.2.7	processing.MCR_unpack . . . . .	59
3.2.8	processing.SIMPLISMA . . . . .	60
3.2.9	processing.baseline_correction . . . . .	61
3.2.10	processing.calc_nc . . . . .	62
3.2.11	processing.calibration . . . . .	63
3.2.12	processing.convdt . . . . .	64
3.2.13	processing.em . . . . .	65
3.2.14	processing.fp . . . . .	66
3.2.15	processing.ft . . . . .	67
3.2.16	processing.gm . . . . .	68
3.2.17	processing.gmb . . . . .	69
3.2.18	processing.ift . . . . .	70
3.2.19	processing.integral . . . . .	71
3.2.20	processing.integral_2D . . . . .	72
3.2.21	processing.interactive_basl_windows . . . . .	73
3.2.22	processing.interactive_fp . . . . .	74
3.2.23	processing.interactive_phase_1D . . . . .	75
3.2.24	processing.interactive_phase_2D . . . . .	76
3.2.25	processing.interactive_qfil . . . . .	77
3.2.26	processing.interactive_xfb . . . . .	78
3.2.27	processing.inv_fp . . . . .	79
3.2.28	processing.inv_xfb . . . . .	80
3.2.29	processing.iterCadzow . . . . .	81
3.2.30	processing.load_baseline . . . . .	82
3.2.31	processing.make_polynomion_baseline . . . . .	83
3.2.32	processing.make_scale . . . . .	84
3.2.33	processing.pkn1 . . . . .	85
3.2.34	processing.ps . . . . .	86
3.2.35	processing.qfil . . . . .	87

3.2.36	processing.qpol . . . . .	88
3.2.37	processing.qsin . . . . .	89
3.2.38	processing.quad . . . . .	90
3.2.39	processing.repack_2D . . . . .	91
3.2.40	processing.rev . . . . .	92
3.2.41	processing.sin . . . . .	93
3.2.42	processing.split_echo_train . . . . .	94
3.2.43	processing.stack_MCR . . . . .	95
3.2.44	processing.sum_echo_train . . . . .	96
3.2.45	processing.tabula_rasa . . . . .	97
3.2.46	processing.td_eff . . . . .	98
3.2.47	processing.tp_hyper . . . . .	99
3.2.48	processing.unpack_2D . . . . .	100
3.2.49	processing.write_basl_info . . . . .	101
3.2.50	processing.xfb . . . . .	102
3.2.51	processing.zf . . . . .	103
3.3	<b>FIGURES package</b> . . . . .	104
3.3.1	figures.ax1D . . . . .	104
3.3.2	figures.ax2D . . . . .	106
3.3.3	figures.ax_heatmap . . . . .	108
3.3.4	figures.dotmd . . . . .	109
3.3.5	figures.dotmd_2D . . . . .	110
3.3.6	figures.figure1D . . . . .	111
3.3.7	figures.figure1D_multi . . . . .	112
3.3.8	figures.figure2D . . . . .	113
3.3.9	figures.figure2D_multi . . . . .	115
3.3.10	figures.fitfigure . . . . .	116
3.3.11	figures.heatmap . . . . .	117
3.3.12	figures.plot_fid . . . . .	118
3.3.13	figures.plot_fid_re . . . . .	119
3.3.14	figures.redraw_contours . . . . .	120
3.3.15	figures.sns_heatmap . . . . .	121
3.3.16	figures.stacked_plot . . . . .	122
3.4	<b>SIM package</b> . . . . .	123
3.4.1	sim.calc_splitting . . . . .	123
3.4.2	sim.f_gaussian . . . . .	124
3.4.3	sim.f_lorentzian . . . . .	125
3.4.4	sim.f_pvoigt . . . . .	126
3.4.5	sim.gaussian_filter . . . . .	127
3.4.6	sim.load_sim_1D . . . . .	128
3.4.7	sim.load_sim_2D . . . . .	129
3.4.8	sim.multiplet . . . . .	130
3.4.9	sim.noisegen . . . . .	131
3.4.10	sim.sim_1D . . . . .	132
3.4.11	sim.sim_2D . . . . .	133
3.4.12	sim.t_2Dgaussian . . . . .	134
3.4.13	sim.t_2Dlorentzian . . . . .	135
3.4.14	sim.t_2Dpvoigt . . . . .	137
3.4.15	sim.t_2Dvoigt . . . . .	138
3.4.16	sim.t_gaussian . . . . .	140

3.4.17	sim.t_lorentzian	141
3.4.18	sim.t_pvoigt	142
3.4.19	sim.t_voigt	143
3.4.20	sim.water7	144
3.5	<b>FIT package</b>	145
3.5.1	fit.Voigt_Fit <b>class</b>	145
3.5.2	fit.ax_histogram	147
3.5.3	fit.bin_data	148
3.5.4	fit.build_baseline	149
3.5.5	fit.calc_fit_lines	150
3.5.6	fit.dic2mat	151
3.5.7	fit.fit_int	152
3.5.8	fit.gaussian_fit	153
3.5.9	fit.gen_iguess	154
3.5.10	fit.get_region	155
3.5.11	fit.histogram	156
3.5.12	fit.integrate	157
3.5.13	fit.integrate_2D	158
3.5.14	fit.interactive_smoothing	159
3.5.15	fit.join_par	160
3.5.16	fit.make_iguess	161
3.5.17	fit.make_signal	162
3.5.18	fit.print_par	163
3.5.19	fit.read_par	164
3.5.20	fit.smooth_spl	165
3.5.21	fit.test_residuals	166
3.5.22	fit.vogt_fit	167
3.5.23	fit.write_log	168
3.5.24	fit.write_par	169
3.6	<b>SPECTRA package</b>	170
3.6.1	Spectrum_1D important attributes	170
3.6.2	Spectrum_2D important attributes	171



# 1. Introduction

KLASSEZ is a python package written to handle 1D and 2D NMR data. The aim of the project is to provide a toolkit, consisting of 'black-box' functions organized in modules, that could be used to read, process and analyze such data in a flexible manner, so to adapt to the needs of the individual users. However, the open-source nature of the package grants the user the chance to open the lid of these black-boxes and understand the gears that stand behind the function call.

The development of the toolkit started with `python 3.8` and therefore it is compatible with that version. Nevertheless, the use of `python 3.10` is advised.

The key objects provided by KLASSEZ are the classes `Spectrum_1D` and `Spectrum_2D`, that are able to fulfil the aims of the package with a few lines of code. The classes are able to read both simulated (i.e. generated with a custom-made input file) and experimental datasets. The latter feature was tested with Bruker data after the removal of the digital filter (run command `convdta` in TopSpin), but should be compatible with other kind of spectrometers, thanks to the remarkable work made by J. J. Helmus and coworkers with their `nmrglue` package<sup>1</sup>. Either the FID or the spectrum processed with external solver can be read from KLASSEZ by using the classes `Spectrum_nD` or `pSpectrum_nD`, respectively.

The `processing` module, besides the classical functions used for the processing of NMR data (window functions, Fourier transform, etc.), includes denoising algorithms based on Multivariate Curve Resolution<sup>2</sup> and on Cadzow method<sup>3</sup>. Details are illustrated in the description of the functions.

Functions to show and analyze data in real time are provided, with dedicated GUIs. However, it is better to rely on the standalone functions, enclosed in the single modules, to save the figures. In fact, the `figures` module offers a wide plethora of functions (all based on `matplotlib`) to plot the data with a high degree of customization for the appearance.

The fitting functions use `lmfit` to build the initial guess and to minimize the difference between the experimental data and the model, generated with a Voigt profile in the time domain and then Fourier-transformed, in the least-square sense (employing the Levenberg-Marquardt algorithm implemented in `scipy`). For this purpose, the class `Voigt_fit` of the `fit` module includes attribute functions to construct an initial guess interactively, fit the data, and save the parameters in dedicated files.

Regarding the development of the package, I would like to acknowledge Letizia Fiorucci for her contribution in the design and the implementation of several functions, and for the alpha-testing.

---

<sup>1</sup><https://www.nmrglue.com/>

<sup>2</sup>Multivariate Curve Resolution: 50 years addressing the mixture analysis problem - A review

<sup>3</sup>Denoising NMR time-domain signal by singular-value decomposition accelerated by graphics processing units



## 2. User guide

### 2.1 Initialize the package

Initialize the package by writing, at the top of your file:

---

```
from klassez import *
```

---

This line also implies:

---

```
import os
import sys
import numpy as np
from scipy import linalg, stats
import matplotlib
import matplotlib.pyplot as plt
import matplotlib.cm as cm
from pprint import pprint as Print
```

---

This means these can be not imported in your code, as KLASSEZ already does it for you.

#### 2.1.1 Extra variables

Initializing KLASSEZ also grants access to CM and COLORS.

CM is a dictionary of colormaps taken from `seaborn` and saved in a dictionary whose keys are their names, so that also `matplotlib` can use them. You can inspect the keys through:

---

```
print(CM.keys())
```

---

COLORS is:

---

```
colors = [ 'tab:blue', 'tab:red', 'tab:green', 'tab:orange', 'tab:cyan', 'tab:purple',
           'tab:pink', 'tab:gray', 'tab:brown', 'tab:olive', 'salmon', 'indigo', 'm', 'c', 'g',
           'r', 'b', 'k', ]
```

---

repeated cyclically ten times and stored as tuple.

Other two 'quality of life' variables are `figures.figsize_small` and `figures.figsize_large`, which correspond to figure panel sizes of  $3.59 \times 2.56$  inches and  $15 \times 8$  inches, respectively. The former suits well for saving figures of spectra with font sizes of about 10 pt, whereas the latter are best for GUIs and withstand font sizes of about 14 pt.

For NMR: the variable `sim.gamma` is a dictionary containing the gyromagnetic ratio, in MHz/T, of all the magnetically-active nuclei. For instance:

---

```
print(sim.gamma['13C'])
>>> 10.70611
```

---

## 2.2 Processing of a 'raw' 1D spectrum

Let us say that your spectrum is saved in the folder `/home/myself/spectra/mydataset/1/`. Initialize the spectrum object through:

---

```
Path = "/home/myself/spectra/mydataset/1/"
s = Spectrum_1D(Path)
```

---

This command will do three main tasks:

- read the binary FID of your spectrum and store it in a complex array `s.fid`;
- load the acquisition parameters, read the interesting keys and store them in a dictionary `s.acqus`;
- initialize a dictionary `s.procs` which contains the processing parameters.

If there is the group delay at the beginning of the FID, you are advised to write

---

```
s.convdta()
```

---

to remove it. However, this is tested for Bruker data and does not always work, because it depends on the version of TopSpin that the spectrometer ran and several other construction parameters. Therefore, if you see a residual of the digital filter in your spectrum, the easiest solution is to run `CONVDTA` inside TopSpin.

A detailed description of `acqu`s and `procs` is shown in table 2.1 and table 2.2.

**Table 2.1:** Description of the `acqu`s dictionary of a `Spectrum_1D` object.

Key	Explanation
B0	Magnetic field strength /T
nuc	Observed nucleus
o1p	Carrier frequency i.e. center of the spectrum, in ppm
o1	Same as o1p, but in Hz
SWp	Sweep width, in ppm
SW	Sweep width, in Hz
SF01	Larmor frequency of the observed nucleus at field B0
TD	Number of sampled complex points
dw	Dwell time, i.e. the sampling interval, in seconds
AQ	Time duration of the FID
t1	Acquisition timescale

To make the Fourier transform of the FID to obtain the spectrum, you must invoke the `process` method, which reads the `procs` dictionary to get the instructions on the processing you want to make on your spectrum. For instance, if you want to obtain a final spectrum of 8k points with an exponential broadening of 25 Hz:

---

```
s.procs["wf"]["mode"] = "em"
s.procs["wf"]["lb"] = 25
s.procs["zf"] = 8192
s.process()
```

---

Calling the `process` method generates new attributes of the class:

- `freq`: the frequency scale, in Hz;

**Table 2.2:** Description of the `procs` dictionary of a `Spectrum_1D` object.

Key	Explanation
<code>wf</code>	Window function. This is a dictionary itself: <ul style="list-style-type: none"> <li>• <code>'mode'</code>: choose function between <ul style="list-style-type: none"> <li>– <code>'em'</code>: exponential</li> <li>– <code>'sin'</code>: sine</li> <li>– <code>'qsin'</code>: squared sine</li> <li>– <code>'gm'</code>: mixed lorentzian-gaussian</li> <li>– <code>'gmb'</code>: mixed lorentzian-gaussian, Bruker style</li> </ul> </li> <li>• <code>'lb'</code>: Exponential line-broadening. Read by <code>em</code>, <code>gm</code> and <code>gmb</code></li> <li>• <code>'gb'</code>: Gaussian line-broadening. Read by <code>gm</code> and <code>gmb</code></li> <li>• <code>'gc'</code>: Center of the gaussian <math>\in [0, 1]</math>. Read by <code>gm</code></li> <li>• <code>'ssb'</code>: Shift of the sine bell. Read by <code>sin</code> and <code>qsin</code></li> <li>• <code>'sw'</code>: Sweep width. Automatically set according to <code>acqu['SW']</code></li> </ul>
<code>zf</code>	Zero-filling. Set the <i>final</i> number of points!
<code>tdef</code>	Number of points to be used for processing
<code>fcor</code>	Scaling factor for the first point of the FID before Fourier transform
<code>p0</code>	Frequency-independent phase correction /degrees
<code>p1</code>	First order phase correction /degrees
<code>pv</code>	Pivot point for the first order phase correction /ppm

- `ppm`: the ppm scale;
- `r`: the real part of the spectrum;
- `i`: the imaginary part of the spectrum;
- `S`: the complex spectrum ( $S = r + ii$ ).

If the spectrum requires phase correction, you can perform it interactively:

---

```
s.adjph()
```

---

or by passing the phase angles, in degrees, to `adjph`. Example, if you know you need to phase your spectrum with 30 degrees of  $\phi^{(0)}$  and  $-55$  degrees of  $\phi^{(1)}$  with the pivot set at 7.32 ppm:

---

```
s.adjph(p0=30, p1=-55, pv=7.32)
```

---

In both cases, the phase angles are updated in the `procs` dictionary.

The spectrum can be calibrated using a dedicated GUI:

---

```
s.cal()
```

---

or specifying the shift value in ppm or in Hz (in this case, be sure to set the `isHz` keyword to `True`).

---

```
s.cal(-3)           # Shift of -3 ppm
s.cal(1000, isHz=True) # Shift of +1 kHz
```

---

Both ppm and freq are updated according to the given values.

A tool for baseline correction and lineshape fitting are also provided:

---

```
s.basl()           # build baseline
s.r -= s.baseline # subtract the baseline
```

---

```
# Make an initial guess interactively and save all the parameters in a file called
  "myguess.inp"
s.F.iguess(input_file="myguess.inp")
# Save a figure of the initial guess in "myguess.png"
s.F.plot(what='iguess', name="myguess")
# Do the fit
s.F.dofit(
    log_file="myfit.log",      # Write the log of the fit in a file called "myfit.log"
    output_file="myfit.out",   # Write the output of my fit in a file called "myfit.out"
    utol=0.5,                  # Tolerance on the chemical shift of +/-0.5 ppm
    vary_phi=False,            # Allow/Not allow to fit dephased peaks
    vary_xg=False,             # Allow/Not allow to change fraction of gaussianity
    res_hist_name="myres",     # Make a figure of the residuals called "myres.png"
    test_res=True              # Perform tests on the goodness of the fit
)
# Plot the output and show it
s.F.plot(what='fit')
```

---

### 2.2.1 The class pSpectrum\_1D

The class `Spectrum_1D` does not work if you want to read the processed data directly from TopSpin (or whatever software you used to acquire and process them). Instead, you should use the class `pSpectrum_1D`, which is designed to perform exactly this task. It inherits most of the attributes and methods of the `Spectrum_1D` class, therefore its usage closely resembles the example reported in the previous section.

## 2.3 Processing of a 'raw' 2D spectrum

Let us say that your spectrum is saved in the folder `/home/myself/spectra/mydataset/21/`. Initialize the spectrum object through:

---

```
Path = "/home/myself/spectra/mydataset/21/"
s = Spectrum_2D(Path)
```

---

The generated `acqus` and `procs` dictionaries include informations on both dimensions.

Then, the sequence of commands resembles the ones of the 1D spectra.

---

```
s.convdta()      # If there is the digital filter
s.process()
s.adjph()
s.plot()
```

---

**Table 2.3:** Description of the `acqus` dictionary of a `Spectrum_2D` object.

Key	Explanation
<code>B0</code>	Magnetic field strength /T
<code>nuc1</code>	Observed nucleus in the indirect dimension
<code>nuc2</code>	Observed nucleus in the direct dimension
<code>o1p</code>	Carrier frequency i.e. center of the indirect dimension, in ppm
<code>o2p</code>	Carrier frequency i.e. center of the direct dimension, in ppm
<code>o1</code>	Same as <code>o1p</code> , but in Hz
<code>o2</code>	Same as <code>o2p</code> , but in Hz
<code>SW1p</code>	Sweep width of the indirect dimension, in ppm
<code>SW2p</code>	Sweep width of the direct dimension, in ppm
<code>SW1</code>	Sweep width of the indirect dimension, in Hz
<code>SW2</code>	Sweep width of the indirect dimension, in Hz
<code>SF01</code>	Larmor frequency of the observed nucleus in F1 at field <code>B0</code>
<code>SF02</code>	Larmor frequency of the observed nucleus in F2 at field <code>B0</code>
<code>TD1</code>	Number of $t_1$ -increments
<code>TD2</code>	Number of sampled complex points
<code>dw1</code>	$t_1$ increments, in seconds
<code>dw2</code>	Dwell time, i.e. the sampling interval, in seconds
<code>AQ1</code>	Sampled timescale of the indirect dimension
<code>AQ2</code>	Time duration of the FID
<code>t1</code>	Evolution timescale
<code>t2</code>	Acquisition timescale

The keys for `adjph` are of the kind: `pXY`, where `X` is the order of the phase correction (0 or 1) and `Y` is the dimension on which to apply it (1 or 2). Explicative table below:

	<b>F1</b>	<b>F2</b>
$\phi^{(0)}$	<code>p01</code>	<code>p02</code>
$\phi^{(1)}$	<code>p11</code>	<code>p12</code>
<code>pivot</code>	<code>pv1</code>	<code>pv2</code>

For further information, rely on the `help` python builtin function.

To read the processed data, use the `pSpectrum_2D` class instead.

**Table 2.4:** Description of the `procs` dictionary of a `Spectrum_2D` object. Each of these dictionary entry is a list of two elements: the first one (index 0) is the processing to apply on the indirect dimension, the second (index 1) on the direct dimension. For instance, `procs[tdeff] = [64, 1024]` means to truncate the indirect evolutions to 64 points and the FIDs to 1024 points.

Key	Explanation
<code>wf</code>	Window function. This is a dictionary itself: <ul style="list-style-type: none"> <li>• <code>'mode'</code>: choose function between <ul style="list-style-type: none"> <li>– <code>'em'</code>: exponential</li> <li>– <code>'sin'</code>: sine</li> <li>– <code>'qsin'</code>: squared sine</li> <li>– <code>'gm'</code>: mixed lorentzian-gaussian</li> <li>– <code>'gmb'</code>: mixed lorentzian-gaussian, Bruker style</li> </ul> </li> <li>• <code>'lb'</code>: Exponential line-broadening. Read by <code>em</code>, <code>gm</code> and <code>gmb</code></li> <li>• <code>'gb'</code>: Gaussian line-broadening. Read by <code>gm</code> and <code>gmb</code></li> <li>• <code>'gc'</code>: Center of the gaussian <math>\in [0, 1]</math>. Read by <code>gm</code></li> <li>• <code>'ssb'</code>: Shift of the sine bell. Read by <code>sin</code> and <code>qsin</code></li> <li>• <code>'sw'</code>: Sweep width. Automatically set according to <code>acqus['SW']</code></li> </ul>
<code>zf</code>	Zero-filling. Set the <i>final</i> number of points!
<code>tdeff</code>	Number of points to be used for processing
<code>fcor</code>	Scaling factor for the first point of the FID before Fourier transform
<code>p0</code>	Frequency-independent phase correction /degrees
<code>p1</code>	First order phase correction /degrees
<code>pv</code>	Pivot point for the first order phase correction /ppm

### 2.3.1 Computing projections

While the 2D spectra give an overall look on the whole experiment, the user might want to extract projection of the direct or the indirect dimension, to focus onto particular features in the spectrum. In order to do so, `klassez` offers two commands: `projf1` and `projf2`, which compute the sum projections on the indirect or on the direct dimension, respectively, and store the result in dictionaries called `trf1` and `trf2`, whose keys are the ppm values correspondent to the projections. Actually, the capitalized versions of the two dictionaries (with the same keys), i.e. `Trf1` and `Trf2`, can be more useful, as they are instances of the `pSpectrum_1D` class and therefore are initialized with ppm scales and other parameters.

Example:

---

```
# Supposed to have a 1H-15N HSQC spectrum

# Extract the direct dimension trace at 115 ppm, 15N scale
s.projf2(115)
# Access to it through
Proj_115 = s.Trf2['115']
```

```
# Extract the indirect dimension trace from 6 to 8 ppm, 1H scale
s.proj1(6, 8)
Proj_indim = s.Trf1['6:8']

# You can plot them:
Proj_115.plot()
Proj_indim.plot()
```

---

## 2.4 Simulating data

The classes `Spectrum_1D` and `Spectrum_2D` are also able to generate simulated data by reading a custom-written input file. The functions they use are `sim.sim_1D` and `sim.sim_2D`.

### 2.4.1 Simulate 1D data

The input file you have to write *must* have the following keys:

- **B0**: Magnetic field strength /T;
- **nuc**: Observed nucleus (e.g. `13C`);
- **o1p**: Carrier frequency i.e. centre of the spectrum /ppm;
- **SWp**: Sweep width /ppm. The spectrum will cover the range  $[\text{o1p} - \text{SWp}/2, \text{o1p} + \text{SWp}/2]$ ;
- **TD**: Number of sampled (complex) points;
- **shifts**: sequence of peak positions /ppm;
- **fwhm**: Full-width at half-maximum of the peaks /Hz;
- **amplitudes**: Intensity of the peaks in the FID;
- **x\_g**: Fraction of gaussianity.  $x_g = 0 \implies$  pure Lorentzian peak,  $x_g = 1 \implies$  pure Gaussian peak;

and *can* have the following keys:

- **phases**: phases of the peaks /degrees. Default: all zeros;
- **mult**: fine structures of the peaks (e.g. doublets of triplets: `dt`). Default: all singlets;
- **Jconst**: coupling constants of the fine structures /Hz. If more of one coupling is expected, provide them as a sequence. Default: not used as the peaks are all singlets.

Key and value must be separated by a tab character. You are allowed to leave empty rows to improve the readability and to insert comments using the `#` character.

Example:

---

```
B0 16.4    # 700 MHz 1H
nuc 1H
o1p 4.7
SWp 40
TD 8192

shifts 1, 3, 5, 7
```

```

fwhm    [10 for k in range(4)]
amplitudes 10, 20, 15, 10
x_g 0, 0.4, 0.6, 1
phases 5, 0, 10, 0

mult    s, t, dt, ddd
Jconst 0, 15, [12, 9.5], [25, 15, 10]

```

---

This input file generates the spectrum in Figure 2.1.

Code:

---

```

#!/usr/bin/env python3

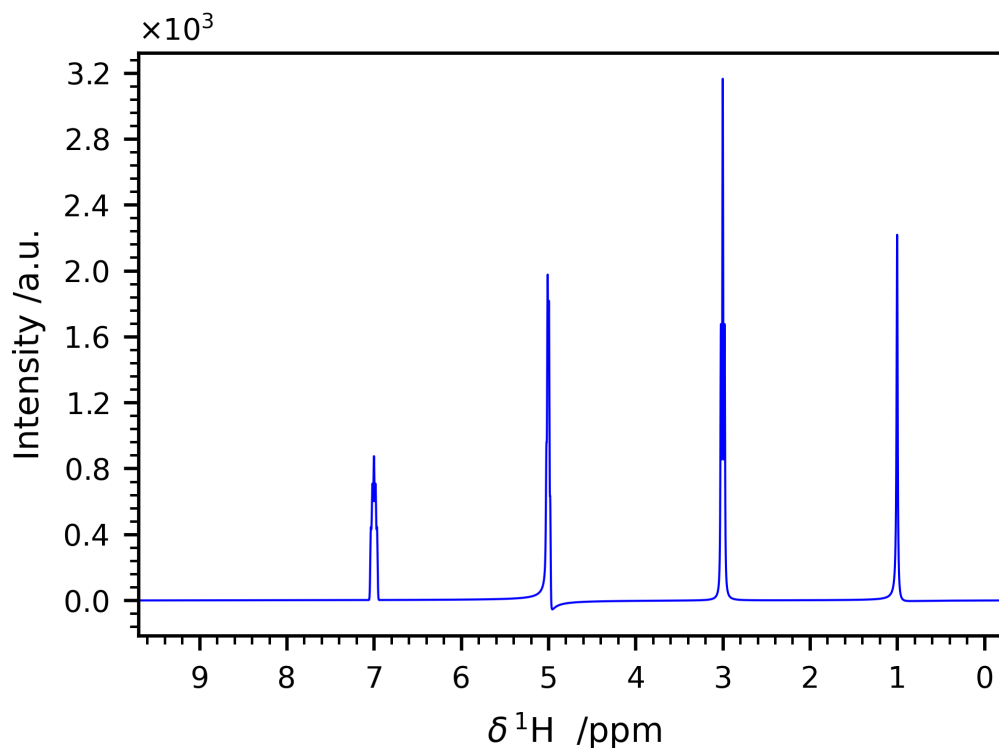
from klassez import *

s = Spectrum_1D('sim_in_1D', isexp=False)
s.process()

figures.figure1D(s.ppm, s.r, name='test_1D', X_label='$\delta$, ^1$H /ppm',
    Y_label='Intensity /a.u.')

```

---



**Figure 2.1:** Example of a simulated 1D spectrum.

## 2.4.2 Simulate 2D data

The same procedure can be followed to simulate 2D spectra. The input file to write is very similar to the one for 1D data, except for the quantities that clearly span over two dimensions. As in NMR textbook, the direct and indirect dimensions will be named F2 and F2 respectively, and dimension-specific quantities will feature the 1 or 2 labels accordingly.

- B0: Magnetic field strength /T;



- `nuc1`: Observed nucleus in F1(e.g.  $^{13}\text{C}$ );
- `nuc2`: Observed nucleus in F2(e.g.  $^1\text{H}$ );
- `o1p`: Carrier frequency i.e. centre of F1 /ppm;
- `o2p`: Carrier frequency i.e. centre of F2 /ppm;
- `SW1p`: Sweep width /ppm. The indirect dimension will cover the range  $[\text{o1p} - \text{SW1p}/2, \text{o1p} + \text{SW1p}/2]$ ;
- `SW2p`: Sweep width /ppm. The direct dimension will cover the range  $[\text{o2p} - \text{SW2p}/2, \text{o2p} + \text{SW2p}/2]$ ;
- `TD1`: Number of sampled (complex) points in F1;
- `TD2`: Number of sampled (complex) points in F2;
- `shifts_f1`: sequence of peak positions in F1 /ppm;
- `shifts_f2`: sequence of peak positions in F2 /ppm;
- `fwhm_f1`: Full-width at half-maximum of the peaks in F1 /Hz;
- `fwhm_f2`: Full-width at half-maximum of the peaks in F2 /Hz;
- `amplitudes`: Intensity of the peaks in the FID;
- `x_g`: Fraction of gaussianity.  $x_g = 0 \implies$  pure Lorentzian peak,  $x_g = 1 \implies$  pure Gaussian peak;

Phase distortions and fine structures are not allowed for multidimensional spectra. The indirect dimension will be generated employing the *States-TPPI* sampling scheme.

Example:

---

```

B0 28.2
nuc1 15N
nuc2 1H
o1p 115
o2p 5
SW1p 40
SW2p 20
TD1 512
TD2 8192

shifts_f1 130.0, 105.0, 120.0, 1.25e2, 130.0, 105.0
shifts_f2 0.0, 0.0, 4.0, 7.0, 1.1e1, 10.5
fwhm_f1 100, 100, 100, 100, 100, 100
fwhm_f2 50, 50, 50, 50, 50, 50
amplitudes 10, 20, 10, 20, 10, 10
x_g 0.0, 0.2, 0.4, 0.6, 0.8, 1.0

```

---

This input file generates the spectrum in Figure 2.2.

Code:

---

```
#!/usr/bin/env python3
```

```
from klassez import *
```

```
s = Spectrum_2D('sim_in_2D', isexp=False)
s.process()
```

```
figures.figure2D(s.ppm_f2, s.ppm_f1, s.rr, lvl=0.005, name='test_2D', X_label='$\delta$',
    ^1$H /ppm', Y_label='$\delta$', ^{15}$N /ppm')
```

---

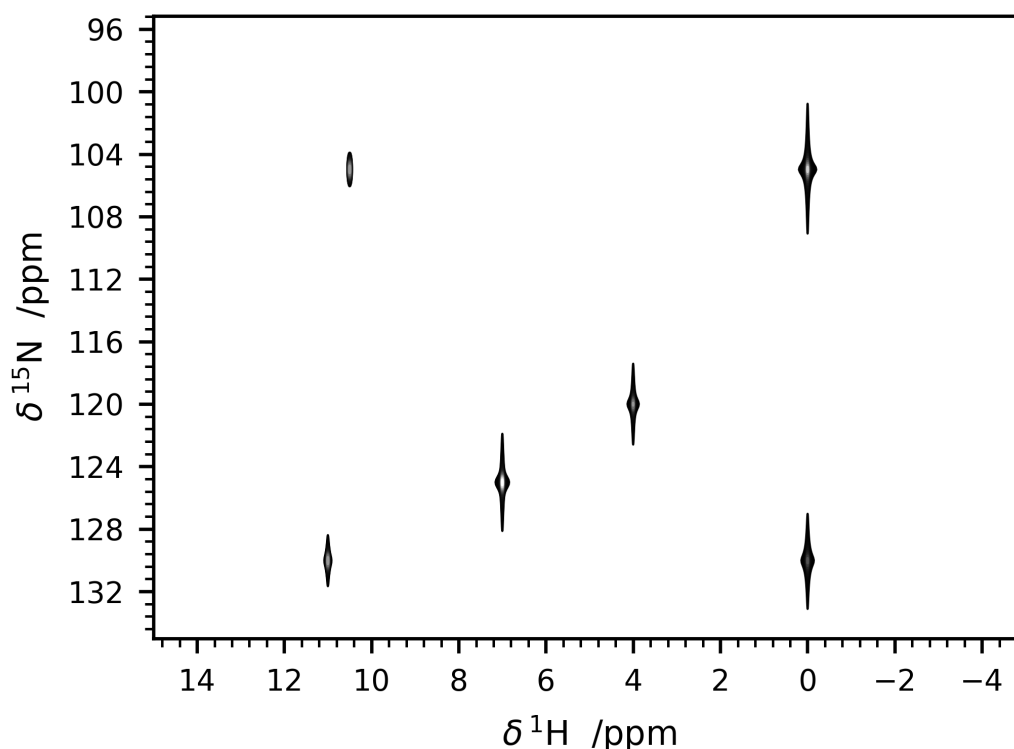


Figure 2.2: Example of a simulated 2D spectrum.

## 2.5 The Pseudo\_2D class

Sometimes, the spectroscopist might find interesting to acquire a series of 1D experiments in which one (or more) parameters are changed according to a certain schedule. This kind of experiments are 2D in principle, but their processing and analysis resemble the one of 1D spectra. Therefore, they lie somewhere in between 1D spectra and 2D spectra, hence they are often referred to as *pseudo\_2D*.

Also in this case, *klassez* offers a specific class to deal with this kind of data: *Pseudo\_2D*. *Pseudo\_2D* is a subclass of *Spectrum\_2D*; however, many functions have been adapted to resemble the 1D version.

*Pseudo\_2D* does not encode for a routine to automatically simulate data. If you want to, you should give a 1D-like input file (just like the one in section 2.4.1), and replace the attribute *fid* with your FID, generated as you wish. With a real dataset this is not required, as it is able to read everything automatically.

---

```
path_to_pseudo = "/home/myself/spectra/mydataset/899/"
s = Pseudo_2D(path_to_pseudo)
s.convdta() # If there is the digital filter
```

---

The `process()` function applies apodization, zero-filling and Fourier transform only on the direct dimension, reading the parameters from a `procs` dictionary like the one of `Spectrum_1D`. The attributes `freq_f1` and `ppm_f1` are initialized with `np.arange(N)`, where  $N$  is the number of experiments that your FID comprises of.

The phase adjustment is performed on a reference spectrum, then applied on the whole 2D matrix. By default, the chosen spectrum is the first one, but you can choose the one that fits the most your needs.

---

```
s.process()
s.adjph(expno = 10) # Calls interactive_phase_1D on the 10th experiment
```

---

The method `plot` shows the 2D contour map of the spectrum, just like the one of `Spectrum_2D`. However, this is not always the most intelligent way to plot the data in order to gather information. This is the reason why this class features two unique additional methods that plot data: `plot_md` and `plot_stacked`. Both rely on the parameter `which`, that is a string of code (i.e. it should be interpreted by `eval`) that identifies which experiment to show by pointing at their index. `which = 'all'` results in pointing at all spectra.

---

```
s.plot() # 2D contour map
s.plot_md(which="3, 5, 11") # Plot the 3rd, the 5th and the 11th spectrum, superimposed
s.plot_stacked(which="np.arange(0,100,5)") # Makes a stacked plot with a spectrum every 5
```

---

The method `integrate` differs a little bit from the one coded in `Spectrum_1D`.

---

```
s.integrate(which=2) # Interactive panel on the 3rd spectrum
```

---

Even if you select the integration limits on a single spectrum, the method `integrate` will compute the integrals throughout the whole range of experiment. This means that each entry of `integrals` will be an array as long as the number of experiment.

## 3. List of modules and functions

### 3.1 MISC package

This package contains miscellaneous functions for the calculation of several properties, and generally for the handling of NMR spectra.

#### 3.1.1 `misc.SNR(data, signal=None, n_reg=None)`

Computes the signal to noise ratio of a 1D spectrum, as:

$$SNR = \frac{S}{2\sigma_n}$$

##### Parameters

- `data` : *1darray*  
The spectrum of which you want to compute the SNR
- `signal` : *float, optional*  
If provided, uses this value as maximum signal. Otherwise, it is selected as the maximum value in `data`
- `n_reg` : *list or tuple, optional*  
If provided, contains the points that delimit the noise region. Otherwise, the whole spectrum is used.

##### Returns

- `snr` : *float*  
The SNR of the spectrum
-

### 3.1.2 misc.SNR\_2D(data, n\_reg=None)

Computes the signal to noise ratio of a 2D spectrum.

#### Parameters

- data : *1darray*  
The spectrum of which you want to compute the SNR
- n\_reg : *list or tuple*  
If provided, the points of F1 scale and F2 scale, respectively, of which to extract the projections.  
Otherwise, opens the tool for interactive selection.

#### Returns

- snr\_f1 : *float*  
The SNR of the indirect dimension
  - snr\_f2 : *float*  
The SNR of the direct dimension
-

### 3.1.3 misc.avg\_antidiag(X)

Given a matrix  $X$  without any specific structure, finds the closest Hankel matrix in the Frobenius norm sense by averaging the antidiagonals.

**Parameters:**

- $X$ : *2darray*  
Input matrix

**Returns:**

- $X_p$ : *2darray*  
Hankel matrix obtained from  $X$
-

### 3.1.4 misc.binomial\_triangle(n)

Calculates the n-th row of the binomial triangle. The first row is n=1, not 0. Example:

---

```
binomial_triangle(4)
>>> 1 3 3 1
```

---

#### Parameters:

- n: *int*  
Row index

#### Returns:

- row: *1darray*  
The n-th row of binomial triangle.
-

### 3.1.5 misc.calcrs(fqscale)

Calculates the frequency resolution of an axis scale, i.e. how many Hz is a 'tick'.

#### Parameters

- `fqscale` : *1darray*  
Scale to be processed

#### Returns

- `res` : *float*  
The resolution of the scale
-



### 3.1.6 misc.cmap2list(cmap, N=10, start=0, end=1)

Extract the colors from a colormap and return it as a list.

#### Parameters:

- **cmap:** *matplotlib.Colormap Object*  
The colormap from which you want to extract the list of colors
- **N:** *int*  
Number of samples to extract
- **start:** *float*  
Start point of the sampling. 0 = beginning of the cmap; 1 = end of the cmap.
- **end:** *float*  
End point of the sampling. 0 = beginning of the cmap; 1 = end of the cmap.

#### Returns:

- **colors:** *list*  
List of the extracted colors.
-

### 3.1.7 `misc.edit_checkboxes(checkbox, xadj=0, yadj=0, length=None, height=None, color=None)`

Edit the size of the box to be checked, and adjust the lines accordingly.

#### Parameters:

- `checkbox`: *matplotlib.widgets.CheckBox Object*  
The checkbox to edit
  - `xadj`: *float*  
modifier value for bottom left corner x-coordinate of the rectangle, in `checkbox.ax` coordinates
  - `yadj`: *float*  
modifier value for bottom left corner y-coordinate of the rectangle, in `checkbox.ax` coordinates
  - `length`: *float*  
length of the rectangle, in `checkbox.ax` coordinates
  - `height`: *float*  
height of the rectangle, in `checkbox.ax` coordinates
  - `color`: *str or list or None*  
If it is not `None`, change color to the lines
-

### 3.1.8 misc.find\_nearest(array, value)

Finds the value in `array` which is the nearest to `value`.

#### Parameters

- `array` : *1darray*  
Self-explanatory
- `value` : *float*  
Value to be found

#### Returns

- `val` : *float*  
The closest value in `array` to `value`
-

### 3.1.9 misc.freq2ppm(x, B0=701.125, o1p=0)

Converts x from Hz to ppm.

#### Parameters

- x : *float*  
Value to be converted
- B0 : *float*  
Field frequency, in MHz. Default: 700 MHz
- o1p : *float*  
Carrier frequency, in ppm. Default: 0.

#### Returns

- y : *float*  
The converted value
-

### 3.1.10 `misc.get_trace(data, ppm_f2, ppm_f1, a, b=None, column=True)`

Takes as input a 2D dataset and the ppm scales of direct and indirect dimensions respectively. Calculates the projection on the given axis summing from `a` (ppm) to `b` (ppm). Default: indirect dimension projection (i.e. `column=True`), change it to `'False'` for the direct dimension projection. Returns the calculated 1D projection.

#### Parameters

- `data` : *2darray*  
Spectrum of which to extract the projections
- `ppm_f2` : *1darray*  
ppm scale of the direct dimension
- `ppm_f1` : *1darray*  
ppm scale of the indirect dimension
- `a` : *float*  
The ppm value from which to start extracting the projection.
- `b` : *float, optional*  
If provided, the ppm value at which to stop extracting the projection. Otherwise, returns only the `a` trace.
- `column` : *bool*  
If `True`, extracts the F1 projection. If `False`, extracts the F2 projection.

#### Returns

- `y` : *1darray*  
Computed projection
-

### 3.1.11 `misc.get_ylim(data_inp)`

Calculates the y-limits of a plot as follows:

- Bottom:  $\min(\text{data}) - 5\% \max(\text{data})$
- Top:  $\max(\text{data}) + 5\% \max(\text{data})$

#### Parameters:

- `data_inp`: *ndarray or list*  
Input data. If it is a list, `data_inp` is converted to array.
-

### 3.1.12 misc.hankel(data, n=None)

Computes a Hankel matrix from `data`. If `data` is a `1darray` of length  $N$ , computes the correspondent Hankel matrix of dimensions  $(N - n + 1, n)$ . If `data` is a `2darray`, computes the closest Hankel matrix in the Frobenius norm sense by averaging the values on the antidiagonals.

#### Parameters:

- `data`: *1darray*  
Vector to be Hankel-ized, of length  $N$
- `n`: *int*  
Number of columns that the Hankel matrix will have

#### Returns:

- `H`: *2darray*  
Hankel matrix of dimensions  $(N - n + 1, n)$
-

### 3.1.13 misc.hz2pt(fqscale, hz)

Converts `hz` from frequency units to points, on the basis of its scale.

#### Parameters

- `fqscale` : *1darray*  
Scale to be processed
- `hz` : *float*  
Value to be converted

#### Returns

- `pt` : *float*  
The frequency value converted in points
-



### 3.1.14 misc.makeacqus\_1D(dic)

Given a NMRGLUE dictionary from a 1D spectrum (generated by `ng.bruker.read`), this function builds the `acqus` file with only the 'important' parameters.

#### Parameters

- `dic` : *dict*  
NMRglue dictionary returned by `ng.bruker.read`

#### Returns

- `acqus` : *dict*  
Dictionary with only few parameters
-

### 3.1.15 misc.makeacqus\_2D(dic)

Given a NMRGLUE dictionary from a 2D spectrum (generated by `ng.bruker.read`), this function builds the `acqus` file with only the 'important' parameters.

#### Parameters

- `dic` : *dict*  
NMRglue dictionary returned by `ng.bruker.read`

#### Returns

- `acqus` : *dict*  
Dictionary with only few parameters
-

### 3.1.16 `misc.mathformat(ax, axis='y', limits=(-2,2))`

Apply exponential formatting to the given axis of the given figure panel. The offset text size is uniformed to the tick labels' size.

#### Parameters:

- `ax`: *matplotlib.Subplot Object*  
Panel of the figure to edit
  - `axis`: *str*  
'x', 'y' or 'both'.
  - `limits`: *tuple*  
tuple of ints that indicate the order of magnitude range outside which the exponential format is applied.
-

### 3.1.17 misc.molfrac(n)

Computes the 'molar fraction'  $\mathbf{x}$  of the array  $\mathbf{n}$ . Returns also the total amount.

$$x_i = \frac{n_i}{N}, \quad N = \sum_i n_i$$

#### Parameters

- $\mathbf{n}$ : *list or 1darray*  
list of values

#### Returns

- $\mathbf{x}$ : *list or 1darray*  
molar fraction array
  - $N$ : *float*  
sum of all the elements in  $\mathbf{n}$
-

### 3.1.18 `misc.noise_std(y)`

Calculates the standard deviation of the noise using the Bruker formula. Taken  $y$  as an array of  $N$  points, and  $y_i$  its  $i$ -th entry:

$$\sigma_n = \frac{1}{\sqrt{N-1}} \sqrt{\sum_{i=1}^N y_i^2 - \frac{1}{N} \left[ \left( \sum_{i=1}^N y_i \right)^2 + \frac{3 \left( \sum_{i=1}^{N/2} i (y_{N-i+1} - y_i)^2 \right)}{N^2 - 1} \right]}$$

#### Parameters

- $y$  : *1darray*

The spectral region you would like to use to calculate the standard deviation of the noise.

#### Returns

- `noisestd` : *float*

The standard deviation of the noise.

---

### 3.1.19 misc.nuc\_format(nuc)

Converts the `nuc` key you may find in `acqus` in the formatted label, e.g. `'13C'`  $\rightarrow$  `'${13}$C'`

#### Parameters:

- `nuc`: *str*  
Unformatted string

#### Returns:

- `fnuc`: *str*  
Formatted string.
-

### 3.1.20 misc.polyn(x, c)

Computes  $p(x)$ , polynomial of degree  $n - 1$ , where  $n$  is the number of provided coefficients.

#### Parameters

- $x$  : *1darray*  
Scale upon which to build the polynomial
- $c$  : *list or 1darray*  
Sequence of the polynomial coefficient, starting from the 0-th order coefficient

#### Returns

- $px$  : *1darray*  
Polynomial of degree  $n - 1$ .
-

### 3.1.21 `misc.ppm2freq(x, B0=701.125, o1p=0)`

Converts `x` from ppm to Hz.

#### Parameters

- `x` : *float*  
Value to be converted
- `B0` : *float*  
Field frequency, in MHz. Default: 700 MHz
- `o1p` : *float*  
Carrier frequency, in ppm. Default: 0.

#### Returns

- `y` : *float*  
The converted value
-



### 3.1.22 misc.ppmfind(ppm\_scale, value)

Finds the exact value in ppm\_scale.

#### Parameters

- ppm\_scale : *1darray*  
Self-explanatory
- value : *float*  
The value to be found

#### Returns

- I : *int*  
The index correspondent to V in ppm\_scale
  - V : *float*  
The closest value to value in ppm\_scale
-

### 3.1.23 `misc.pretty_scale(ax, limits, axis='x', n_major_ticks=10)`

This function computes a pretty scale for your plot. Calculates and sets a scale made of `n_major_ticks` numbered ticks, spaced by `5 * n_major_ticks` unnumbered ticks. After that, the plot borders are trimmed according to the given limits.

#### Parameters

- `ax`: *matplotlib.AxesSubplot object*  
Panel of the figure of which to calculate the scale
  - `limits`: *tuple*  
limits to apply of the given axis. (`left`, `right`)
  - `axis`: *str*  
'x' for x-axis, 'y' for y-axis
  - `n_major_ticks`: *int*  
Number of numbered ticks in the final scale. An oculated choice gives very pleasant results.
-

### 3.1.24 misc.print\_dict(mydict)

Prints a dictionary one entry per row, in the format `key: value`. Nested dictionaries are printed with an indentation

#### Parameters:

- mydict: *dict*  
The dictionary you want to print

#### Returns:

- outstring: *str*  
The printed text formatted as single string
-

### 3.1.25 `misc.print_list(mylist)`

Prints a list, one entry per row.

#### Parameters:

- `mylist`: *list*  
The list you want to print

#### Returns:

- `outstring`: *str*  
The printed text formatted as single string
-

### 3.1.26 misc.procpars(txt)

Takes as input the path of a file containing a 'key' in the first column and a 'value' in the second column. Returns a dictionary of shape **key** : **value**.

#### Parameters

- txt : *str*  
Path to a file that contains 'key' in first column and 'value' in the second

#### Returns

- procpars : *dict*  
Dictionary of shape **key** : **value**.
-

### 3.1.27 misc.readlistfile(datafile)

Takes as input the path of a file containing one entry for each row. Returns a list of the aforementioned entries.

#### Parameters

- datafile: *str*  
Path to a file that contains one entry for each row

#### Returns

- files : *list*  
List of the entries contained in the file
-

### 3.1.28 `misc.select_for_integration(ppm_f1, ppm_f2, data, Neg=True)`

Select the peaks of a 2D spectrum to integrate. First, select the area where your peak is located by dragging the red square. Then, select the center of the peak by right\_clicking. Finally, click 'ADD' to store the peak. Repeat the procedure for as many peaks as you want.

#### Parameters:

- `ppm_f1` : *1darray*  
ppm scale of the indirect dimension
- `ppm_f2` : *1darray*  
ppm scale of the direct dimension
- `data` : *2darray*  
Spectrum
- `Neg` : *bool*  
Choose if to show the negative contours ( `True`) or not (`False` )

#### Returns:

- `peaks`: *list of dict*  
For each peak there are two keys, '`f1`' and '`f2`', whose meaning is obvious. For each of these keys, you have '`u`': center of the peak /ppm, and '`lim`': the limits of the square you drew before.
-

### 3.1.29 misc.select\_traces(ppm\_f1, ppm\_f2, data, Neg=True, grid=False)

Select traces from a 2D spectrum, save the coordinates in a list. Left click to select a point, right click to remove it.

#### Parameters

- ppm\_f1 : *1darray*  
ppm scale of the indirect dimension
- ppm\_f2 : *1darray*  
ppm scale of the direct dimension
- data : *2darray*  
Spectrum
- Neg : *bool*  
Choose if to show the negative contours (**True**) or not (**False**)
- grid : *bool*  
Choose if to display the grid (**True**) or not (**False**)

#### Returns

- coord: *list*  
List containing the [x,y] coordinates of the selected points.
-



### 3.1.30 `misc.set_fontsizes(ax, fontsize=10)`

Automatically adjusts the fontsizes of all the figure elements. In particular:

- title = `fontsize`
- axis labels = `fontsize - 2`
- ticks labels = `fontsize - 3`
- legend entries = `fontsize - 4`

#### Parameters:

- ax: *matplotlib.Subplot Object*  
Subplot of interest
  - fontsize: *float*  
Starting fontsize
-

### 3.1.31 `misc.set_ylim(ax, data_inp)`

Sets the y-limits of `ax` as follows:

- Bottom:  $\min(\text{data}) - 5\% \max(\text{data})$
- Top:  $\max(\text{data}) + 5\% \max(\text{data})$

#### Parameters:

- `ax`: *matplotlib.Subplot Object*  
Panel of the figure where to apply this scale
  - `data_inp`: *ndarray or list*  
Input data. If it is a list, `data_inp` is converted to array.
-

### 3.1.32 `misc.show_cmap(cmap, N=10, start=0, end=1)`

Plot the colors extracted from a colormap.

#### Parameters:

- `cmap`: *matplotlib.Colormap Object*  
The colormap from which you want to extract the list of colors
  - `N`: *int*  
Number of samples to extract
  - `start`: *float*  
Start point of the sampling. 0 = beginning of the cmap; 1 = end of the cmap.
  - `end`: *float*  
End point of the sampling. 0 = beginning of the cmap; 1 = end of the cmap.
-

### 3.1.33 misc.split\_acqus\_2D(acqus)

Split the acqus dictionary of a 2D spectrum into two separate 1D-like acqus dictionaries.

#### Parameters:

- acqus: *dict*  
acqus dictionary of a 2D spectrum

#### Returns:

- acqu1s: *dict*  
acqus dictionary of the indirect dimension
  - acqu2s: *dict*  
acqus dictionary of the direct dimension
-

### 3.1.34 misc.split\_procs\_2D(procs)

Split the procs dictionary of a 2D spectrum into two separate 1D-like procs dictionaries.

#### Parameters

- procs: *dict*  
procs dictionary of a 2D spectrum

#### Returns

- proc1s: *dict*  
procs dictionary of the indirect dimension
  - proc2s: *dict*  
procs dictionary of the direct dimension
-

### 3.1.35 `misc.trim_data(ppm_scale, y, sx, dx)`

Trims the frequency scale and correspondant 1D dataset `y` from `sx` (ppm) to `dx` (ppm).

#### Parameters

- `ppm_scale` : *1darray*  
ppm scale of the spectrum
- `y` : *1darray*  
spectrum
- `sx` : *float*  
ppm value where to start trimming
- `dx` : *float*  
ppm value where to finish trimming

#### Returns

- `xtrim` : *1darray*  
Trimmed ppm scale
  - `ytrim` : *1darray*  
Trimmed spectrum
-

### 3.1.36 misc.unhankel(H)

Concatenates the first row and the last column of the matrix H, which should have Hankel-like structure, so to build the array of independent parameters.

#### Parameters:

- H: *2darray*  
Hankel-like matrix

#### Returns:

- h: *1darray*  
First row and last column of H, concatenated
-

### 3.1.37 misc.write\_acqus\_1D(acqus, path='.', filename=None)

Writes the input file for a simulated spectrum, basing on a dictionary of parameters.

#### Parameters

- `acqu` : *dict*  
The dictionary containing the parameters for the simulation
  - `path` : *str, optional*  
Directory where the file will be saved.
  - `filename`: *str, optional*  
Name of the file to be saved in `path`. The default name is `sim_in_1D`
-



### 3.1.38 `misc.write_acqus_2D`(`acqus`, `path`='sim\_in\_2D')

Writes the input file for a simulated spectrum, basing on a dictionary of parameters.

#### Parameters

- `acqus` : *dict*  
The dictionary containing the parameters for the simulation
  - `path` : *str, optional*  
Directory where the file will be saved. The default name is `sim_in_1D`.
-

### 3.1.39 misc.write\_help(request, file=None)

Gets the documentation of `request`, and tries to save it in a text file.

#### Parameters:

- `request`: *function or class or package*  
Whatever you need documentation of
  - `file`: *str or None or False*  
Name of the output documentation file. If it is `None`, a default name is given. If it is `False`, the output is printed on screen.
-

### 3.1.40 misc.write\_ser(path, fid)

Writes the FID file in directory `path`, in a TopSpin-readable way (i.e. little endian, int32). The binary file is named *'fid'* if 1D, *'ser'* if multiD.

Apparently, bidimensional spectra are not saved correctly.

#### Parameters

- `path` : *str*  
Directory where to save the file
  - `fid` : *ndarray*  
FID array to be written
-

## 3.2 PROCESSING package

This package contains functions for the processing of NMR spectra, either in time domain or in frequency domain, and the transition between the two domains.

---

### 3.2.1 processing.Cadzow(data, n, nc, print\_head=True)

This functions performs Cadzow denoising on `data`, which is a 1D array of  $N$  points. The algorithm works as follows:

1. Transform `data` in a Hankel matrix  $\mathbb{H}$  of dimensions  $(N - n, n)$
2. Make SVD on  $\mathbb{H} = \mathbb{U}\mathbb{S}\mathbb{V}^\dagger$
3. Keep only the first `nc` singular values, and put all the rest to 0 ( $\mathbb{S} \rightarrow \mathbb{S}'$ )
4. Rebuild  $\mathbb{H}' = \mathbb{U}\mathbb{S}'\mathbb{V}^\dagger$
5. Average the antidiagonals to rebuild the Hankel-type structure, then make 1D array

#### Parameters

- `data`: *1darray*  
Input data
- `n`: *int*  
Number of columns of the Hankel matrix.
- `nc`: *int*  
Number of singular values to keep.
- `print_head`: *bool*  
Set it to `True` to display the fancy heading.

#### Returns

- `datap`: *1darray*  
Denoised data
-

### 3.2.2 `processing.Cadzow_2D(data, n, nc, i=True, itermax=100, f=0.005, print_time=True)`

Performs the Cadzow denoising method on a 2D spectrum, one transient at the time. This function calls `Cadzow` if `i=False`, or `iterCadzow` if `i=True`.

#### Parameters

- `data`: *2darray*  
Input data
- `n`: *int*  
Number of columns of the Hankel matrix.
- `nc`: *int*  
Number of singular values to keep.
- `i`: *bool*  
Calls `processing.Cadzow` if `i=False`, or `processing.iterCadzow` if `i=True`.
- `itermax`: *int*  
Maximum number of iterations allowed.
- `f`: *float*  
Factor for the arrest criterion.
- `print_time`: *bool*  
Set it to `True` to display the time spent.

#### Returns

- `datap`: *2darray*  
Denoised data
-

### 3.2.3 processing.EAE(data)

Shuffles data if the spectrum is acquired with '*FnMODE*':'*Echo-Antiecho*'. **NOTE:** introduces -90° phase shift in F1, to be corrected after the processing.

---

```
pdata = np.zeros_like(data)
pdata[:,2] = (data[:,2].real - data[1:,2].real) + 1j*(data[:,2].imag - data[1:,2].imag)
pdata[1:,2] = -(data[:,2].imag + data[1:,2].imag) + 1j*(data[:,2].real + data[1:,2].real)
```

---

#### Parameters

- data : *ndarray*  
Data to be shuffled.

#### Returns

- pdata : *ndarray*  
Shuffled data.
-

### 3.2.4 processing.LRD(data, nc)

Denoising method based on Low-Rank Decomposition. The algorithm performs a singular value decomposition on `data`, then keeps only the first `nc` singular values while setting all the others to 0. Finally, rebuilds the data matrix using the modified singular values.

#### Parameters:

- `data`: *2darray*  
Data to be denoised
- `nc`: *int*  
Number of components, i.e. number of singular values to keep

#### Returns:

- `data_out`: *2darray*  
Denoised data
-

### 3.2.5 `processing.MCR(input_data, nc, f=10, tol=1e-5, itermax=1e4, H=True, oncols=True)`

This is an implementation of Multivariate Curve Resolution for the denoising of 2D NMR data.

Let us consider a matrix  $\mathbb{D}$ , of dimensions  $m \times n$ , where the starting data are stored. The final purpose of MCR is to decompose the  $\mathbb{D}$  matrix as follows:

$$\mathbb{D} = \mathbb{C}\mathbb{S} + \mathbb{E}$$

where  $\mathbb{C}$  and  $\mathbb{S}$  are matrices of dimension  $m \times \text{nc}$  and  $\text{nc} \times n$ , respectively, and  $\mathbb{E}$  contains the part of the data that are not reproduced by the factorization.

Being  $\mathbb{D}$  the FID of a NMR spectrum,  $\mathbb{C}$  will contain time evolutions of the indirect dimension, and  $\mathbb{S}$  will contain transients in the direct dimension.

The total MCR workflow can be separated in two parts: a first algorithm that produces an initial guess for the three matrices  $\mathbb{C}$ ,  $\mathbb{S}$  and  $\mathbb{E}$  (**SIMPLISMA**), and an optimization step that aims at the removal of the unwanted features of the data by iteratively filling the  $\mathbb{E}$  matrix (**MCR ALS**).

This function returns the denoised datasets,  $\mathbb{CS}$ , and the single  $\mathbb{C}$  and  $\mathbb{S}$  matrices.

#### Parameters

- `input_data`: *2darray or 3darray*  
a 3D array containing the set of 2D NMR datasets to be coprocessed stacked along the first dimension. A single 2D array can be passed, if the denoising of a single dataset is desired.
- `nc`: *int*  
number of purest components to be looked for;
- `f`: *float*  
percentage of allowed noise;
- `tol`: *float*  
tolerance for the arrest criterion;
- `itermax`: *int*  
maximum number of allowed iterations
- `H`: *bool*  
`True` for horizontal stacking of data (default), `False` for vertical;
- `oncols`: *bool*  
`True` to estimate  $\mathbb{S}$  with `processing.SIMPLISMA`, `False` to estimate  $\mathbb{C}$ .

#### Returns

- `CS_f`: *2darray or 3darray*  
Final denoised data matrix
  - `C_f`: *2darray or 3darray*  
Final  $\mathbb{C}$  matrix
  - `S_f`: *2darray or 3darray*  
Final  $\mathbb{S}$  matrix
-



### 3.2.6 processing.MCR\_ALS(D, C, S, itermax=10000, tol=1e-5)

Performs alternating least squares to get the final  $\mathbb{C}$  and  $\mathbb{S}$  matrices. Being the fundamental MCR equation:

$$\mathbb{D} = \mathbb{C}\mathbb{S} + \mathbb{E}$$

At the  $k$ -th step of the iterative cycle:

1.  $\mathbb{C}^{(k)} = \mathbb{D}\mathbb{S}^{+(k-1)}$
2.  $\mathbb{S}^{(k)} = \mathbb{C}^{+(k)}\mathbb{D}$
3.  $\mathbb{E}^{(k)} = \mathbb{D} - \mathbb{C}^{(k)}\mathbb{S}^{(k)}$

Defined  $r_C$  and  $r_S$  as the Frobenius norm of the difference of  $\mathbb{C}$  and  $\mathbb{S}$  matrices between two subsequent steps:

$$r_C = \|\mathbb{C}^{(k)} - \mathbb{C}^{(k-1)}\|_F \quad r_S = \|\mathbb{S}^{(k)} - \mathbb{S}^{(k-1)}\|_F$$

The convergence is reached when:

$$r_C \leq \text{tol} \quad \wedge \quad r_S \leq \text{tol}$$

#### Parameters

- **D**: *2darray*  
Input data, of dimensions  $m \times n$
- **C**: *2darray*  
Estimation of the  $\mathbb{C}$  matrix, of dimensions  $m \times \text{nc}$ .
- **S**: *2darray*  
Estimation of the  $\mathbb{S}$  matrix, of dimensions  $\text{nc} \times n$ .
- **itermax**: *int*  
Maximum number of iterations
- **tol**: *float*  
Threshold for the arrest criterion.

#### Returns

- **C**: *2darray*  
Optimized  $\mathbb{C}$  matrix, of dimensions  $m \times \text{nc}$ .
  - **S**: *2darray*  
Optimized  $\mathbb{S}$  matrix, of dimensions  $\text{nc} \times n$ .
-

### 3.2.7 processing.MCR\_unpack(C, S, nds, H=True)

Reverts matrix augmentation of `stack_MCR`. If `H=True`, converts `C` from dimensions  $(Y, \text{nds})$  to  $(X, Y, \text{nds})$  and `S` from dimensions  $(\text{nds}, X * Z)$  to  $(X, \text{nds}, Z)$ ; if `H=False` converts `C` from dimensions  $(Y, \text{nds})$  to  $(X, Y, \text{nds})$  and `S` from dimensions  $(\text{nds}, X * Z)$  to  $(X, \text{nds}, Z)$ .

#### Parameters

- `C`: *2darray*  
MCR  $\mathbb{C}$  matrix
- `S`: *2darray*  
MCR  $\mathbb{S}$  matrix
- `nds`: *int*  
Number of datasets to be unpacked.
- `H`: *bool*  
`True` for horizontal stacking, `False` for vertical stacking.

#### Returns

- `C_f`: *3darray*  
Not-augmented  $\mathbb{C}$  matrix.
  - `S_f`: *3darray*  
Not-augmented  $\mathbb{S}$  matrix.
-

### 3.2.8 processing.SIMPLISMA(D, nc, f=10, oncols=True)

Finds the first `nc` purest components of matrix `D` using the SIMPLISMA algorithm, proposed by Windig and Guilment (*DOI: 10.1021/ac00014a016*). If `oncols=True`, this function estimates  $\mathbb{S}$  with SIMPLISMA, then calculates  $\mathbb{C} = \mathbb{D}\mathbb{S}^+$ . If `oncols=False`, this function estimates  $\mathbb{C}$  with SIMPLISMA, then calculates  $\mathbb{S} = \mathbb{C}^+\mathbb{D}$ . `f` defines the percentage of allowed noise.

#### Parameters

- `D`: *2darray*  
Input data, of dimensions  $m \times n$
- `nc`: *int*  
Number of components to be found. This determines the final size of the  $\mathbb{C}$  and  $\mathbb{S}$  matrices.
- `f`: *float*  
Percentage of allowed noise.
- `oncols`: *bool*  
If `True`, SIMPLISMA estimates the  $\mathbb{S}$  matrix, otherwise estimates  $\mathbb{C}$ .

#### Returns

- `C`: *2darray*  
Estimation of the  $\mathbb{C}$  matrix, of dimensions  $m \times \text{nc}$ .
  - `S`: *2darray*  
Estimation of the  $\mathbb{S}$  matrix, of dimensions  $\text{nc} \times n$ .
-

### 3.2.9 `processing.baseline_correction(ppm, data, basl_file='spectrum.basl', winlim=None)`

Interactively corrects the baseline of a given spectrum and saves the parameters in a file. The program starts with an interface to partition the spectrum in windows to correct separately. Then, for each window, an interactive panel opens to allow the user to compute the baseline.

#### Parameters:

- `ppm`: *1darray*  
PPM scale of the spectrum
  - `data`: *1darray*  
The spectrum of which to adjust the baseline
  - `basl_file`: *str*  
Name for the baseline parameters file
  - `winlim`: *list or str or None*  
List of the breakpoints for the window. If it is `str`, it points to a file to be read with `np.loadtxt`. If it is `None`, the partitioning is done interactively.
-

### 3.2.10 `processing.calc_nc(data, s_n)`

Calculates the optimal number of components, given the standard deviation of the noise. The threshold value is calculated as stated in Theorem 1 of reference: <https://arxiv.org/abs/1710.09787v2>

**Parameters:**

- `data`: *2darray*  
Input data
- `s_n`: *float*  
Noise standard deviation

**Returns:**

- `n_c`: *int*  
Number of components
-

### 3.2.11 `processing.calibration(ppmscale, S)`

Scroll the ppm scale of spectrum to make calibration. The interface offers two guidelines: the red one, labelled 'reference signal' remains fixed, whereas the green one ('calibration value') moves with the ppm scale.

The ideal calibration procedure consists in placing the red line on the signal you want to use as reference, and the green line on the ppm value that the reference signal must assume in the calibrated spectrum. Then, scroll with the mouse until the two lines are superimposed.

#### Parameters

- `ppmscale`: *1darray*  
The ppm scale to be calibrated
- `S`: *1darray*  
The spectrum to calibrate

#### Returns

- `offset`: *float*  
Difference between original scale and new scale. This must be summed up to the original ppm scale to calibrate the spectrum.
-

### 3.2.12 processing.convdta(data, grpdly=0, scaling=1)

Removes the digital filtering to obtain a spectrum similar to the command CONVDTA performed by TopSpin. However, they will differ a little bit because of the digitization. These differences are not invisible to human's eye.

#### Parameters:

- data: *ndarray*  
FID with digital filter
- grpdly: *int*  
Number of points that the digital filter consists of. Key *\$GRPDLY* in *acqus* file
- scaling: *float*  
Scaling factor of the resulting FID. Needed to match TopSpin's intensities.

#### Returns:

- data\_in: *ndarray*  
FID without the digital filter. It will have *grpdly* points less than *data*.
-

### 3.2.13 processing.em(data, lb, sw)

Exponential apodization.

Being the FID an array of  $N$  points in its last dimension, and taken  $x = \text{np.arange}(N)/N$ :

$$\text{apod} = \exp\left[-\pi \frac{\text{lb}}{2\text{sw}} x\right]$$

#### Parameters

- data : *ndarray*  
FID to be apodized
- lb : *float*  
Lorentzian broadening, in Hz.  $\text{lb} > 0$
- sw : *float*  
Spectral width in Hz, used for normalization.

#### Returns

- apod \* data : *ndarray*  
Apodized FID.
-



### 3.2.14 `processing.fp(data, wf=None, zf=None, fcor=0.5, tdeff=0)`

Performs the full processing of a 1D NMR FID.

#### Parameters

- data: *1darray*  
Data to be processed.
- wf: *dict*  
{'mode': function to be used, 'parameters': see window functions documentation}
- zf: *int*  
final size of spectrum
- fcor: *float*  
Weighting factor for the first point of the FID.
- tdeff: *int*  
Number of FID points to be employed for the processing. `tdeff = 0` means whole FID.

#### Returns

- data: *ndarray*  
Processed spectrum.
-

### 3.2.15 `processing.ft(data, alt=False, fcor=0.5, Numpy=True)`

Fourier transform in NMR sense, i.e. with positive exponential. This means to perform IFT reverting the  $1/N$  scaling.

#### Parameters

- `data` : *ndarray*  
FID to be Fourier-transformed.
- `alt`: *bool*  
Negates the sign of the odd points, then takes the complex conjugate. Required for States-TPPI processing.
- `fcor`: *float*  
Weighting factor for FID 1-st point. Default value (0.5) prevents baseline offset
- `Numpy`: *bool*  
If `True` (\*\*STRONGLY ADVISED\*\*) performs the FT using the FFT algorithm encoded in Numpy. Otherwise, performs it manually using the definition of discrete FT.

#### Returns

- `dataft` : *ndarray*  
Transformed FID.
-

### 3.2.16 processing.gm(data, lb, gb, sw, gc=0)

Gaussian apodization.

Being the FID an array of  $N$  points in its last dimension, and taken  $x = \text{np.arange}(N)$ :

$$\text{apod} = \exp[a - b^2], \quad a = \pi \frac{\text{lb}}{\text{sw}} x, \quad b = 0.6\pi \frac{\text{gb}}{\text{sw}} (\text{gc}(N - 1) - x)$$

#### Parameters

- data : *ndarray*  
FID to be apodized
- lb : *float*  
Lorentzian broadening, in Hz.  $\text{lb} < 0$
- gb : *float*  
Gaussian broadening, in Hz.  $\text{gb} > 0$
- sw : *float*  
Spectral width in Hz, used for normalization.
- gc: *float*  
Gaussian center, relatively to the FID length:  $0 \leq \text{gc} \leq 1$

#### Returns

- apod \* data : *ndarray*  
Apodized FID.
-

### 3.2.17 processing.gmb(data, lb, gb, sw)

Gaussian apodization, Bruker-like. It does not work very well.

Being the FID an array of  $N$  points in its last dimension, and taken  $t = \text{np.arange}(N)/\text{sw}$ :

$$\text{apod} = \exp[\alpha t - (\beta t)^2], \quad \alpha = \pi \text{lb}, \quad \beta = \frac{-\alpha N}{2 \text{gb sw}}$$

#### Parameters

- data : *ndarray*  
FID to be apodized
- lb : *float*  
Lorentzian broadening, in Hz.  $\text{lb} < 0$
- gb : *float*  
Gaussian broadening, in Hz.  $\text{gb} > 0$
- sw : *float*  
Spectral width in Hz, used for normalization.
- gc: *float*  
Gaussian center, relatively to the FID length:  $0 \leq \text{gc} \leq 1$

#### Returns

- apod \* data : *ndarray*  
Apodized FID.
-

### 3.2.18 `processing.ift(data, alt=False, fcor=0.5, Numpy=True)`

Inverse Fourier transform in NMR sense, i.e. with negative exponential. This means to perform FT multiplying by  $N$  afterwards.

#### Parameters

- `data` : *ndarray*  
FID to be inverse Fourier-transformed.
- `alt`: *bool*  
Negates the sign of the odd points, then takes the complex conjugate. Required for States-TPPI processing.
- `fcor`: *float*  
Weighting factor for FID 1-st point. Default value (0.5) prevents baseline offset
- `Numpy`: *bool*  
If `True` (**\*\*\*STRONGLY ADVISED\*\*\***) performs the FT using the FFT algorithm encoded in Numpy. Otherwise, performs it manually using the definition of discrete FT.

#### Returns

- `dataft` : *ndarray*  
Inverse transformed FID.
-

### 3.2.19 `processing.integral(fx, x=None, lims=None)`

Calculates the primitive of `fx`. If `fx` is a multidimensional array, the integrals are computed along the last dimension.

#### Parameters:

- `fx`: *ndarray*  
Function (array) to integrate
- `x`: *1darray or None*  
Independent variable. Determines the integration step. If `None`, it is set as the point scale
- `lims`: *tuple or None*  
Integration range. If `None`, the whole function is integrated

#### Returns:

- `Fx`: *ndarray*  
Integrated function.
-

### 3.2.20 `processing.integral_2D(ppm_f1, t_f1, SFO1, ppm_f2, t_f2, SFO2, u_1=None, fwhm_1=200, utol_1=0.5, u_2=None, fwhm_2=200, utol_2=0.5, plot_result=False)`

Calculate the integral of a 2D peak. The idea is to extract the traces correspondent to the peak center and fit them with a gaussian function in each dimension. Then, once got the intensity of each of the two gaussians, multiply them together in order to obtain the 2D integral. This procedure should be equivalent to what CARA does.

#### Parameters:

- `ppm_f1`: *1darray*  
PPM scale of the indirect dimension
- `t_f1`: *1darray*  
Trace of the indirect dimension, real part
- `SFO1`: *float*  
Larmor frequency of the nucleus in the indirect dimension
- `ppm_f2`: *1darray*  
PPM scale of the direct dimension
- `t_f2`: *1darray*  
Trace of the direct dimension, real part
- `SFO2`: *float*  
Larmor frequency of the nucleus in the direct dimension
- `u_1`: *float*  
Chemical shift in F1 /ppm. Defaults to the center of the scale
- `fwhm_1`: *float*  
Starting FWHM /Hz in the indirect dimension
- `utol_1`: *float*  
Allowed tolerance for `u_1` during the fit. (`u_1-utol_1`, `u_1+utol_1`)
- `u_2`: *float*  
Chemical shift in F2 /ppm. Defaults to the center of the scale
- `fwhm_2`: *float*  
Starting FWHM /Hz in the direct dimension
- `utol_2`: *float*  
Allowed tolerance for `u_2` during the fit. (`u_2-utol_2`, `u_2+utol_2`)
- `plot_result`: *bool*  
True to show how the program fitted the traces.

#### Returns:

- `I_tot`: *float*  
Computed integral.
-

### 3.2.21 `processing.interactive_basl_windows(ppm, data)`

Allows for interactive partitioning of a spectrum in windows. Double left click to add a bar, double right click to remove it. Returns the location of the red bars as a list.

#### Parameters:

- ppm: *1darray*  
PPM scale of the spectrum
- data: *1darray*  
Spectrum to be partitioned

#### Returns:

- coord: *list*  
List containing the coordinates of the windows, plus `ppm[0]` and `ppm[-1]`
-



### 3.2.22 `processing.interactive_fp(fid0, acqu, procs)`

Perform the processing of a 1D NMR spectrum interactively. The GUI offers the opportunity to test different window functions, as well as different `tdeff` values and final sizes. The active parameters appear as blue text.

#### Parameters:

- `fid0`: *1darray*  
FID to process
- `acqu`: *dict*  
Dictionary of acquisition parameters
- `procs`: *dict*  
Dictionary of processing parameters

#### Returns:

- `pdata`: *1darray*  
Processed spectrum
  - `procs`: *dict*  
Updated dictionary of processing parameters
-

### 3.2.23 processing.interactive\_phase\_1D(ppmscale, S)

Allows for interactive phase adjustment of 1D NMR spectra. Employs `processing.ps` for the actual phase correction. Press the **z** key on the keyboard to toggle the automatic adjustment of vertical scale on or off.

#### Parameters

- `ppmscale`: *1darray*  
PPM scale of the spectrum
- `S`: *1darray*  
The spectrum to be phased

#### Returns

- `phased_data`: *1darray*  
Phased spectrum.
-

### 3.2.24 `processing.interactive_phase_2D(ppm_f1, ppm_f2, S)`

Interactively adjust the phases of a 2D spectrum. `S` must be hypercomplex, therefore must be passed before to unpack it into the 4 real files. The phase correction is done using `processing.ps` as follows:

1. `S = processing.ps(S, p0=p0_f2, p1=p1_f2, pivot=pivot_f2)`
2. Transpose: normal if `FnMODE='QF'`, hypercomplex otherwise
3. `S = phase(S, p0=p0_f1, p1=p1_f1, pivot=pivot_f1)`
4. Transpose back

#### Parameters

- `ppm_f1`: *1darray*  
Indirect dimension ppm scale
- `ppm_f1`: *1darray*  
Direct dimension ppm scale
- `S`: *2darray*  
Hypercomplex spectrum

#### Returns

- `S`: *2darray*  
Phased spectrum
-

### 3.2.25 `processing.interactive_qfil(ppm, data_in)`

Interactive function to design a gaussian filter with the aim of suppressing signals in the spectrum. You can adjust position and width of the filter scrolling with the mouse.

#### Parameters:

- ppm: *1darray*  
Scale on which the filter will be built
- data\_in: *1darray*  
Spectrum on which to apply the filter.

#### Returns:

- u: *float*  
Position of the gaussian filter
  - s: *float*  
Width of the gaussian filter (Standard deviation)
-

### 3.2.26 `processing.interactive_xfb(fid0, acqu, procs, lvl0=0.1, show_cnt=True)`

Perform the processing of a 2D NMR spectrum interactively. The GUI offers the opportunity to test different window functions, as well as different `tdeff` values and final sizes. The active parameters appear as blue text. When changing the parameters, give it some time to compute. The figure panel is quite heavy.

#### Parameters:

- `fid0`: *2darray*  
FID to process
- `acqu`: *dict*  
Dictionary of acquisition parameters
- `procs`: *dict*  
Dictionary of processing parameters
- `lvl0`: *float*  
Starting level of the contours
- `show_cnt`: *bool*  
Choose if to display data using contours (**True**) or heatmap (**False**)

#### Returns:

- `pdata`: *2darray*  
Processed spectrum
  - `procs`: *dict*  
Updated dictionary of processing parameters
-

### 3.2.27 processing.inv\_fp(data, wf=None, size=None, fcor=0.5)

Performs the full inverse processing of a 1D NMR spectrum (`data`). Required parameters are:

#### Parameters

- `data`: *1darray*  
Input data
- `wf`: *dict*  
{'mode': function to be used, 'parameters': different from each function}
- `size`: *int*  
initial size of the FID
- `fcor`: *float*  
weighting factor for the FID first point

#### Returns

- `data`: *1darray*  
Processed data
-

### 3.2.28 `processing.inv_xfb(data, wf=[None, None], size=[None, None], fcor=[0.5, 0.5], FnMODE='States-TPPI')`

Performs the full processing of a 2D NMR FID (`data`). Required parameters are:

#### Parameters

- `data`: *2darray*  
Input data
- `wf`: *list of dict*  
list of two entries [`F1`, `F2`]. Each entry is a dictionary of window functions
- `zf`: *list*  
list of two entries [`zf F1`, `zf F2`]
- `fcor`: *list*  
first fid point weighting factor [`F1`, `F2`]
- `u`: *bool*  
If `True`, unpacks the hypercomplex spectrum and returns the 4 real files, using `processing.unpack_2D`
- `tdeff`: *list of int*  
number of points of the FID to be used for the processing, [`F1`, `F2`]

#### Returns

- `data`: *2darray*  
Processed data
-

### 3.2.29 processing.iterCadzow(data, n, nc, itermax=100, f=0.005, print\_head=True, print\_time=True)

This functions performs Cadzow denoising on `data`, which is a 1D array of  $N$  points, in an iterative manner. The algorithm works as follows:

1. Transform `data` in a Hankel matrix  $\mathbb{H}$  of dimensions  $(N - n, n)$
2. Make SVD on  $\mathbb{H} = \mathbb{U}\mathbb{S}\mathbb{V}^\dagger$
3. Keep only the first `nc` singular values, and put all the rest to 0 ( $\mathbb{S} \rightarrow \mathbb{S}'$ )
4. Rebuild  $\mathbb{H}' = \mathbb{U}\mathbb{S}'\mathbb{V}^\dagger$
5. Average the antidiagonals to rebuild the Hankel-type structure, then make 1D array
6. Check arrest criterion: if it is not reached, go to step 1, otherwise exit from the cycle and return the processed data.

The arrest criterion is on the array of singular values  $S$ , which is the main diagonal of the matrix  $\mathbb{S}$ . At step  $k$  and Python indexing system:

$$\left| \frac{S^{(k-1)}[\text{nc} - 1]}{S^{(k-1)}[0]} - \frac{S^{(k)}[\text{nc} - 1]}{S^{(k)}[0]} \right| < f \frac{S^{(0)}[\text{nc} - 1]}{S^{(0)}[0]}$$

#### Parameters

- `data`: *1darray*  
Input data
- `n`: *int*  
Number of columns of the Hankel matrix.
- `nc`: *int*  
Number of singular values to keep.
- `itermax`: *int*  
Maximum number of iterations allowed.
- `f`: *float*  
Factor for the arrest criterion.
- `print_head`: *bool*  
Set it to `True` to display the fancy heading.
- `print_time`: *bool*  
Set it to `True` to display the time spent.

#### Returns

- `datap`: *1darray*  
Denoised data
-



### 3.2.30 processing.load\_baseline(filename, ppm, data)

Read the baseline parameters from a file and builds the baseline itself.

#### Parameters:

- filename: *str*  
Location of the baseline file
- ppm: *1darray*  
PPM scale of the spectrum
- data: *1darray*  
Spectrum of which to correct the baseline

#### Returns:

- baseline: *1darray*  
Computed baseline
-

### 3.2.31 processing.make\_polynomion\_baseline(ppm, data, limits)

Interactive baseline correction with 4th degree polynomion.

#### Parameters:

- ppm: *1darray*  
PPM scale of the spectrum
- data: *1darray*  
spectrum
- limits: *tuple*  
Window limits (left, right).

#### Returns:

- mode: *str*  
Baseline correction mode: 'polynomion' as default, 'spline' if you press the button
  - C\_f: *1darray or str*  
Baseline polynomion coefficients, or 'callintsmooth' if you press the spline button
-

### 3.2.32 processing.make\_\_scale(size, dw, rev=True)

Computes the frequency scale of the NMR spectrum, given the number of points and the employed dwell time (the REAL one, not the TopSpin one!). `rev=True` is required for the correct frequency arrangement in the NMR sense.

#### Parameters

- size: *int*  
Number of points of the frequency scale
- dw : *float*  
Time spacing in the time dimension
- rev: *bool*  
Reverses the scale

#### Returns

- fqscale: *1darray*  
The computed frequency scale.
-

### 3.2.33 `processing.pknl(data, grpdly=0, onfid=False)`

Compensate for the Bruker group delay at the beginning of FID through a first-order phase correction of

$$\phi^{(1)} = 360 * \text{GRPDLY}$$

This should be applied after apodization and zero-filling.

#### Parameters:

- `data`: *ndarray*  
Input data. Be sure it is complex!
- `grpdly`: *int*  
Number of points that make the group delay.
- `onfid`: *bool*  
If it is `True`, performs FT before to apply the phase correction, and IFT after.

#### Returns:

- `datap`: *ndarray*  
Corrected data
-

### 3.2.34 `processing.ps(data, ppmscale=None, p0=None, p1=None, pivot=None, interactive=False)`

Applies phase correction on the last dimension of data. The pivot is set at the center of the spectrum by default. Missing parameters will be inserted interactively.

Being `data` a 1D array of  $N$  points, as well as `ppmscale`, the following parameters are defined:

$$p_V = (\text{index of pivot on ppmscale})/N$$

$$x = \text{np.arange}(N)/N - p_V$$

Basically,  $p_V$  and  $x$  are `pivot` and `ppmscale`, normalized in order to fit a scale that ranges from 0 to 1. This is done with the aim to 'standardize' the behaviour of the function, making it nucleus-independent.

The following phase-correction function is applied:

$$\Phi = \exp[i(p_0 + p_1 x)]$$

If `data` is a 2D array, the function  $\Phi$  is applied row-wise.

#### Parameters:

- `data`: *ndarray*  
Input data
- `ppmscale`: *1darray or None*  
PPM scale of the spectrum. Required for `pivot` and `interactive` phase correction
- `p0`: *float*  
Zero-order phase correction angle /degrees
- `p1`: *float*  
First-order phase correction angle /degrees
- `pivot`: *float or None*.  
First-order phase correction pivot /ppm. If `None`, it is set at the center of the spectrum.
- `interactive`: *bool*  
If `True`, all the parameters will be ignored and the interactive phase correction panel will be opened.

#### Returns:

- `datap`: *ndarray*  
Phased data
  - `final_values`: *tuple*  
Employed values of the phase correction. (`p0`, `p1`, `pivot`)
-

### 3.2.35 processing.qfil(ppm, data, u, s)

Suppress signals in the spectrum using a gaussian filter.

#### Parameters:

- ppm: *1darray*  
Scale on which to build the filter
- data: *ndarray*  
Data to be processed. The filter is applied on the last dimension
- u: *float*  
Position of the filter
- s: *float*  
Width of the filter (standard deviation)

#### Returns:

- pdata: *ndarray*  
Filtered data
-

### 3.2.36 processing.qpol(fid)

Fits the FID with a 4-th degree polynomion, then subtracts it from the original FID. The real and imaginary channels are treated separately.

#### Parameters

- `fid` : *ndarray*  
Self-explanatory.

#### Returns

- `fid` : *ndarray*  
Processed FID.
-

### 3.2.37 processing.qsin(data, ssb)

Sine-squared apodization.

Being `data` an array of  $N$  points in its last dimension, and taken  $x = \text{np.arange}(N)/N$ , if `ssb` = 0 or `ssb` = 1:

$$\text{apod} = \sin^2\left(\pi x\right)$$

else if `ssb`  $\geq 2$ :

$$\text{apod} = \sin^2\left(\pi \frac{1}{\text{ssb}} + \pi \left(1 - \frac{1}{\text{ssb}}\right)x\right)$$

#### Parameters

- `data` : *ndarray*  
FID to be apodized.
- `ssb` : *int*  
Shifting parameter for the sine bell.
  - `ssb` = 0 : from 0 to  $\pi$
  - `ssb` = 1 : same as `ssb` = 0
  - `ssb` = 2 : from  $\pi/2$  to  $\pi$
  - `ssb` = 3 : from  $\pi/3$  to  $\pi$

#### Returns

- `apod * data` : *ndarray*  
Apodized FID.
-



### 3.2.38 processing.quad(fid)

Subtracts from the FID the arithmetic mean of its last quarter. The real and imaginary channels are treated separately.

#### Parameters

- *fid* : *ndarray*  
Self-explanatory.

#### Returns

- *fid* : *ndarray*  
Processed FID.
-

### 3.2.39 `processing.repack_2D(rr, ir, ri, ii)`

Reconstruct hypercomplex 2D NMR data given the 4 real arrays. See `processing.unpack_2D` for details.

#### Returns

- data
-

### 3.2.40 `processing.rev(data)`

Reverses the last dimension of `data`.

#### Parameters

- `data : ndarray`  
matrix to be reverted

#### Returns

- `data[...,::-1] : ndarray`  
Self-explanatory
-

### 3.2.41 `processing.sin(data, ssb)`

Sine apodization.

Being `data` an array of  $N$  points in its last dimension, and taken  $x = \text{np.arange}(N)/N$ , if `ssb` = 0 or `ssb` = 1:

$$\text{apod} = \sin\left(\pi x\right)$$

else if `ssb`  $\geq 2$ :

$$\text{apod} = \sin\left(\pi \frac{1}{\text{ssb}} + \pi \left(1 - \frac{1}{\text{ssb}}\right)x\right)$$

#### Parameters

- `data` : *ndarray*  
FID to be apodized.
- `ssb` : *int*  
Shifting parameter for the sine bell.
  - `ssb` = 0 : from 0 to  $\pi$
  - `ssb` = 1 : same as `ssb` = 0
  - `ssb` = 2 : from  $\pi/2$  to  $\pi$
  - `ssb` = 3 : from  $\pi/3$  to  $\pi$

#### Returns

- `apod * data` : *ndarray*  
Apodized FID.
-

### 3.2.42 `processing.split_echo_train(datao, n, n_echoes, i_p=0)`

`datao` is a 1D or 2D dataset whose direct dimension has been acquired with the CPMG sequence, i.e. the FID is made of echoes separated one from each other by `n` points. The first good point of the FID is `i_p`. This function separates the first `n_echoes` echoes and stores them in a 2D array of shape `(n_echoes, datao.shape[0], n/2)`.

#### Parameters

- `datao` : *ndarray*  
1D or 2D CPMG FID
- `n` : *int*  
Number of points that separate an echo from the next one
- `n_echoes` : *int*  
Number of echoes to add together
- `i_p` : *int*  
First point offset.

#### Returns

- `data_p` : *ndarray*  
Processed FID.
-

### 3.2.43 processing.stack\_MCR(input\_data, H=True)

Performs matrix augmentation converting `input_data` from dimensions  $(X, Y, Z)$  to  $(Y, X * Z)$  if `H=True`, or  $(X * Y, Z)$  if `H=False`.

#### Parameters

- `input_data`: *3darray*  
Contains the spectra to be stacked together. The index that runs on the datasets must be the first one.
- `H`: *bool*  
`True` for horizontal stacking, `False` for vertical stacking.

#### Returns

- `data`: *2darray*  
Augmented data matrix.
-

### 3.2.44 `processing.sum_echo_train(datao, n, n_echoes, i_p=0)`

`datao` is a 1D or 2D dataset whose direct dimension has been acquired with the CPMG sequence, i.e. the FID is made of echoes separated one from each other by `n` points. The first good point of the FID is `i_p`. This function sums up the first `n_echoes` echoes and returns the resulting FID.

#### Parameters

- `datao` : *ndarray*  
1D or 2D CPMG FID
- `n` : *int*  
Number of points that separate an echo from the next one
- `n_echoes` : *int*  
Number of echoes to add together
- `i_p` : *int, optional*  
First point offset.

#### Returns

- `data_p` : *ndarray*  
Processed FID.
-

### 3.2.45 `processing.tabula_rasa(data, lvl=0.05, cmap=cm.Blues_r)`

This function is to be used in SIFT algorithm. Allows interactive selection using a Lasso widget of the region of the spectrum which contain signal. Returns a masking matrix, of the same shape as data, whose entries are 1 inside the selection and 0 outside.

#### Parameters

- `data` : *2darray*  
The data to be plotted
- `lvl` : *float*  
Level of the contours, expressed as fraction of the maximum intensity
- `cmap` : *matplotlib.cm Object*  
Color of the contours

#### Returns

- `mask`: *2darray*  
Matrix that contains 1 inside the selection and 0 outside.
-



### 3.2.46 processing.td\_eff(data, tdeff)

Uses only the first `tdeff` points of data. It is applied before any other processing. The length of the list `tdeff` must match the dimension of `data`, if `data` is multidimensional.

#### Parameters

- data: *ndarray*  
The FID to be trimmed.
- tdeff: *int or list*  
The number of points to be used. If `data` is 1D, `tdeff` can be passed as integer, otherwise it has to be: `[F1, F2, ..., Fn]`. A 0 entry in the list means to not trim the corresponding dimension.

#### Returns

- data: *ndarray*  
Trimmed data according to `tdeff`.
-

### 3.2.47 processing.tp\_hyper(data)

Computes the hypercomplex transpose of data. Needed for the processing of data acquired in a phase-sensitive manner in the indirect dimension.

To explain how a hypercomplex transposition works, we will focus on the simpler example of a  $4 \times 6$  matrix,  $\mathbb{A}$ , with complex coefficients  $a_{i,j} = x_{i,j} + iy_{i,j}$ .

$$\mathbb{A} = \begin{pmatrix} x_{1,1} + iy_{1,1} & x_{1,2} + iy_{1,2} & x_{1,3} + iy_{1,3} & x_{1,4} + iy_{1,4} \\ x_{2,1} + iy_{2,1} & x_{2,2} + iy_{2,2} & x_{2,3} + iy_{2,3} & x_{2,4} + iy_{2,4} \\ x_{3,1} + iy_{3,1} & x_{3,2} + iy_{3,2} & x_{3,3} + iy_{3,3} & x_{3,4} + iy_{3,4} \\ x_{4,1} + iy_{4,1} & x_{4,2} + iy_{4,2} & x_{4,3} + iy_{4,3} & x_{4,4} + iy_{4,4} \\ x_{5,1} + iy_{5,1} & x_{5,2} + iy_{5,2} & x_{5,3} + iy_{5,3} & x_{5,4} + iy_{5,4} \\ x_{6,1} + iy_{6,1} & x_{6,2} + iy_{6,2} & x_{6,3} + iy_{6,3} & x_{6,4} + iy_{6,4} \end{pmatrix}$$

The hypercomplex tranpose of the matrix  $\mathbb{A}$ , here referred as  $\mathbb{B}$ , is:

$$\mathbb{B} = \begin{pmatrix} x_{1,1} + ix_{2,1} & x_{3,1} + ix_{4,1} & x_{5,1} + ix_{6,1} \\ y_{1,1} + iy_{2,1} & y_{3,1} + iy_{4,1} & y_{5,1} + iy_{6,1} \\ x_{1,2} + ix_{2,2} & x_{3,2} + ix_{4,2} & x_{5,2} + ix_{6,2} \\ y_{1,2} + iy_{2,2} & y_{3,2} + iy_{4,2} & y_{5,2} + iy_{6,2} \\ x_{1,3} + ix_{2,3} & x_{3,3} + ix_{4,3} & x_{5,3} + ix_{6,3} \\ y_{1,3} + iy_{2,3} & y_{3,3} + iy_{4,3} & y_{5,3} + iy_{6,3} \\ x_{1,4} + ix_{2,4} & x_{3,4} + ix_{4,4} & x_{5,4} + ix_{6,4} \\ y_{1,4} + iy_{2,4} & y_{3,4} + iy_{4,4} & y_{5,4} + iy_{6,4} \end{pmatrix}$$

#### Parameters

- data : *2darray*  
Complex data matrix.

#### Returns

- datatp : *2darray*  
Hypercomplex transpose of data.

### 3.2.48 processing.unpack\_2D(data)

Separates fully processed 2D NMR data into 4 distinct ser files:

```
rr = Re{data}[:, 2]
ir = Im{data}[:, 2]
ri = Re{data}[1 :: 2]
ii = Im{data}[1 :: 2]
```

#### Parameters

- data : *2darray*  
Complex data matrix.

#### Returns

- rr, ir, ri, ii: *2darray, 2darray, 2darray, 2darray*  
See above.
-

### 3.2.49 `processing.write_basl_info(f, limits, mode, data)`

Writes the baseline parameters of a certain window in a file.

#### Parameters:

- `f`: *TextIO object*  
File where to write the parameters
  - `limits`: *tuple*  
Limits of the spectral window. (`left`, `right`)
  - `mode`: *str*  
Baseline correction mode: `'polynomion'` or `'spline'`
  - `data`: *float or 1darray*  
It can be either the spline smoothing factor or the polynomion coefficients
-

### 3.2.50 `processing.xfb(data, wf=[None, None], zf=[None, None], fcor=[0.5,0.5], tdeff=[0,0], u=True, FnMODE='States-TPPI')`

Performs the full processing of a 2D NMR FID.

#### Parameters

- `data`: *2darray*  
FID data matrix
- `wf`: *list*  
List of two entries [F1, F2]. Each entry is a dictionary: {'mode': function to be used, 'parameters': different from each function}
- `zf`: *list*  
List of two entries [zf F1, zf F2]
- `fcor`: *list*  
Weighting factor for the first point of the FID. [F1, F2]
- `tdeff`: *list*  
Number of FID points to be employed for the processing. `tdeff = [0,0]` means whole FID. [F1, F2]
- `u`: *bool*  
if `True`, unpacks the hypercomplex spectrum and returns the 4 files.

#### Returns

- `data`: *2darray*  
Processed data.
-

### 3.2.51 processing.zf(data, size)

Zero-filling of data up to `size`.

#### Parameters

- `data` : *ndarray*  
FID / Spectrum to be zero-filled
- `size` : *int*  
Final size of the FID / Spectrum

#### Returns

- `datazf` : *ndarray*  
Zero-filled FID / Spectrum
-

## 3.3 FIGURES package

This package contains a series of functions to make plots of various nature.

---

### 3.3.1 `figures.ax1D(ax, ppm, data, norm=False, xlims=None, ylims=None, c='b', lw=0.5, X_label='$\delta$, $ F1 /ppm', Y_label='Intensity /a.u.', n_xticks=10, n_yticks=10, label=None, fontsize=10)`

Makes the figure of a 1D NMR spectrum, placing it in a given figure panel. This allows the making of modular figures.

The plot can be customized in a very flexible manner by setting the function keywords properly.

#### Parameters

- `ax`: *matplotlib.subplot Object*  
panel where to put the figure
- `ppm`: *1darray*  
ppm scale of the spectrum
- `data`: *1darray*  
spectrum to be plotted
- `norm`: *bool*  
if `True`, normalizes the intensity to 1.
- `xlims`: *list or tuple*  
Limits for the x-axis. If `None`, the whole scale is used.
- `ylims`: *list or tuple*  
Limits for the y-axis. If `None`, the whole scale is used.
- `c`: *str*  
Colour of the line.
- `lw`: *float*  
linewidth
- `X_label`: *str*  
text of the x-axis label;
- `Y_label`: *str*  
text of the y-axis label;
- `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
- `n_yticks`: *int*  
Number of numbered ticks on the x-axis of the figure
- `label`: *str*  
label to be put in the legend.
- `fontsize`: *float*  
Biggest font size in the figure.

**Returns:**

- line: *Line2D Object*  
Line object returned by `plt.plot`.
-



**3.3.2** `figures.ax2D(ax, ppm_f2, ppm_f1, datax, xlims=None, ylims=None, cmap=None, c_fac=1.4, lvl=0.1, lw=0.5, X_label='$\delta\,$ F2 /ppm', Y_label='$\delta\,$ F1 /ppm', title=None, n_xticks=10, n_yticks=10), fontsize=10`

Makes a 2D contour plot like the one in `figures.figure2D`, but in a specified panel. Allows for the buildup of modular figures. The contours are drawn according to the formula:

$$c_l = \text{contour\_start} * \left( \text{contour\_factor} ** \text{np.arange}(\text{contour\_num}) \right)$$

where `contour_start = np.max(data) * lvl`, `contour_num = 16` and `contour_factor = c_fac`. Increasing the value of `c_fac` will decrease the number of contour lines, whereas decreasing the value of `c_fac` will increase the number of contour lines.

### Parameters

- `ax`: *matplotlib.subplot Object*  
panel where to put the figure
- `ppm_f2`: *1darray*  
ppm scale of the direct dimension
- `ppm_f1`: *1darray*  
ppm scale of the indirect dimension
- `datax`: *2darray*  
the 2D NMR spectrum to be plotted
- `xlims`: *tuple*  
limits for the x-axis (`left`, `right`). If `None`, the whole scale is used.
- `ylims`: *tuple*  
limits for the y-axis (`left`, `right`). If `None`, the whole scale is used.
- `cmap`: *matplotlib.cm Object*  
Colour for the contour
- `c_fac`: *float*  
Contour factor parameter
- `lvl`: *float*  
height with respect to maximum at which the contour are computed
- `X_label`: *str*  
text of the x-axis label;
- `Y_label`: *str*  
text of the y-axis label;
- `lw`: *float*  
linewidth of the contours
- `title`: *str*  
Figure title.

- `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `n_yticks`: *int*  
Number of numbered ticks on the y-axis of the figure
  - `fontsize`: *float*  
Biggest font size in the figure.
-

### 3.3.3 `figures.ax_heatmap(ax, data, zlim='auto', z_sym=True, cmap=None, xscale=None, yscale=None, rev=(False, False), n_xticks=10, n_yticks=10, n_zticks=10, fontsize=10)`

Computes a heatmap of data on the given `ax`.

#### Parameters:

- `ax`: *matplotlib.Subplot object*  
Panel where to draw the heatmap
- `data`: *2darray*  
Input data
- `zlim`: *tuple or 'auto' or 'abs'*  
Vertical limits of the heatmap, that determine the extent of the colorbar. 'auto' means `(min(data), max(data))`, 'abs' means `(min(|data|), max(|data|))`.
- `z_sym`: *bool*  
`True` to symmetrize the vertical scale around 0.
- `cmap`: *matplotlib.cm object*  
Colormap of the heatmap.
- `xscale`: *1darray or None*  
x-scale. `None` means `np.arange(data.shape[1])`
- `yscale`: *1darray or None*  
y-scale. `None` means `np.arange(data.shape[0])`
- `rev`: *tuple of bool*  
Reverse scale. Format: `(x, y)`.
- `n_xticks`: *int*  
Number of ticks of the x axis
- `n_yticks`: *int*  
Number of ticks of the y axis
- `n_zticks`: *int*  
Number of ticks of the color bar
- `fontsize`: *float*  
Biggest font size to apply to the figure.

#### Returns:

- `im`: *matplotlib.AxesImage*  
The heatmap
  - `cax`: *matplotlib.Subplot object*  
figure panel where the colorbar is drawn
-

### 3.3.4 `figures.dotmd(ppmscale, S, labels=None, n_xticks=10)`

Interactive display of multiple 1D spectra.

#### Parameters

- `ppmscale`: *1darray or list*  
ppm scale of the spectra. If only one scale is supplied, all the spectra are plotted using the same scale. Otherwise, each spectrum is plotted using its scale. There is a 1:1 correspondance between `ppmscale` and `S`.
  - `S`: *list*  
spectra to be plotted
  - `labels`: *list*  
labels to be put in the legend.
  - `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
-

### 3.3.5 `figures.dotmd_2D(ppm_f1, ppm_f2, S, labels=None, name='dotmd_2D', X_label='$\delta$, $ F2 /ppm', Y_label='$\delta$, $ F1 /ppm', n_xticks=10, n_yticks=10, Neg=True)`

Interactive display of multiple 2D spectra. They have to share the same scales.

- `ppm_f1`: *1darray*  
ppm scale of the indirect dimension. If only one scale is supplied, all the spectra are plotted using the same scale. Otherwise, each spectrum is plotted using its scale. There is a 1:1 correspondance between `ppm_f1` and `S`.
  - `ppm_f2`: *1darray*  
ppm scale of the direct dimension. If only one scale is supplied, all the spectra are plotted using the same scale. Otherwise, each spectrum is plotted using its scale. There is a 1:1 correspondance between `ppm_f2` and `S`.
  - `S`: *list*  
spectra to be plotted
  - `labels`: *list*  
labels to be put in the legend.
  - `name`: *str*  
If you choose to save the figure, this is its filename.
  - `X_label`: *str*  
text of the x-axis label;
  - `Y_label`: *str*  
text of the y-axis label;
  - `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `n_yticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `Neg`: *bool*  
If `True`, show the negative contours.
-

### 3.3.6 `figures.figure1D(ppm, data, norm=False, xlims=None, ylims=None, c='b', lw=0.5, name=None, X_label='$\delta$', $F1 /ppm', Y_label='Intensity /a.u.', n_xticks=10, n_yticks=10, hideylabels=False)`

Makes the figure of a 1D NMR spectrum. The plot can be customized in a very flexible manner by setting the function keywords properly.

The final figure can be either shown or saved.

#### Parameters

- `ppm`: *1darray*  
ppm scale of the spectrum
  - `data`: *1darray*  
spectrum to be plotted
  - `norm`: *bool*  
if `True`, normalizes the intensity to 1.
  - `xlims`: *list or tuple*  
Limits for the x-axis. If `None`, the whole scale is used.
  - `ylims`: *list or tuple*  
Limits for the y-axis. If `None`, they are automatically set.
  - `c`: *str*  
Colour of the line.
  - `lw`: *float*  
linewidth
  - `name`: *str*  
filename of the figure, if it has to be saved;
  - `X_label`: *str*  
text of the x-axis label;
  - `Y_label`: *str*  
text of the y-axis label;
  - `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `n_yticks`: *int*  
Number of numbered ticks on the y-axis of the figure
  - `hideylabels`: *bool*  
if `True`, does not show label and tick labels of the y axis.
-

**3.3.7** `figures.figure1D_multi(ppm0, data0, xlims=None, ylims=None, norm=False, c=None, name=None, X_label='$\delta$ F1 /ppm', Y_label='Intensity /a.u.', n_xticks=10, n_yticks=10, hideylabels=False, labels=None)`

Creates the superimposed plot of a series of 1D NMR spectra.

**Parameters:**

- `ppm0`: *list or 1darray*  
ppm scale of the spectra
  - `data0`: *list*  
List containing the spectra to be plotted
  - `xlims`: *tuple or None*  
Limits for the x-axis. If `None`, the whole scale is used.
  - `ylims`: *tuple or None*  
Limits for the y-axis. If `None`, they are automatically set.
  - `norm`: *False or float or str*  
If it is `False`, it does nothing. If it is float, divides all spectra for that number. If it is `str('#')`, normalizes all the spectra to the #-th spectrum. If it is whatever else string, normalizes all spectra to themselves.
  - `c`: *tuple or None*  
List of the colors to use for the traces. `None` uses the default ones.
  - `name`: *str or None*  
Filename of the figure, if it has to be saved. If it is `None`, the figure is shown instead.
  - `X_label`: *str*  
text of the x-axis label
  - `Y_label`: *str*  
text of the y-axis label
  - `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `n_yticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `hideylabels`: *bool*  
if `True`, does not show label and tick labels of the y axis.
  - `labels`: *list or None or False*  
List of the labels to be shown in the legend. If it is `None`, the default entries are used (i.e., '1, 2, 3,...'). If it is `False`, the legend is not shown.
-

**3.3.8** `figures.figure2D(ppm_f2, ppm_f1, datax, xlims=None, ylims=None, cmap=None, c_fac=1.4, lvl=0.09, name=None, X_label='$\delta$, $F2 /ppm', Y_label='$\delta$, $F1 /ppm', lw=0.5, Negatives=False, cmapneg=None, n_xticks=10, n_yticks=10)`

Creates the contour plot of a 2D NMR spectrum. The contours are drawn according to the formula:

$$cl = \text{contour\_start} * \left( \text{contour\_factor} ** \text{np.arange}(\text{contour\_num}) \right)$$

where `contour_start = np.max(data) * lvl`, `contour_num = 16` and `contour_factor = c_fac`. Increasing the value of `c_fac` will decrease the number of contour lines, whereas decreasing the value of `c_fac` will increase the number of contour lines.

### Parameters

- `ppm_f2`: *1darray*  
ppm scale of the direct dimension
- `ppm_f1`: *1darray*  
ppm scale of the indirect dimension
- `datax`: *2darray*  
the 2D NMR spectrum to be plotted
- `xlims`: *tuple*  
limits for the x-axis (`left`, `right`). If `None`, the whole scale is used.
- `ylims`: *tuple*  
limits for the y-axis (`left`, `right`). If `None`, the whole scale is used.
- `cmap`: *matplotlib.cm Object*  
Colour for the contour
- `c_fac`: *float*  
Contour factor parameter
- `lvl`: *float*  
height with respect to maximum at which the contour are computed
- `name`: *str*  
filename of the figure, if it has to be saved;
- `X_label`: *str*  
text of the x-axis label;
- `Y_label`: *str*  
text of the y-axis label;
- `lw`: *float*  
linewidth of the contours
- `Negatives`: *bool*  
set it to `True` if you want to see the negative part of the spectrum
- `cmapneg`: *matplotlib.cm Object*  
Colour of the negative contours



- `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `n_yticks`: *int*  
Number of numbered ticks on the y-axis of the figure
-

**3.3.9** `figures.figure2D_multi(ppm_f2, ppm_f1, datax, xlims=None, ylims=None, lvl='default', name=None, X_label='$\delta\$', F2 /ppm', Y_label='$\omega\$', F1 /ppm', lw=0.5, Negatives=False, n_xticks=10, n_yticks=10, labels=None)`

Generates the figure of multiple, superimposed spectra.

#### Parameters

- `ppm_f2`: *1darray*  
ppm scale of the direct dimension
  - `ppm_f1`: *1darray*  
ppm scale of the indirect dimension
  - `datax`: *list*  
the 2D NMR spectra to be plotted
  - `xlims`: *tuple*  
limits for the x-axis (`left`, `right`). If `None`, the whole scale is used.
  - `ylims`: *tuple*  
limits for the y-axis (`left`, `right`). If `None`, the whole scale is used.
  - `lvl`: *'default' or list*  
height with respect to maximum at which the contour are computed. If `'default'`, each spectrum is at 10% of maximum height. Otherwise, each entry of the list corresponds to the contour height of the respective spectrum.
  - `name`: *str*  
filename of the figure, if it has to be saved;
  - `X_label`: *str*  
text of the x-axis label;
  - `Y_label`: *str*  
text of the y-axis label;
  - `lw`: *float*  
linewidth of the contours
  - `Negatives`: *bool*  
set it to `True` if you want to see the negative part of the spectrum
  - `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `n_yticks`: *int*  
Number of numbered ticks on the y-axis of the figure
  - `labels`: *list*  
entries of the legend. If `None`, the legend is not drawn.
-

### 3.3.10 `figures.fitfigure(S, ppm_scale, t_AQ, V, C=False, SFO1=701.125, o1p=0, limits=None, s_labels=None, X_label='δ F1 /ppm', n_xticks=None, name=None)`

Makes the figure to show the result of a quantitative fit.

The figure will display the experimental data, the total fitting function, and the individual signals that contribute to it. The baseline is displayed only if specified.

The final plot can be either shown or saved.

#### Parameters:

- *S: 1darray*  
Spectrum to be fitted
  - *ppm\_scale: 1darray*  
Self-explanatory
  - *V: 2darray*  
matrix (# signals, parameters)
  - *C: 1darray or False*  
Coefficients of the polynomion to be used as baseline correction. If it is **False**, the baseline is not computed nor shown.
  - *limits: tuple or None*  
Trim limits for the spectrum (`left`, `right`). If **None**, the whole spectrum is used.
  - *s\_labels: list or None or False*  
Legend entries for the single components. If **None**, they are computed automatically as 1, 2, 3, etc. If it is **False**, they are not shown in the legend.
  - *X\_label: str*  
label for the x-axis.
  - *n\_xticks: int*  
number of numbered ticks that will appear in the ppm scale. An oculated choice can be very satisfying.
  - *name: str or None*  
Name with which to save the figure. If **None**, the picture is shown instead of being saved.
-

### 3.3.11 `figures.heatmap(data, zlim='auto', z_sym=True, cmap=None, xscale=None, yscale=None, rev=(False, False), n_xticks=10, n_yticks=10, n_zticks=10, fontsize=10, name=None)`

Computes a heatmap of data.

#### Parameters:

- `data`: *2darray*  
Input data
  - `zlim`: *tuple or 'auto' or 'abs'*  
Vertical limits of the heatmap, that determine the extent of the colorbar. 'auto' means `(min(data), max(data))`, 'abs' means `(min(|data|), max(|data|))`.
  - `z_sym`: *bool*  
True to symmetrize the vertical scale around 0.
  - `cmap`: *matplotlib.cm object*  
Colormap of the heatmap.
  - `xscale`: *1darray or None*  
x-scale. None means `np.arange(data.shape[1])`
  - `yscale`: *1darray or None*  
y-scale. None means `np.arange(data.shape[0])`
  - `rev`: *tuple of bool*  
Reverse scale. Format: `(x, y)`.
  - `n_xticks`: *int*  
Number of ticks of the x axis
  - `n_yticks`: *int*  
Number of ticks of the y axis
  - `n_zticks`: *int*  
Number of ticks of the color bar
  - `fontsize`: *float*  
Biggest font size to apply to the figure.
  - `name`: *str or None*  
Filename for the figure. Set to None to show the figure.
-

### 3.3.12 `figures.plot_fid(fid, name=None)`

Makes a double-panel figure that shows the real and imaginary part of the FID. X-scale and Y-scale are automatically adjusted. If the FID is of a 2D spectrum, it is flattened before to be plot.

#### Parameters

- `fid`: *ndarray*  
The data to be plot. The entries can be complex numbers.
  - `name`: *str*  
If `None`, displays the figure. Otherwise, represents the filename/path where to save the figure.
-

### 3.3.13 `figures.plot_fid_re(fid, scale=None, c='b', lims=None, name=None)`

Makes a single-panel figure that shows either the real or imaginary part of the FID. X-scale and Y-scale are automatically adjusted. If the FID is of a 2D spectrum, it is flattened before to be plot.

#### Parameters

- `fid`: *ndarray*  
The data to be plot. The entries have to be real numbers.
  - `scale`: *1darray*  
Custom x-scale for the plot. If `None`, it is computed automatically.
  - `c`: *str*  
Color of the line. Default blue (`'b'`)
  - `lims`: *tuple*  
Limits of the x-axis. Syntax: (`left`, `right`).
  - `name`: *str*  
If `None`, displays the figure. Otherwise, represents the filename/path where to save the figure.
-

### 3.3.14 `figures.redraw_contours(ax, ppm_f2, ppm_f1, S, lvl, cnt, Neg=False, Ncnt=None, lw=0.5, cmap=[None, None])`

Redraws the contours in interactive 2D visualizations.

#### Parameters:

- `ax`: *matplotlib.Subplot Object*  
Panel of the figure where to draw the contours
- `ppm_f2`: *1darray*  
ppm scale of the direct dimension
- `ppm_f1`: *1darray*  
ppm scale of the indirect dimension
- `S`: *2darray*  
Spectrum
- `lvl`: *float*  
Level at which to draw the contours
- `cnt`: *matplotlib.contour.QuadContourSet object*  
Pre-existing contours
- `Neg`: *bool*  
Choose if to draw the negative contours (**True**) or not (**False**)
- `Ncnt`: *matplotlib.contour.QuadContourSet object*  
Pre-existing negative contours
- `lw`: *float*  
Linewidth
- `cmap`: *list*  
Colour of the contours. Format: [`cmap +`, `cmap -`]

#### Returns:

- `cnt`: *matplotlib.contour.QuadContourSet object*  
Updated contours
  - `Ncnt`: *matplotlib.contour.QuadContourSet object or None*  
Updated negative contours if `Neg` is **True**, **None** otherwise
-

### 3.3.15 `figures.sns_heatmap(data, name=None)`

Computes a heatmap of `data`, which is a matrix. This function employs the `seaborn` package. Specify `name` if you want to save the figure.

#### Parameters:

- `data`: *2darray*  
Data of which to compute the heatmap. Make sure the entries are real numbers.
  - `name`: *str or None*  
Filename of the figure to be saved. If `None`, the figure is shown instead.
-



**3.3.16** `figures.stacked_plot(ppmscale, S, xlims=None, lw=0.5, name=None, X_label='$\delta\$', $ F1 /ppm', Y_label='Normalized intensity /a.u.', n_xticks=10, labels=None)`

Creates a stacked plot of all the spectra contained in the list `S`. Note that `S` MUST BE a list. All the spectra must share the same scale.

- `ppmscale`: *1darray*  
ppm scale of the spectrum
  - `S`: *list*  
spectra to be plotted
  - `xlims`: *list or tuple*  
Limits for the x-axis. If `None`, the whole scale is used.
  - `lw`: *float*  
linewidth
  - `name`: *str*  
filename of the figure, if it has to be saved;
  - `X_label`: *str*  
text of the x-axis label;
  - `Y_label`: *str*  
text of the y-axis label;
  - `n_xticks`: *int*  
Number of numbered ticks on the x-axis of the figure
  - `labels`: *list*  
labels to be put in the legend.
-

## 3.4 SIM package

This package contains function for the simulation of various features of NMR spectra, being them monodimensional or bidimensional. Functions for the simulation of whole spectra are also provided.

---

### 3.4.1 `sim.calc_splitting(u0, I0, m=1, J=0)`

Calculate the frequency and the intensities of a NMR signal splitted by scalar coupling.

**Parameters:**

- `u0`: *float*  
Frequency of the non-splitted signal (Hz)
- `I0`: *float*  
Total intensity of the non-splitted signal.
- `m`: *int*  
Multiplicity, i.e. number of expected signals after the splitting
- `J`: *float*  
Scalar coupling constant (Hz)

**Returns:**

- `u_s`: *1darray*  
Frequencies of the splitted signal (Hz)
  - `I_s`: *1darray*  
Intensities of the splitted signal
-

### 3.4.2 `sim.f_gaussian(x, u, s, A=1)`

Gaussian function in the frequency domain:

$$\mathcal{G}(x) = \frac{A}{\sqrt{2\pi}s} \exp\left[-\frac{1}{2}\left(\frac{x-u}{s}\right)^2\right]$$

#### Parameters

- `x`: *1darray*  
Independent variable
- `u`: *float*  
Peak position
- `s`: *float*  
Standard deviation
- `A`: *float*  
Intensity

#### Returns

- `f`: *1darray*  
Gaussian function.
-

### 3.4.3 `sim.f_lorentzian(x, u, fwhm, A=1)`

Lorentzian function in the time domain:

$$\mathcal{L}(x) = \frac{A}{\pi} \frac{\gamma}{(x - u)^2 + \gamma^2}$$

#### Parameters

- `x`: *1darray*  
Independent variable
- `u`: *float*  
Peak position
- `fwhm`: *float*  
Full-width at half-maximum,  $\Gamma = 2\gamma$
- `A`: *float*  
Intensity

#### Returns

- `f`: *1darray*  
Lorentzian function.
-

### 3.4.4 `sim.f_pvoigt(x, u, fwhm, A=1, x_g=0)`

Pseudo-Voigt function in the frequency domain:

$$S(x) = x_g \mathcal{G}(x) + (1 - x_g) \mathcal{L}(x)$$

This is practically done by:

---

```
s = fwhm / 2.355
S = A* (sim.f_gaussian(x, u, s, A=x_g) + sim.f_lorentzian(x, u, fwhm, A=1-x_g))
```

---

#### Parameters

- `x`: *1darray*  
Independent variable
- `u`: *float*  
Peak position
- `fwhm`: *float*  
Full-width at half-maximum
- `A`: *float*  
Intensity
- `x_g`: *float*  
Fraction of gaussianity

#### Returns

- `S`: *1darray*  
Pseudo-Voigt function.
-

### 3.4.5 `sim.gaussian_filter(ppm, u, s)`

Compute a gaussian filter to be used in order to suppress signals in the spectrum.

#### Parameters:

- ppm: *1darray*  
Scale on which to build the filter
- u: *float*  
Position of the filter
- s: *float*  
Width of the filter (standard deviation)

#### Returns:

- G: *1darray*  
Computed gaussian filter
-

### 3.4.6 `sim.load_sim_1D(File)`

Creates a dictionary from the spectral parameters listed in the input file.

Template of the input file:

---

```

B0 16.4          # Magnetic field strength /Tesla
nuc 1H           # Observed nucleus
o1p 4.7          # Pulse carrier frequency /ppm
SWp 30           # Spectral width /ppm
TD 4096          # Number of sampled complex points
shifts 0, 4.5, 3 # Peak shifts /ppm, separated by commas
fwhm 100, 100, 100 # Full-width at half maximum of the peaks /Hz, separated by commas
amplitudes 1, 4, 3 # Amplitudes of the peaks, separated by commas
x_g 0.0, 0.5, 1.0 # Fraction of gaussianity of the peaks (1 = 100% gaussian, 0 = 100%
                  lorentzian)

```

---

Use the `tab` character to separate the variable from its value. Comments are placed here to explain what the variables stand for, they are not needed in a real input file. However, if you want to put them, use the `#` character to denote them. Note that the variable names **CANNOT** be changed for any reason: the penalty is a massive sequence of errors. If only one value is supplied for the fields `shifts`, `fwhm`, `amplitudes` and `x_g`, that value must be followed by a comma in order to allow the program to recognize the list as a tuple.

#### Parameters

- File: *str*  
Path to the input file location

#### Returns

- dic: *dict*  
Dictionary of the parameters, ready to be read from the simulation functions.
-

### 3.4.7 `sim.load_sim_2D(File, states=True)`

Creates a dictionary from the spectral parameters listed in the input file.

Template of the input file:

---

```

B0 28.2          # Magnetic field strength /Tesla
nuc1 15N         # Observed nucleus in indirect dimension (F1)
nuc2 1H          # Observed nucleus in direct dimension (F2)
o1p 115         # Pulse carrier frequency /ppm in F1
o2p 5           # Pulse carrier frequency /ppm in F2
SW1p 40         # F1 Spectral width /ppm
SW2p 20         # F2 Spectral width /ppm
TD1 64          # Number of t1 increment in indirect dimension
TD2 256         # Number of sampled complex points for each transient
shifts_f1 130.0, 105.0, 120.0    # Peak F1 shifts /ppm, separated by commas
shifts_f2 0.0, 0.0, 7.0         # Peak F2 shifts /ppm, separated by commas
fwhm_f1 100, 100, 100           # Full-width at half maximum of the peaks in F1
    (/Hz), separated by commas
fwhm_f2 500, 500, 500           # Full-width at half maximum of the peaks in F2
    (/Hz), separated by commas
amplitudes 100.0, 200.0, 100.0  # Intensity of the peaks, separated by commas
x_g 0.0, 0.5, 1.0              # # Fraction of gaussianity of the peaks (1 = 100%
    gaussian, 0 = 100% lorentzian)

```

---

Use the `tab` character to separate the variable from its value. Comments are placed here to explain what the variables stand for, they are not needed in a real input file. However, if you want to put them, use the `#` character to denote them. Note that the variable names **CANNOT** be changed for any reason: the penalty is a massive sequence of errors. If only one value is supplied for the fields `shifts`, `fwhm`, `amplitudes` and `x_g`, that value must be followed by a comma in order to allow the program to recognize the list as a tuple.

#### Parameters

- `File`: *str*  
Path to the input file location
- `states`: *bool*  
If *FnMODE* is *States* or *States-TPPI*, set it to `True` to get the correct timescale.

#### Returns

- `dic`: *dict*  
Dictionary of the parameters, ready to be read from the simulation functions.
-



### 3.4.8 `sim.multiplet(u, I, m='s', J=[])`

Split a given signal according to a scalar coupling pattern.

#### Parameters:

- `u`: *float*  
Frequency of the non-splitted signal (Hz)
- `I`: *float*  
Intensity of the non-splitted signal
- `m`: *str*  
Organic chemistry-like multiplet, i.e. s, d, dqt, etc.
- `J`: *float or list*  
Scalar coupling constants. The number of constants should match the number of coupling branches

#### Returns:

- `u_in`: *list*  
List of the splitted frequencies (Hz)
  - `I_in`: *list*  
Intensities of the splitted signal
-

### 3.4.9 `sim.noisegen(size, o2, t2, s_n=1)`

Simulates additive noise in the time domain, in the form of a matrix of dimensions `size`.

This model for the noise depicts it as a white noise vector (i.e. random number that fit a gaussian distribution with 0 mean and standard deviation equal to `s_n`), modulated for the carrier frequency `o2`.

We consider the noise as the sum of a correlated contribution, due to the fact that the signal in a real spectrometer travels through the same cables until the ADC, and of a non-correlated contribution, which arises from the separation into real channel and imaginary channel.

This translates in the following code:

---

```
# correlated part of noise until ADC
white_corr = np.random.normal(0, s_n, size)
# white noise in FID has to be centered on the offset frequency
noise_corr = white_corr * np.exp(1j* 2 * np.pi * o2 * t2)

# uncorrelated part of noise: quadrature detection
white_re = np.random.normal(0, s_n, size)
white_im = np.random.normal(0, s_n, size)
# cosine-modulated in the real channel and sine-modulated in the imaginary channel
noise_re = white_re * np.cos( 2* np.pi * o2 * t2)
noise_im = white_im * np.sin( 2* np.pi * o2 * t2)

# final noise is sum of the two parts
noise = noise_corr + (noise_re + 1j*noise_im)
```

---

#### Parameters

- `size`: *int or tuple*  
Dimension of the noise matrix
- `o2`: *float*  
Carrier frequency, in Hz.
- `t2`: *1darray*  
Time scale of the last temporal dimension.
- `s_n`: *float*  
Standard deviation of the noise.

#### Returns

- `noise`: *2darray*  
Noise matrix, of dimensions `size`.
-

### 3.4.10 `sim.sim_1D(File, pv=False)`

Simulates a 1D NMR spectrum from the instructions written in `File`.

The instructions on how to write the input file are reported in the caption of `sim.load_sim_1D`.

#### Parameters

- `File`: *str*  
Path to the input file location
- `pv`: *bool*  
`True` for pseudo-Voigt model, `False` for Voigt model.

#### Returns

- `fid`: *1darray*  
FID of the simulated spectrum.
-

### 3.4.11 `sim.sim_2D(File, states=True, alt=True, pv=False)`

Simulates a 2D NMR spectrum from the instructions written in `File`. The indirect dimension is sampled with *FnMODE=States-TPPI* as default.

The instructions on how to write the input file are reported in the caption of `sim.load_sim_2D`.

#### Parameters

- `File`: *str*  
Path to the input file location
- `states`: *bool*  
Set it to `True` to allow for correct spectral arrangement in the indirect dimension.
- `alt`: *bool*  
Set it to `True` to allow for correct spectral arrangement in the indirect dimension.
- `pv`: *bool*  
`True` for pseudo-Voigt model, `False` for Voigt model.

#### Returns

- `fid`: *2darray*  
FID of the simulated spectrum.
-

### 3.4.12 `sim.t_2Dgaussian(t1, t2, v1, v2, s1, s2, A=1, states=True, alt=True)`

Bidimensional gaussian peak. The working code requires `states=True` and `alt=True`.

The signal is generated as follows:

---

```
# States acquires twice the same point of the indirect dimension time domain
t1[1::2] = t1[:,2]
# TPPI cycles the receiver phase of 90 degrees at each transient acquisition
freq_1 = np.zeros(len(t1), dtype='complex64')
for k in range(4):
    t1t = t1[k::4]
    freq_1[k::4] = np.cos( (2 * np.pi * v1 * t1t) - (0.5 * np.pi * np.mod(k,4) ))

# NMR signal in the direct dimension
F2 = np.exp(1j*2*np.pi*v2*t2) * np.exp(-(s2**2 * t2**2)/2)
# NMR signal in the indirect dimension
F1 = freq_1 * np.exp(-(s1**2 * t1**2)/2)

# The full FID is reconstructed by doing the external product between the two vectors
S = A * F1.reshape(-1,1) @ F2.reshape(1,-1)
```

---

#### Parameters

- `t1: 1darray`  
Indirect evolution timescale
- `t2: 1darray`  
Timescale of the direct dimension
- `v1: float`  
Peak position in the indirect dimension
- `v2: float`  
Peak position in the direct dimension
- `s1: float`  
Standard deviation in the indirect dimension
- `s2: float`  
Standard deviation in the direct dimension
- `A: float`  
Intensity
- `states: bool`  
Set to `True` for *FnMODE = States-TPPI*
- `alt: bool`  
Set to `True` for *FnMODE = States-TPPI*

#### Returns

- `S: 2darray`  
2D Gaussian function.
-

### 3.4.13 `sim.t_2Dlorentzian(t1, t2, v1, v2, fwhm1, fwhm2, A=1, states=True, alt=True)`

Bidimensional Lorentzian peak. The working code requires `states=True` and `alt=True`.

The signal is generated as follows:

---

```

hwhm1 = fwhm1 / 2
hwhm2 = fwhm2 / 2

# States acquires twice the same point of the indirect dimension time domain
t1[1::2] = t1[:,2]
# TPPI cycles the receiver phase of 90 degrees at each transient acquisition
freq_1 = np.zeros(len(t1), dtype='complex64')
for k in range(4):
    t1t = t1[k::4]
    freq_1[k::4] = np.cos( (2 * np.pi * v1 * t1t) - (0.5 * np.pi * np.mod(k,4) ))

# NMR signal in the direct dimension
F2 = np.exp(1j*2*np.pi*v2*t2) * np.exp(-(hwhm2 * t2))
# NMR signal in the indirect dimension
F1 = freq_1 * np.exp(-(hwhm1 * t1))

# The full FID is reconstructed by doing the external product between the two vectors
S = A * F1.reshape(-1,1) @ F2.reshape(1,-1)

```

---

#### Parameters

- `t1`: *1darray*  
Indirect evolution timescale
- `t2`: *1darray*  
Timescale of the direct dimension
- `v1`: *float*  
Peak position in the indirect dimension
- `v2`: *float*  
Peak position in the direct dimension
- `fwhm1`: *float*  
Full-width at half maximum in the indirect dimension
- `fwhm2`: *float*  
Full-width at half maximum in the direct dimension
- `A`: *float*  
Intensity
- `states`: *bool*  
Set to `True` for 'FnMODE'!'States-TPPI
- `alt`: *bool*  
Set to `True` for 'FnMODE'!'States-TPPI

## Returns

- S: *2darray*  
Lorentzian function.
-

### 3.4.14 `sim.t_2Dpvoigt(t1, t2, v1, v2, fwhm1, fwhm2, A=1, x_g=0.5, states=True, alt=True)`

Generates a 2D pseudo-voigt signal in the time domain. `x_g` states for the fraction of gaussianity, whereas `A` defines the overall amplitude of the total peak. Indexes '1' and '2' on the variables stand for 'F1' and 'F2', respectively.

---

```
# stdev computed for the gaussian part.
s1 = fwhm1 / 2.355
s2 = fwhm2 / 2.355
# Passing 's' to 'gaussian' and 'fwhm' to 'lorentzian' makes the two parts of the
# pseudo-voigt signal to have the same width and allow proper summation
G = sim.t_2Dgaussian(t1, t2, v1, v2, s1, s2, A=x_g, states=states, alt=alt)
L = sim.t_2Dlorentzian(t1, t2, v1, v2, fwhm1, fwhm2, A=(1-x_g), states=states, alt=alt)
fid = A * (G + L)
```

---

#### Parameters

- `t1`: *1darray*  
Indirect evolution timescale
- `t2`: *1darray*  
Timescale of the direct dimension
- `v1`: *float*  
Peak position in the indirect dimension
- `v2`: *float*  
Peak position in the direct dimension
- `fwhm1`: *float*  
Full-width at half maximum in the indirect dimension
- `fwhm2`: *float*  
Full-width at half maximum in the direct dimension
- `A`: *float*  
Intensity
- `x_g`: *float*  
Fraction of gaussianity
- `states`: *bool*  
Set to `True` for *FnMODE=States-TPPI*
- `alt`: *bool*  
Set to `True` for *FnMODE=States-TPPI*

#### Returns

- `fid`: *2darray*  
2D Pseudo-Voigt function.
-



### 3.4.15 `sim.t_2Dvoigt(t1, t2, v1, v2, fwhm1, fwhm2, A=1, x_g=0.5, states=True, alt=True)`

Generates a 2D Voigt signal in the time domain. `x_g` states for the fraction of gaussianity, whereas `A` defines the overall amplitude of the total peak. Indexes '1' and '2' on the variables stand for 'F1' and 'F2', respectively.

---

```
# stdev computed for the gaussian part.
s1 = fwhm1 / 2.355
s2 = fwhm2 / 2.355
# hwhm computed for the lorentzian part.
hwhm1 = fwhm1 / 2
hwhm2 = fwhm2 / 2

# States acquires twice the same point of the indirect dimension time domain
t1[1::2] = t1[:,2]

# direct dimension
# frequency
freq_2 = np.exp(1j * 2 * np.pi * v2 * t2)

# Add line-broadening, fist lorentzian then gaussian, using:
# hwhm' = (1 - x_g) * hwhm      for L
# s' = x_g * s                  for G
F2 = freq_2 * np.exp(-(1-x_g)*hwhm2 * t2) * np.exp(-((x_g*s2)**2 * t2**2)/2)

# indirect dimension
# Redfield cycles the receiver phase of 90 degrees at each transient acquisition
freq_1 = np.zeros(len(t1), dtype='complex64')
for k in range(4):
    t1t = t1[k::4]
    freq_1[k::4] = np.cos( (2 * np.pi * v1 * t1t) - (0.5 * np.pi * np.mod(k,4) ))

# Add line-broadening, fist lorentzian then gaussian, using:
# hwhm' = (1 - x_g) * hwhm      for L
# s' = x_g * s                  for G
F1 = freq_1 * np.exp(-(1-x_g) * hwhm1 * t1) * np.exp(-((x_g*s1)**2 * t1**2)/2)

# The full FID is reconstructed by doing the external product between the two vectors
S = A * F1.reshape(-1,1) @ F2.reshape(1,-1)
return S
```

---

#### Parameters

- `t1`: *1darray*  
Indirect evolution timescale
- `t2`: *1darray*  
Timescale of the direct dimension
- `v1`: *float*  
Peak position in the indirect dimension
- `v2`: *float*  
Peak position in the direct dimension

- *fwhm1: float*  
Full-width at half maximum in the indirect dimension
- *fwhm2: float*  
Full-width at half maximum in the direct dimension
- *A: float*  
Intensity
- *x\_g: float*  
Fraction of gaussianity
- *states: bool*  
Set to **True** for 'FnMODE': 'States-TPPI'
- *alt: bool*  
Set to **True** for 'FnMODE': 'States-TPPI'

## Returns

- *S: 2darray*  
Voigt function.
-

### 3.4.16 `sim.t_gaussian(t, u, s, A=1, phi=0)`

Gaussian function in the time domain.

$$g(x) = Ae^{i\phi} e^{i2\pi ut} e^{-s^2 t^2 / 2}$$

#### Parameters

- `t`: *1darray*  
Independent variable
- `u`: *float*  
Peak position
- `s`: *float*  
Standard deviation
- `A`: *float*  
Intensity
- `phi`: *float*  
Phase, in radians

#### Returns

- `S`: *1darray*  
Gaussian function.
-

### 3.4.17 `sim.t_lorentzian(t, u, fwhm, A=1, phi=0)`

Lorentzian function in the time domain.

$$\ell(x) = Ae^{i\phi} e^{i2\pi ut} e^{-\gamma t}$$

#### Parameters

- `t`: *1darray*  
Independent variable
- `u`: *float*  
Peak position
- `fwhm`: *float*  
Full-width at half-maximum,  $\Gamma = 2\gamma$
- `A`: *float*  
Intensity
- `phi`: *float*  
Phase, in radians

#### Returns

- `S`: *1darray*  
Lorentzian function.
-

### 3.4.18 `sim.t_pvoigt(t, u, fwhm, A=1, x_g=0, phi=0)`

Pseudo-Voigt function in the time domain:

$$s(x) = x_g g(x) + (1 - x_g) \ell(x)$$

---

```
s = fwhm / 2.355
```

```
S = A * (sim.t_gaussian(t, u, s, A=x_g, phi=phi) + sim.t_lorentzian(t, u, fwhm, A=1-x_g,
    phi=phi))
```

---

#### Parameters

- `t`: *1darray*  
Independent variable
- `u`: *float*  
Peak position
- `fwhm`: *float*  
Full-width at half-maximum
- `A`: *float*  
Intensity
- `x_g`: *float*  
Fraction of gaussianity
- `phi`: *float*  
Phase, in radians

#### Returns

- `S`: *1darray*  
Pseudo-Voigt function.
-

### 3.4.19 `sim.t_voigt(t, u, fwhm, A=1, x_g=0, phi=0)`

Voigt function in the time domain. The parameter `x_g` affects the linewidth of the lorentzian and gaussian contributions.

---

```
s = fwhm / 2.355
S = A * np.exp(1j*phi) * sim.t_gaussian(t, u/2, s*x_g) * sim.t_lorentzian(t, u/2,
    fwhm*(1-x_g))
```

---

#### Parameters

- `t`: *1darray*  
Independent variable
- `u`: *float*  
Peak position
- `fwhm`: *float*  
Full-width at half-maximum
- `A`: *float*  
Intensity
- `x_g`: *float*  
Fraction of gaussianity
- `phi`: *float*  
Phase, in radians

#### Returns

- `S`: *1darray*  
Voigt function.
-

### 3.4.20 `sim.water7(N, t2, vW, fwhm=300, A=1, spread=701.125)`

Simulates a feature like the water ridge in HSQC spectra, in the time domain.

This signal is modelled as a gaussian signal which does not encode for any frequency in the indirect dimension, and whose chemical shift moves due to field drifts through the various transients according to a gaussian distribution. This signal is on-phase in the even transients and 90°-dephased in the odd transients.

#### Parameters

- `N`: *int*  
Number of transients
- `t2`: *1darray*  
Time scale of the last temporal dimension.
- `vW`: *float*  
Nominal peak position, in Hz.
- `fwhm`: *float*  
Nominal full-width at half maximum of the peak.
- `A`: *float*  
Signal intensity.
- `spread`: *float*  
Standard deviation of the peak position distribution, in Hz.

#### Returns

- `ridge`: *2darray*  
Matrix of the ridge.
-

## 3.5 FIT package

Functions for performing fits.

---

### 3.5.1 fit.Voigt\_Fit

class

This class offers an 'interface' to fit a 1D NMR spectrum.

#### Attributes:

- ppm\_scale: *1darray*  
Self-explanatory
- S : *1darray*  
Spectrum to fit. Only real part
- t\_AQ: *1darray*  
acquisition timescale of the spectrum
- SFO1: *float*  
Larmor frequency of the nucleus
- o1p : *float*  
Pulse carrier frequency
- nuc : *str or None*  
Nucleus. Used to write the X\_scale of the plot.
- input\_file : *str*  
filename of the input file
- output\_file : *str*  
filename of the output file
- log\_file : *str*  
filename of the log file
- limits : *tuple*  
borders of the fitting window
- Vi : *2darray*  
array with the values of the signals used as initial guess
- Ci : *1darray*  
coefficients of the baseline polynomion as initial guess
- Vf : *2darray*  
array with the values of the signals after the fit
- Cf : *1darray*  
coefficients of the baseline polynomion after the fit
- s\_labels : *list*  
legend entries for the single signals.



## Methods

- `__init__(self, ppm_scale, S, t_AQ, SFO1, o1p, nuc=None):`  
Add common variables
  - `iguess(self, input_file, limits=None):`  
Create initial guess and writes the input file if not present
  - `dofit(self, log_file='fit.log', output_file='fit.out', utol=0.5, vary_phi=False, vary_xg=False, res_hist_name='histogram_of_residuals', test_res=True):`  
Fit the data, writes the output file and the log file
  - `plot(self, what, name=None, s_labels=None, X_label='δ F1 /ppm', n_major_ticks=10):`  
plot either the initial guess (**what='iguess'**) or the fitted data (**what='fit'**).
-

### 3.5.2 `fit.ax_histogram(ax, data, nbins=100, density=True, f_lims=None, xlabel=None, x_symm=False, barcolor='tab:blue')`

Computes an histogram of `data` and tries to fit it with a gaussian lineshape. The parameters of the gaussian function are calculated analytically directly from `data`.

#### Parameters:

- `ax` : *matplotlib.subplot Object*  
panel of the figure where to put the histogram
- `data` : *ndarray*  
the data to be binned
- `nbins` : *int*  
number of bins to be calculated
- `density` : *bool*  
`True` for normalize histogram
- `f_lims` : *tuple or None*  
limits for the x-axis of the figure
- `xlabel` : *str or None*  
Text to be displayed under the x-axis
- `x_symm` : *bool*  
set it to `True` to make symmetric x-axis with respect to 0
- `barcolor`: *str*  
Color of the bars of the histogram

#### Returns:

- `m` : *float*  
Mean of `data`
  - `s` : *float*  
Standard deviation of `data`.
-

### 3.5.3 `fit.bin_data(data0, nbins=100, density=True, x_symm=False)`

Computes the histogram of `data`, sampling it into `nbins` bins. If `data` is multidimensional, it is flattened.

#### Parameters:

- `data` : *ndarray*  
the data to be binned
- `nbins` : *int*  
number of bins to be calculated
- `density` : *bool*  
True for normalize `data`
- `x_symm` : *bool*  
set it to `True` to make symmetric x-axis with respect to 0

#### Returns:

- `hist`: *1darray*  
The bin intensity
  - `bin_scale`: *1darray*  
Scale built with the mean value of the bin widths.
-

### 3.5.4 `fit.build_baseline(ppm_scale, C, L=None)`

Builds the baseline calculating the polynomial with the given coefficients, and summing up to the right position.

#### Parameters:

- `ppm_scale`: *1darray*  
ppm scale of the spectrum
- `C`: *list*  
Baseline coefficients. No baseline corresponds to `False`.
- `L`: *list*  
List of window regions. If it is `None`, the baseline is built on the whole `ppm_scale`

#### Returns:

- `baseline`: *1darray*  
Self-explanatory.
-

### 3.5.5 `fit.calc_fit_lines(ppm_scale, limits, t_AQ, SFO1, o1p, N, V, C=False)`

Given the values extracted from a fit input/output file, calculates the signals, the total fit function, and the baseline.

#### Parameters:

- `ppm_scale`: *1darray*  
PPM scale of the spectrum
- `limits`: *tuple*  
(`left`, `right`) in ppm
- `t_AQ`: *1darray*  
Acquisition timescale
- `SFO1`: *float*  
Larmor frequency of the nucleus /ppm
- `o1p`: *float*  
Pulse carrier frequency /ppm
- `N`: *int*  
Size of the final spectrum.
- `V`: *2darray*  
Matrix containing the values to build the signals.
- `C`: *1darray*  
Baseline polynomial coefficients. `False` to not use the baseline

#### Returns:

- `sgn`: *list*  
Voigt signals built using `V`
  - `Total`: *1darray*  
sum of all the arrays in `sgn`
  - `baseline`: *1darray*  
Polynomial built using `C` as coefficients. `False` if `C` is `False`.
-

### 3.5.6 `fit.dic2mat(dic, peak_names, ns, A=None)`

This is used to make the matrix of the parameters starting from a dictionary like the one produced by *lmfit*. The column of the total intensity is not added, unless the parameter `A` is passed. In this case, the third column (which is the one with the relative intensities) is corrected using the function `misc.molfrac`.

#### Parameters:

- `dic` : *dict*  
input dictionary
- `peak_names` : *list*  
list of the parameter entries to be looked for
- `ns` : *int*  
number of signals to unpack
- `A` : *float or None*  
Total intensity.

#### Returns:

- `V` : *2darray*  
Matrix containing the parameters.
-

### 3.5.7 `fit.fit_int(y, y_c)`

Calculate the intensity according to the least square fit as:

$$I = \frac{\sum \text{obs} \cdot \text{calc}}{\sum \text{calc}^2}$$

#### Parameters:

- `y`: *ndarray*  
Observed data.
- `y_c`: *ndarray*  
Calculated data

#### Returns:

- `I`: *float*  
Calculated intensity
-

### 3.5.8 `fit.gaussian_fit(x, y)`

Fit `y` with a gaussian function, built using `x` as independent variable. The gaussian function is built with `sim.f_gaussian`.

#### Parameters:

- `x` : *1darray*  
x-scale
- `y` : *1darray*  
data to be fitted

#### Returns:

- `u` : *float*  
mean
  - `s` : *float*  
standard deviation
  - `A` : *float*  
Integral
-



### 3.5.9 `fit.gen_iguess(x, experimental, param, model, model_args=[])`

GUI for the interactive setup of a Parameters object to be used in a fitting procedure. Once you initialized the Parameters object with the name of the parameters and a dummy value, you are allowed to set the value, minimum, maximum and vary status through the textboxes given in the right column, and see their effects in real time. Upon closure of the figure, the Parameters object with the updated entries is returned. A maximum of 18 parameters will fit the figure.

#### Parameters:

- `x`: *1darray*  
Independent variable
- `experimental`: *1darray*  
The objective values you are trying to fit
- `param`: *lmfit.Parameters Object*  
Initialized parameters object
- `model`: *function*  
Function to be used for the generation of the fit model. `param` must be the first argument.
- `model_args`: *list*  
List of args to be passed to `model`, after `param`

#### Returns:

- `param`: *lmfit.Parameters Object*  
Updated Parameters Object
-

### 3.5.10 `fit.get_region(ppmscale, S, rev=True)`

Interactively select the spectral region to be fitted. Returns the border ppm values.

#### Parameters:

- `ppmscale`: *1darray*  
The ppm scale of the spectrum
- `S`: *1darray*  
The spectrum to be trimmed
- `rev`: *bool*  
Choose if to reverse ppm scale and data (**True**) or not (**False**).

#### Returns:

- `left`: *float*  
Left border of the selected spectral window
  - `right`: *float*  
Right border of the selected spectral window
-

### 3.5.11 `fit.histogram(data, nbins=100, density=True, f_lims= None, xlabel=None, x_symm=False, name=None, barcolor='tab:blue')`

Computes an histogram of `data` and tries to fit it with a gaussian lineshape. The parameters of the gaussian function are calculated analytically directly from `data`.

#### Parameters:

- `data` : *ndarray*  
the data to be binned
- `nbins` : *int*  
number of bins to be calculated
- `density` : *bool*  
`True` for normalize data
- `f_lims` : *tuple or None*  
limits for the x-axis of the figure
- `xlabel` : *str or None*  
Text to be displayed under the x-axis
- `x_symm` : *bool*  
set it to `True` to make symmetric x-axis with respect to 0
- `name` : *str*  
name for the figure to be saved
- `barcolor`: *str*  
Color of the bars of the histogram

#### Returns:

- `m` : *float*  
Mean of `data`
  - `s` : *float*  
Standard deviation of `data`.
-

### 3.5.12 `fit.integrate(ppm0, data0, X_label='$\delta$', $F1 /ppm')`

Allows interactive integration of a NMR spectrum through a dedicated GUI. Returns the values as a dictionary, where the keys are the selected regions truncated to the 2nd decimal figure. The returned dictionary contains some predefined keys, as follows:

- `total`: total integrated area
- `ref_pos`: location of the reference peak (`ppm1 : ppm2`)
- `ref_int`: absolute integral of the reference peak
- `ref_val`: for how many nuclei the reference peak integrates

The absolute integral of the  $x$ -th peak,  $I_x$ , must be calculated according to the formula:

$$I_x = I_x^{\text{relative}} \cdot \frac{\text{ref\_int}}{\text{ref\_val}}$$

#### Parameters:

- `ppm`: *1darray*  
PPM scale of the spectrum
- `data`: *1darray*  
Spectrum to be integrated.
- `X_label`: *str*  
Label of the x-axis

#### Returns:

- `f_vals`: *dict*  
Dictionary containing the values of the integrated peaks.
-

### 3.5.13 `fit.integrate_2D(ppm_f1, ppm_f2, data, SFO1, SFO2, fwhm_1=200, fwhm_2=200, utol_1=0.5, utol_2=0.5, plot_result=False)`

Function to select and integrate 2D peaks of a spectrum, using dedicated GUIs. Calls `integral_2D` to do the dirty job.

#### Parameters:

- `ppm_f1`: *1darray*  
PPM scale of the indirect dimension
- `ppm_f2`: *1darray*  
PPM scale of the direct dimension
- `data`: *2darray*  
real part of the spectrum
- `SFO1`: *float*  
Larmor frequency of the nucleus in the indirect dimension
- `SFO2`: *float*  
Larmor frequency of the nucleus in the direct dimension
- `fwhm_1`: *float*  
Starting FWHM /Hz in the indirect dimension
- `fwhm_2`: *float*  
Starting FWHM /Hz in the direct dimension
- `utol_1`: *float*  
Allowed tolerance for `u_1` during the fit. (`u_1-utol_1`, `u_1+utol_1`)
- `utol_2`: *float*  
Allowed tolerance for `u_2` during the fit. (`u_2-utol_2`, `u_2+utol_2`)
- `plot_result`: *bool*  
True to show how the program fitted the traces.

#### Returns:

- `I`: *dict*  
Computed integrals. The keys are '`<ppm f1>:<ppm f2>`' with 2 decimal figures.
-

### 3.5.14 `fit.interactive_smoothing(x, y, cmap='RdBu')`

Interpolate the given data with a 3rd-degree spline. Type the desired smoothing factor in the box and see the outcome directly on the figure. When the panel is closed, the smoothed function is returned.

#### Parameters:

- `x`: *1darray*  
Scale of the data
- `y`: *1darray*  
Data to be smoothed
- `cmap`: *str*  
Name of the colormap to be used to represent the weights

#### Returns:

- `sx`: *1darray*  
Location of the spline points
  - `sy`: *1darray*  
Smoothed y
  - `s_f`: *float*  
Employed smoothing factor for the spline
  - `weights`: *1darray*  
Weights vector
-

### 3.5.15 `fit.join_par(filenamees, ppm_scale, joined_name=None)`

Load a series of parameters fit files. Join them together, returning a unique array of signal parameters, a list of coefficients for the baseline, and a list of tuples for the regions. Also, uses the coefficients and the regions to directly build the baseline according to the ppm windows.

#### Parameters:

- `filenamees`: *list*  
List of directories of the input files.
- `ppm_scale`: *1darray*  
ppm scale of the spectrum. Used to build the baseline
- `joined_name`: *str or None*  
If it is not `None`, concatenates the files in the list `filenamees` and saves them in a single file named `joined_name`.

#### Returns:

- `V`: *2darray*  
Array of joined signal parameters
  - `C`: *list*  
Baseline polynomion coefficients. No baseline corresponds to `False`.
  - `L`: *list*  
List of window regions.
  - `baseline`: *1darray*  
Baseline built from `C` and `L`.
-

### 3.5.16 `fit.make_iguess(S, ppm_scale, t_AQ, limits=None, SFO1=701.125, o1p=0, rev=True, name='i_guess.inp')`

Compute the initial guess for the quantitative fit of 1D NMR spectrum in an interactive manner. When the panel is closed, the values are saved in a file.

#### Parameters:

- `S` : *1darray*  
Spectrum to be fitted
- `ppm_scale` : *1darray*  
Self-explanatory
- `t_AQ` : *1darray*  
Acquisition timescale
- `limits` : *tuple or None*  
Trim limits for the spectrum (`left`, `right`). If `None`, the whole spectrum is used.
- `SFO1` : *float*  
Larmor frequency /MHz
- `o1p` : *float*  
pulse carrier frequency /ppm
- `rev` : *bool*  
choose if you want to reverse the x-axis scale (`True`) or not (`False`).
- `name` : *str*  
name of the file where to save the parameters

#### Returns:

- `V_f` : *2darray*  
matrix (# signals, parameters)
  - `C_f` : *1darray or False*  
Coefficients of the polynomion to be used as baseline correction. If the 'baseline' checkbox in the interactive figure panel is not checked, `C_f` is `False`.
-



### 3.5.17 `fit.make_signal(t, u, s, k, x_g, phi, A, SFO1=701.125, o1p=0, N=None)`

Generates a Voigt signal using `sim.t_voigt` on the basis of the specified parameters. Then, makes the Fourier transform and returns it.

#### Parameters:

- `t` : *ndarray*  
acquisition timescale
- `u` : *float*  
chemical shift, in ppm
- `s` : *float*  
full-width at half-maximum, in hertz
- `k` : *float*  
relative intensity
- `x_g` : *float*  
fraction of gaussianity
- `phi` : *float*  
phase of the signal, in degrees
- `A` : *float*  
total intensity
- `SFO1` : *float*  
Larmor frequency, in MHz
- `o1p` : *float*  
pulse carrier frequency, in ppm
- `N` : *int or None*  
length of the final signal array. If `None`, the signal is not zero-filled before to be transformed.

#### Returns:

- `sgn` : *1darray*  
generated signal in the frequency domain (just the real part).
-

### 3.5.18 `fit.print_par(V, C, limits=[None,None])`

Prints on screen the same thing that `fit.write_par` writes in a file.

#### Parameters:

- `V` : *2darray*  
matrix (# signals, parameters)
  - `C` : *1darray or False*  
Coefficients of the polynomial to be used as baseline correction. If it is `False`, they are not printed.
  - `limits` : *tuple or None*  
Trim limits for the spectrum (`left`, `right`).
-

### 3.5.19 `fit.read_par(filename)`

Reads the input file of the fit and returns the values.

#### Parameters:

- `filename`: *str*  
directory and name of the input file to be read

#### Returns:

- `V`: *2darray*  
matrix (# signals, parameters)
  - `C`: *1darray or False*  
Coefficients of the polynomial to be used as baseline correction. If the corresponding flag is not found in the file, `C_f` is set to `False`.
  - `limits`: *tuple*  
Trim limits for the spectrum (`left`, `right`).
-

### 3.5.20 `fit.smooth_spl(x, y, s_f=1, size=0, weights=None)`

Fit the input data with a 3rd-order spline, given the smoothing factor to be applied.

#### Parameters:

- `x`: *1darray*  
Location of the experimental points
- `y`: *1darray*  
Input data to be fitted
- `s_f`: *float*  
Smoothing factor of the spline. 0=best straight line, 1=naive spline.
- `size`: *int*  
Size of the spline. If `size` is 0, the same dimension as `y` is chosen.
- `weights`: *1darray*  
Array of weights of the spline points. `None` assign the same weight to all points in `x`.

#### Returns:

- `x_s`: *1darray*  
Location of the spline data points.
  - `y_s`: *1darray*  
Spline that fits the data.
-

### 3.5.21 `fit.test_residuals(R, nbins=100, density=False)`

Test the residuals of a fit to see if it was reliable. Returns two values: `SYSDEV` and `Q_G`.

`SYSDEV` is inspired by Svergun's Gnom, and it tells if there are systematic deviations basing on the number of sign changes in the residual. Optimal value must be 1. Values greater than 1 show positive bias, values lesser than 1 show negative bias.

Let  $R$  be an array of  $N$  points, and let  $N_s$  be the number of sign changes in  $R$ . Then:

$$\text{SYSDEV} = \frac{N_s}{N/2}$$

`Q_G` is to see the discrepancy between a gaussian function built with the mean and standard deviation of the residuals and the gaussian built fitting the histogram of the residuals. Values go from 0 (worst case) to 1 (best case).

Let  $x$  be the scale of the bin edges, and let  $G_T(x)$  and  $G_F(x)$  be the theoretical and the fitted gaussians respectively.

$$G_T(x) = \frac{A_T}{\sqrt{2\pi}\sigma_T} \exp\left[-\frac{1}{2}\left(\frac{x - \mu_T}{\sigma_T}\right)^2\right] \quad G_F(x) = \frac{A_F}{\sqrt{2\pi}\sigma_F} \exp\left[-\frac{1}{2}\left(\frac{x - \mu_F}{\sigma_F}\right)^2\right]$$

Then:

$$\text{Q\_G} = \frac{\int |G_T(x) - G_F(x)| dx}{\int G_T(x) dx + \int G_F(x) dx}$$

The numerator of this fraction expresses the area of the superposition region of the two gaussians. This value is then related to the worst possible case, i.e. when the two gaussians are totally disjointed.

#### Parameters:

- `R` : *1darray*  
Array of the residuals
- `nbins` : *int*  
number of bins of the histogram, which is also the length of  $x$ .
- `density` : *bool*  
True to normalize the histogram, False otherwise.

#### Returns:

- `SYSDEV` : *float*  
Read full caption
  - `Q_G` : *float*  
Read full caption
-

**3.5.22** `fit.voigt_fit(S, ppm_scale, V, C, t_AQ, limits=None, SFO1=701.125, o1p=0, utol=0.5, vary_phi=False, vary_xg=True, hist_name=None, write_out='fit.out', test_res=True)`

Fits an NMR spectrum with a set of signals, whose parameters are specified in the `V` matrix. There is the possibility to use a baseline through the parameter `C`. The signals are computed in the time domain and then Fourier transformed.

#### Parameters:

- `S` : *1darray*  
Spectrum to be fitted
- `ppm_scale` : *1darray*  
Self-explanatory
- `V` : *2darray*  
matrix (# signals, parameters)
- `C` : *1darray or False*  
Coefficients of the polynomial to be used as baseline correction. If it is `False`, the baseline correction is not used.
- `t_AQ` : *1darray*  
Acquisition timescale
- `limits` : *tuple or None*  
Trim limits for the spectrum (`left`, `right`). If `None`, the whole spectrum is used.
- `SFO1` : *float*  
Larmor frequency /MHz
- `o1p` : *float*  
pulse carrier frequency /ppm
- `utol` : *float*  
tolerance for the chemical shift. The peak center can move in the range  $[\mu - \text{utol}, \mu + \text{utol}]$ .
- `vary_xg`: *bool*  
If it is `False`, the parameter `x_g` cannot be varied during the fitting procedure. Useful when fitting with pure Gaussians or pure Lorentzians.
- `vary_basl`: *bool*  
If it is `False`, the baseline is kept fixed at the initial parameters.

#### Returns:

- `C_f` : *1darray or False*  
Coefficients of the polynomial to be used as baseline correction, or just `False` if not used.
  - `V_f` : *2darray*  
matrix (# signals, parameters) after the fit
  - `result` : *lmfit.fit\_result Object*  
container of all information on the fit
-

### 3.5.23 `fit.write_log(input_file, output_file, limits, V_i, C_i, V_f, C_f, result, runtime, test_res=True, log_file='fit.log')`

Write a log file with all the information of the fit.

#### Parameters:

- `input_file`: *str*  
Location and filename of the input file
  - `output_file`: *str*  
Location and filename of the output file
  - `limits`: *tuple*  
Delimiters of the spectral region that was fitted. (`left`, `right`)
  - `V_i`: *2darray*  
Initial parameters of the fit
  - `C_i`: *1darray or False*  
Coefficients of the starting polynomial used for baseline correction. If `False`, they are not written.
  - `V_f`: *2darray*  
Final parameters of the fit
  - `C_f`: *1darray or False*  
Coefficients of the final polynomial used for baseline correction. If `False`, they are not written.
  - `result`: *lmfit.FitResult Object*  
Object returned by *lmfit* after the fit.
  - `runtime`: *datetime.datetime Object*  
Time taken for the fit
  - `test_res`: *bool*  
Choose if to test the residual with the `fit.test_residual` function (`True`) or not (`False`)
  - `log_file`: *str*  
Filename of the log file to be saved.
-

### 3.5.24 `fit.write_par(V, C, limits, filename='i_guess.inp')`

Write the parameters of the fit, whether they are input or output.

#### Parameters:

- *V* : *2darray*  
matrix (# signals, parameters)
  - *C* : *1darray or False*  
Coefficients of the polynomial to be used as baseline correction. If it is **False**, they will not appear in the written file.
  - *limits* : *tuple*  
Trim limits for the spectrum (**left**, **right**).
  - *filename*: *str or TextIO Object*  
directory and name of the file to be written, or directly a file opened with **open** and writing rights.
-



## 3.6 SPECTRA package

All the classes in the `Spectra` module are automatically imported together with `klassez` itself.

They are:

- `Spectrum_1D`
- `pSpectrum_1D`
- `Spectrum_2D`
- `pSpectrum_2D`
- `Pseudo_2D`

Classes of spectra with same dimensionality share more or less the same attributes. A list of them is reported in the following sections. Regarding the methods, please refer to the examples reported in the *User guide* section to understand how to use them, or use the functions `help()`, `vars()`, `dir()` to get detailed info on how they exactly work.

### 3.6.1 Spectrum\_1D important attributes

Initialized attributes:

- `fid` : *1darray*  
FID of the experiment.
- `acqus` : *dict*  
Dictionary containing the important acquisition parameters.
- `procs` : *dict*  
Dictionary containing the important processing parameters.
- `ngdic` : *dict*  
Dictionary containing all the information about your experiments, as returned by *nmrglue*. Available for non-simulated data only.

Created after `process()`:

- `freq` : *1darray*  
Frequency scale of the spectrum.
- `ppm` : *1darray*  
PPM scale of the spectrum.
- `S` : *1darray*  
Complex spectrum.
- `r` : *1darray*  
Real part of the spectrum.
- `i` : *1darray*  
Imaginary part of the spectrum.
- `F` : *fit.Voigt\_Fit Object*  
Fitting interface initialized with the current attributes of the class.

### 3.6.2 Spectrum\_2D important attributes

Initialized attributes:

- **fid** : *2darray*  
FID of the experiment.
- **acqus** : *dict*  
Dictionary containing the important acquisition parameters.
- **procs** : *dict*  
Dictionary containing the important processing parameters.
- **ngdic** : *dict*  
Dictionary containing all the information about your experiments, as returned by *nmrglue*. Available for non-simulated data only.

Created after **process()**:

- **freq\_f1** : *1darray*  
Frequency scale of the indirect dimension of the spectrum.
- **freq\_f2** : *1darray*  
Frequency scale of the direct dimension of the spectrum.
- **ppm\_f1** : *1darray*  
PPM scale of the indirect dimension of the spectrum.
- **ppm\_f2** : *1darray*  
PPM scale of the direct dimension of the spectrum.
- **S** : *2darray*  
(Hyper)Complex spectrum, depending on the acquisition scheme of the indirect dimension.
- **rr** : *2darray*  
Real part of the spectrum.
- **ir** : *2darray*  
Imaginary part of the direct dimension, real part of the indirect dimension.
- **ri** : *2darray*  
Real part of the direct dimension, imaginary part of the indirect dimension.
- **ii** : *2darray*  
Imaginary part of the direct dimension, imaginary part of the indirect dimension.
- **Trf1**: *dict of pSpectrum\_1D Objects*  
Projection of the indirect dimension, obtained by invoking **projf1(a,b)**. The keys of the dictionary are [**f**'{a}:{b}'].
- **Trf2**: *dict of pSpectrum\_1D Objects*  
Projection of the direct dimension, obtained by invoking **projf2(a,b)**. The keys of the dictionary are [**f**'{a}:{b}'].