



Exploring the limits of network topology estimation using diffusion-based tractography and tracer studies in the macaque cortex

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ABSTRACT

Reconstructing the anatomical pathways of the brain to study the human connectome has become an important endeavour for understanding brain function and dynamics. Reconstruction of the cortico-cortical connectivity matrix *in vivo* often relies on noninvasive diffusion-weighted imaging (DWI) techniques but the extent to which they can accurately represent the topological characteristics of structural connectomes remains unknown. We addressed this question by constructing connectomes using DWI data collected from macaque monkeys *in vivo* and with data from published invasive tracer studies. We found the strength of fiber tracts was well estimated from DWI and topological properties like degree and modularity were captured by tractography-based connectomes. Rich-club/core-periphery type architecture could also be detected but the classification of hubs using betweenness centrality, participation coefficient and core-periphery identification techniques was inaccurate. Our findings indicate that certain aspects of cortical topology can be faithfully represented in noninvasively-obtained connectomes while other network analytic measures warrant cautionary interpretations.

1. Introduction

Network structure is thought to play a prominent role in supporting healthy brain function (Fornito et al., 2015; Griffa et al., 2013). A large body of work has been devoted to the analysis of the brain's structural topology in order to characterize and infer how functional networks emerge from large-scale structural connectivity, or the “connectome” (Park and Friston, 2013; Sporns, 2014; Zuo et al., 2016). In humans, the characterization of network structure relies mainly on noninvasive techniques such as tractography using diffusion-weighted magnetic resonance imaging (DWI). A number of influential observations about brain organization in both health and disease have been made based on DWI data (Baum et al., 2017; Crossley et al., 2014; Perry et al., 2015; van den Heuvel et al., 2010; van den Heuvel and Sporns, 2011; Zalesky et al., 2011). Recent validation studies in the macaque have demonstrated how a general correspondence exists between DWI-based estimates of structural connectivity, specifically “connection strength” (usually taken as some derivative of the number of streamlines between two regions), and those derived from the gold standard invasive technique of using tract tracers to map axonal projections. DWI-based tractography has been

shown to correctly detect the presence of a large proportion of connections across the visual system (Azadbakht et al., 2015) and DWI-based estimates of connection strengths are correlated to those obtained from tracer studies (Donahue et al., 2016; van den Heuvel et al., 2015). However, even with extremely high-resolution DWI, probabilistic tractography suffers from a steep trade-off between sensitivity and specificity whereby obtaining a large proportion of true positive connections is accompanied by a large number of false positives and the optimal parameter settings for tractography (e.g., curvature thresholds) can vary widely depending on the location of the seed (Thomas et al., 2014; also see Maier-Hein et al., 2017). The ability of tractography to properly reconstruct the connectivity of the human brain and, in particular, the interpretation of detected streamlines (Jones et al., 2013), remains a matter of debate.

Existing validation studies in the macaque have only examined the accuracy of DWI-based tractography at the level of the individual connection. However, a major use of tractography has been to study the human brain at the level of large-scale whole-brain networks. The extent to which tractography can accurately capture the brain's structural topology remains unknown. While some studies have shown that

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connectomes generated from tracer studies exhibit similar network organization principles as those reported using DWI data (e.g., [de Reus and van den Heuvel, 2013a](#); [Harriger et al., 2012](#)), it is still unclear whether the topologies of networks obtained from the two different modalities actually coincide. Most previous studies have also been limited to tractography within a single hemisphere and usually using only a few *ex vivo* specimens, where DWI scans are of optimal quality and are not affected by artifacts such as motion or physiological noise. In this study, we used DWI data obtained from 10 macaque monkeys in combination with macaque connectivity described by published tracer studies to determine whether probabilistic tractography can accurately represent whole-brain structural topology *in vivo*. Given that tractography's accuracy varies greatly as a function of its parameter settings ([Dauguet et al., 2007](#); [Jones et al., 2013](#); [Thomas et al., 2014](#)), we first systematically varied tractography parameters to determine the optimal settings for constructing whole-brain connectomes in the macaque. Using these optimized connectomes in conjunction with network analytic tools, we then determined the extent to which connectomes derived from DWI accurately captured the structural network characteristics of the macaque brain. We replicate previous findings that tractography can detect the presence and/or absence of connections above chance levels and can also provide reasonable estimates of connection strengths. In the macaque, more accurate connectomes were obtained by lowering the curvature threshold and discarding a small percentage of the weakest connections. However, owing to the high false positive rates in tractography-based connectomes, their ability to accurately capture critical aspects of structural topology was dependent on the robustness of the network analytic measure in question to misidentified connections.

2. Results

Probabilistic tractography was performed using an FSL-based pipeline on diffusion-weighted magnetic resonance imaging data collected from 10 macaque monkeys at 7T. Two different parcellations, a single-hemisphere one (“Markov-Kennedy” ([Markov et al., 2014](#))) and a whole-cortex parcellation (“RM-CoCo” ([Bezgin et al., 2012](#); [Kötter and Wanke, 2005](#))) were used and tractography parameters (angular threshold and distance correction) were systematically varied (see Materials and Methods). Tractography-derived connectivity matrices for various parameter combinations for an example subject are shown alongside the tracer-derived matrices in [Fig. 1](#). For the purposes of this paper, we use the term connection “strength” to refer to the number or proportion of axons running between two regions in the case of tract tracing data and the number or proportion of streamlines running between two regions for DWI-based tractography.

The tracer networks were symmetrized to enable comparison to the undirected tractography networks. The density of the Markov-Kennedy tracer network was 79.3% while that of the RM-CoCo tracer was 55.9% (intra-hemispheric: 79.2%; inter-hemispheric: 35.8%).

2.1. Effects of varying tractography parameters on accuracy

On the assumption that the tracer-derived networks serve as a “ground truth” for the large-scale anatomical connectivity of the macaque brain, we computed a number of accuracy measures to determine the ability for diffusion-weighted tractography to reconstruct anatomical connectivity from data collected *in vivo*. These included the percentage of connections correctly represented, the area under the ROC curve (AUC), and corresponding measures of sensitivity, specificity and precision. Since these measures could be affected by the high density of the networks being compared (i.e., the chances of correctly detecting a connection using DWI are high simply due to the large number of existing connections in the tracer networks), we compared the observed accuracy values to those computed from 1000 randomly connected networks of the same size and density as the observed networks.

We first consider intra-hemispheric tractography using the Markov-Kennedy parcellation. With the “default” tractography parameter combination (curvature threshold: 0.2; distance correction: off), the percentage of connections correctly represented in the tractography-derived connectivity matrices was on average 79.21% (SD: 0.32) before any thresholding was performed. This, however, was not significantly greater than the null distribution estimated by 1000 randomly connected networks of the same size and density. The mean AUC was 0.68 (SD: 0.02), which was significantly greater than the null and corresponded to a very high sensitivity (M: 0.99, SD: 0.01) but very low specificity (M: 0.01, SD: 0.01). These results are in line with previous macaque studies using *ex vivo* specimens that suggested that probabilistic tractography is accurate at correctly detecting connections ([Azadbakht et al., 2015](#)) but trades off specificity for sensitivity ([Thomas et al., 2014](#)). Precision was, on average, 0.79 (SD: 0.002) for the default parameter settings and significantly greater than the null, indicating a high positive predictive value in DWI tractography (i.e., the great majority of positive results are true positives) despite the high density of the networks.

Curvature thresholds in tractography algorithms constrain the extent to which estimated streamlines can turn as they propagate. By default, FSL's algorithm uses a threshold of 0.2, corresponding to $\sim 78^\circ$. We systematically lowered this threshold (0.4, 0.6, 0.8 or $\sim 66^\circ$, $\sim 53^\circ$, $\sim 37^\circ$) to examine its effect on the accuracy of tractography. There was an effect of curvature threshold on the percentage of correctly detected connections of the unthresholded matrices (i.e., where the x-axis = 0, [Fig. 2A](#);

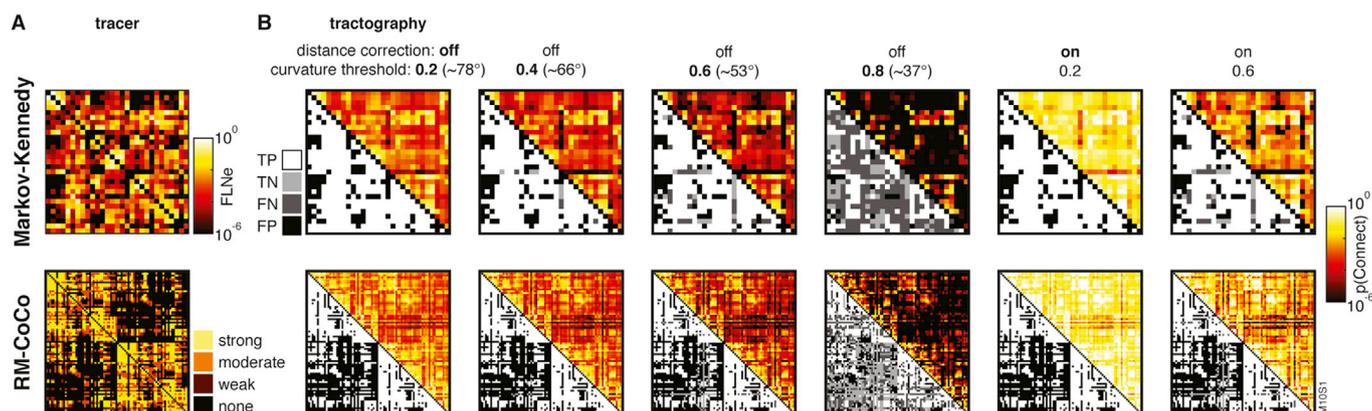


Fig. 1. (A) Tracer-derived connectivity matrices from [Markov et al. \(2014\)](#) (top) and CoCoMac ([Shen et al., 2012](#); [Stephan et al., 2001](#)) (bottom). (B) Tractography-derived matrices (upper triangle) for an example subject for each parcellation (Markov-Kennedy, top; RM-CoCo, bottom) using various tractography parameters. Accuracy of each connection, as compared to tracer-derived matrices, depicted in lower triangles (TP: true positive; TN: true negative; FN: false negative; FP: false positive). For the RM-CoCo parcellation, left hemisphere ROIs are ordered together followed by right hemisphere ROIs, such that inter-hemispheric quadrants are the upper right and lower left of each matrix.

Intrahemispheric: Markov-Kennedy

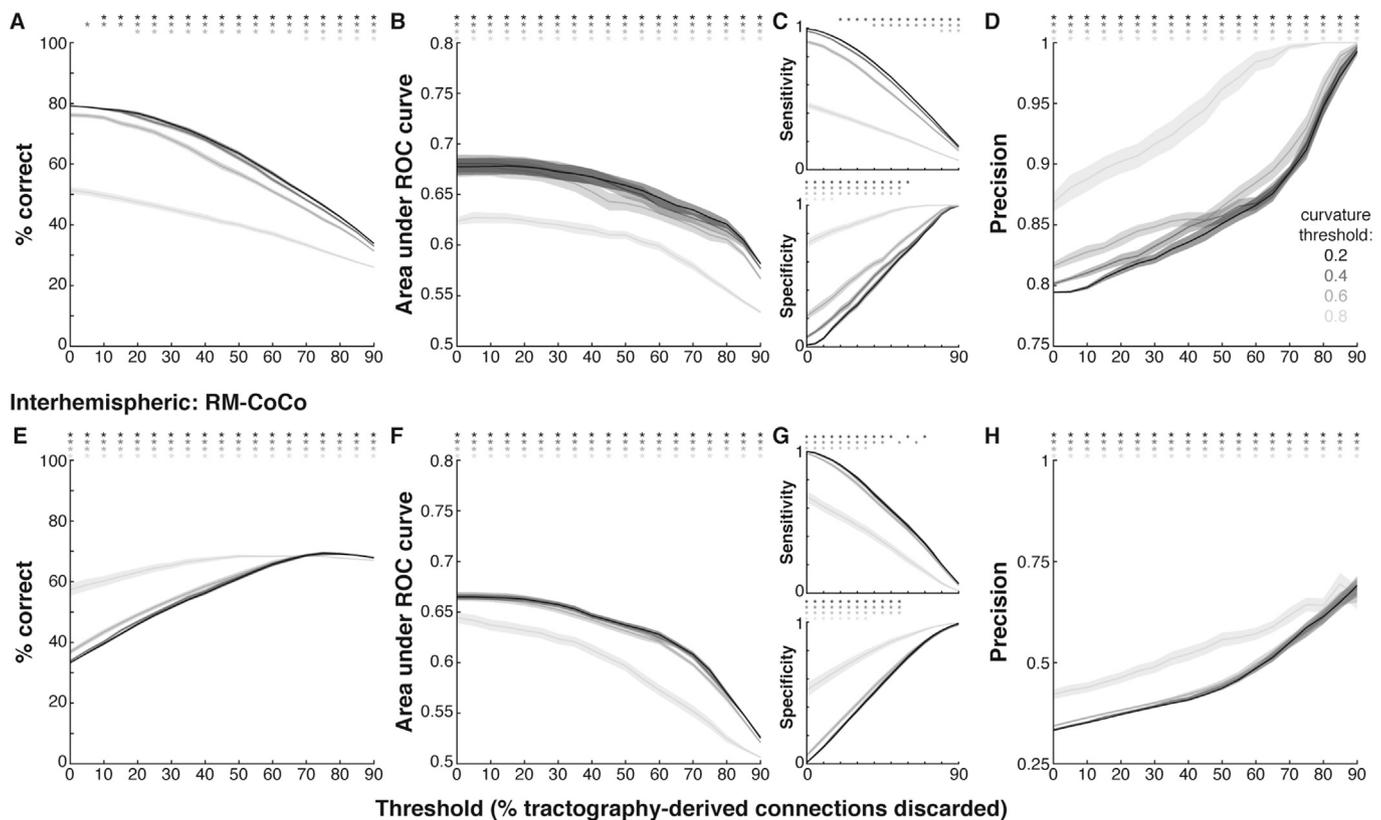


Fig. 2. Accuracy of DWI tractography. (A–D) Accuracy measures for tractography using the Markov-Kennedy parcellation. (E–H) Accuracy measures for interhemispheric tractography using the RM-CoCo parcellation. Curves shown correspond to accuracy using different tractography curvature thresholds as a function of thresholding the tractography-derived connectivity matrices (i.e., discarding connections having the lowest proportion of streamline counts). Points along the curves that are >95% confidence interval derived from the distributions of accuracy measures computed using 1000 randomly generated networks of the same size and densities are denoted with *.

repeated measures one-way ANOVA, $F(3, 9) = 525.85$, $p < 0.001$). Notably, post hoc comparisons indicated that % correct was not significantly different between matrices derived using curvature thresholds of 0.2 and 0.4 (M: 79.14%, SD: 0.12) but was significantly lower for thresholds of 0.6 (M: 76.21%, SD: 0.63) and 0.8 (M: 51.31, SD: 1.02) (Tukey-Kramer tests, $p < 0.05$). For all curvature thresholds, however, % correct was not significantly greater than the null for unthresholded matrices, indicating that these levels of correctly detected connections are as expected for networks of corresponding density. The effect of curvature threshold on the AUC of unthresholded matrices was limited to differences between the lowest threshold (0.8) and all other thresholds (repeated measures one-way ANOVA, $F(3,9) = 39.83$, $p < 0.001$; post hoc Tukey-Kramer tests) (Fig. 2B). AUC was significantly greater than the same-density null for all curvature thresholds. Lowering the curvature threshold to 0.6 and below resulted in a significant drop in sensitivity (repeated measures one-way ANOVA, $F(3,9) = 676.26$, $p < 0.001$; post hoc Tukey-Kramer tests) with no differences for curvature thresholds of 0.2 and 0.4 (Fig. 2C, top). This was accompanied by a significant increase in specificity across all curvature thresholds (Fig. 2C, bottom; repeated measures one-way ANOVA, $F(3,9) = 530.79$, $p < 0.001$; post hoc Tukey-Kramer tests). Precision also significantly increased when the curvature threshold was lowered to 0.6 and below (repeated measures one-way ANOVA, $F(3,9) = 81.26$, $p < 0.001$; post-hoc Tukey-Kramer tests), with no pairwise differences between 0.2 and 0.4 (Fig. 2D). Notably, precision was significantly better than that of the same-density null networks across all curvature thresholds. Intrahemispheric tractography using the RM-CoCo parcellation produced similar results (Fig. S1).

We also explored the accuracy of interhemispheric tractography using

the RM-CoCo parcellation. For the ‘default’ tractography parameter settings, interhemispheric tracking resulted in significantly lower % correct (paired t -test, $t(9) = -1873.4$, $p < 0.0001$) and precision ($t(9) = -5636.7$, $p < 0.0001$) while AUC was not different ($t(9) = 1.74$, $p = 0.12$) (Fig. 2E–H vs. Fig. S1). For these unthresholded matrices, all observed accuracy measures were significantly greater than those for the same-density null networks. Note that since available tracer data for interhemispheric connections are limited, many of the “absent” interhemispheric connections in the tracer matrix are due to a lack of anatomical data. We therefore performed the same analysis on the subset of interhemispheric connections for which the CoCoMac database indicates an explicitly present ($n = 479$) or absent ($n = 125$) connection. While accuracy of tractography was remarkably better for this subset of interhemispheric connections, it was still slightly lower than that of intrahemispheric tractography (Fig. S2). Interestingly, precision for the subset analysis was considerably higher in the analysis of explicit connections (Fig. S2D) than that for all interhemispheric connections (Fig. 2H), suggesting that the numerous false positives contributing to low precision in interhemispheric tracking may in part be due to the absence of evidence in the tracer dataset.

Just as in intrahemispheric tracking, there were no pairwise differences in % correct, AUC or precision values between curvature thresholds of 0.2 or 0.4 for interhemispheric tracking (repeated measures one-way ANOVAs, all $p < 0.001$; post-hoc Tukey-Kramer tests) (Fig. 2E–H). Observed accuracy measures were all significantly greater than for the same-density null networks. Together with the intrahemispheric tracking results, these findings suggest that using a high curvature threshold for macaque data does not result in a notable effect on the accuracy of DWI-

based tractography and instead may lower the specificity when reconstructing anatomical connections across the macaque brain.

The ability of probabilistic tractography to reconstruct white matter fiber tracts is thought to be limited by the distance between ROIs. Factors such as noise, artifacts and actual fiber trajectory increase the uncertainty of tracking with increasing distance (Li et al., 2012). To test whether this was the case for our data, we binned connections by distance and found that % correct dropped as a function of distance for intrahemispheric tracking (Fig. S3A), consistent with previous findings for intrahemispheric tractography (Donahue et al., 2016). There was no consistent effect of distance on % correct for interhemispheric tracking (Fig. S3B). Employing distance correction did little, if anything, to change accuracy measures (data not shown), since our accuracy measures are computed using binarized data and distance correction as implemented in FSL is simply a reweighting scheme that biases the number of streamlines detected for long-distance tracts rather than whether streamlines are detectable.

Distance is also a determining factor in actual connectivity probabilities as observed in tracer-based networks (Beul et al., 2017; Markov et al., 2013), suggesting that the distance between ROIs could be used to estimate the existence of a connection between them. To test this, we used a simple geodesic distance-based logistic regression model to generate connectivity matrices in the Markov-Kennedy parcellation. Remarkably, geodesic distance-based estimates of connectivity led to better correspondence with the tracer data (median AUC: 0.75) than DWI-based reconstructions (Fig. S4).

2.2. Effects of discarding “weakest” connections on accuracy

For the purposes of connectome creation, the outputs of probabilistic tractography algorithms are often thresholded by discarding connections whose streamline counts do not meet a minimum requirement (Roberts et al., 2017; Zalesky et al., 2016). To determine whether such a thresholding technique improves the accuracy of probabilistic tractography, we systematically thresholded our tractography-derived connectivity matrices by discarding between 5 and 90% of the “weakest” connections (i.e., those with lowest weights) in increments of 5%. Accuracy, as measured by % connections correctly detected and AUC, dropped as a function of thresholding the intrahemispheric connectivity matrix (Fig. 2A–B and S1A–B), with a significant drop occurring once 20% or more of the weakest connections were discarded, depending on the accuracy metric, parcellation and curvature threshold used (Table S1). However, the densest networks (i.e., those with the lowest thresholds) were also those whose % correct values were not significantly greater than the same-density null networks (Fig. 2A). AUC (Fig. 2B) and precision (Fig. 2D) was significantly greater than the null at all threshold levels. For interhemispheric tractography, only AUC dropped as a function of discarding the weakest weights, corresponding to a drop in sensitivity (Fig. 2F and G, but see Fig. S2), with a significant drop occurring once a 35% threshold or more was reached (Table S2). Thresholding the tractography-derived connectivity matrices using a consistency-based thresholding scheme (Roberts et al., 2017) instead resulted in similar effects on accuracy (Fig. S5).

2.3. Variability across individuals

Neuroanatomical differences exist between cynomolgous and rhesus macaques (e.g., Amaral et al., 2003) and could contribute to differences in our DWI connectomes. However, the differences in tractography results across animals were relatively small, as evidenced by the narrow range of standard errors in the accuracy figures (e.g., Fig. 2) and the results from the cynomolgous monkey were within the range of the rhesus monkeys (see Fig. S6A for an example). The cross-correlation between individual DWI connectomes was high and the cynomolgous monkey's correlation values were often within the range of the rhesus macaques' (see Figs. S6B–C for examples).

2.4. Do DWI-based connectomes accurately depict network characteristics?

To determine whether network characteristics were accurately captured in DWI-based connectomes, we first constructed an average DWI-based connectome for each parcellation using the set of “optimal” tractography parameters and thresholds that maximized AUC for each animal (see Methods, Fig. S7 and Table S2). We also included for analysis a DWI-based structural network in the RM-CoCo parcellation derived from control animals recently described by Grayson et al. (2017). This independent average network was constructed using different imaging sequences, image preprocessing and probabilistic tractography procedures from those described in the present paper. For replication purposes and to examine the dependence of our results on our specific sample and DWI sequences, we present analyses on this additional DWI-based network in the supplementary materials (see Fig. S8, S10–S11).

The edge weights of the average DWI-based network were correlated with those from the tract-tracing one for the Markov-Kennedy parcellation (Fig. 3A; Pearson correlation coefficient (r) = 0.71, $p < 0.0001$) and also for the RM-CoCo parcellation (Fig. 3B; Spearman rank correlation coefficient (r_s) = 0.45, $p < 0.0001$; also see Fig. S8B). This correlation is in line with (Donahue et al., 2016) or better than (van den Heuvel et al., 2015) previous studies, and suggests that the strength of a white matter fiber tract can be reasonably estimated using tractography-based methods. As distance correction biases the number of streamlines detected for long tracks, and therefore biases the weights of our DWI-based connectomes, we additionally constructed distance-corrected versions of the average DWI-based network for comparison with the tracer networks. The correlation between the tractography-based and the tract-tracing-based edge weights was worse than the correlation obtained without distance correction for both the Markov-Kennedy (Pearson correlation; $r = 0.51$, $p < 0.0001$; Fig. S9A) and RM-CoCo parcellation (Spearman rank correlation; $r_s = 0.40$, $p < 0.0001$; Fig. S9B). Finally, because connection strength varies as a function of distance in both tracer and tractography data (e.g., Donahue et al., 2016) we also computed the partial correlation between tracer and tractography weights while controlling for distance between ROIs. The correlation between the weights was reduced by nearly half for the Markov-Kennedy parcellation ($r = 0.37$, $p < 0.0001$) but only marginally for the RM-CoCo parcellation ($r_s = 0.43$, $p < 0.0001$).

We next computed a number of graph metrics that capture different levels of description of topology for both tracer- and DWI-based networks to determine the extent to which tractography-derived networks can accurately estimate network topology.

2.5. Centrality

Centrality measures are commonly used to provide estimates of the extent to which each node is embedded within a network, describing its potential contribution to network communication (van den Heuvel and Sporns, 2013a). Fig. 3 shows how the nodal degree for DWI- and tract-tracing-based networks was positively correlated for the Markov-Kennedy intrahemispheric parcellation (Fig. 3C; $r_s = 0.45$, $p < 0.01$) as well as the RM-CoCo whole brain parcellation (Fig. 3D; $r_s = 0.24$, $p = 0.01$). Betweenness centrality, however, was not correlated for either parcellation (Markov-Kennedy: $r_s = 0.14$, $p = 0.23$; RM-CoCo: $r_s = 0.14$, $p = 0.11$) (but see Fig. S8C). Network “hubs” are often singled out for investigation because of the special topological role they are thought to play in network communication and are identified as those nodes with high centrality. To determine whether hubs in the tractography-based networks coincide with those in the tracer-based networks, we identified hub nodes as those having centrality values greater than the 80th percentile for each centrality measure. Although some overlap exists in the identified hubs from tractography- and tracer-based networks (Fig. 3C–F; red data points), a number of hubs in the tracer-based networks were not considered hubs in the tractography-based networks (Fig. 3C–F; black data points) and vice

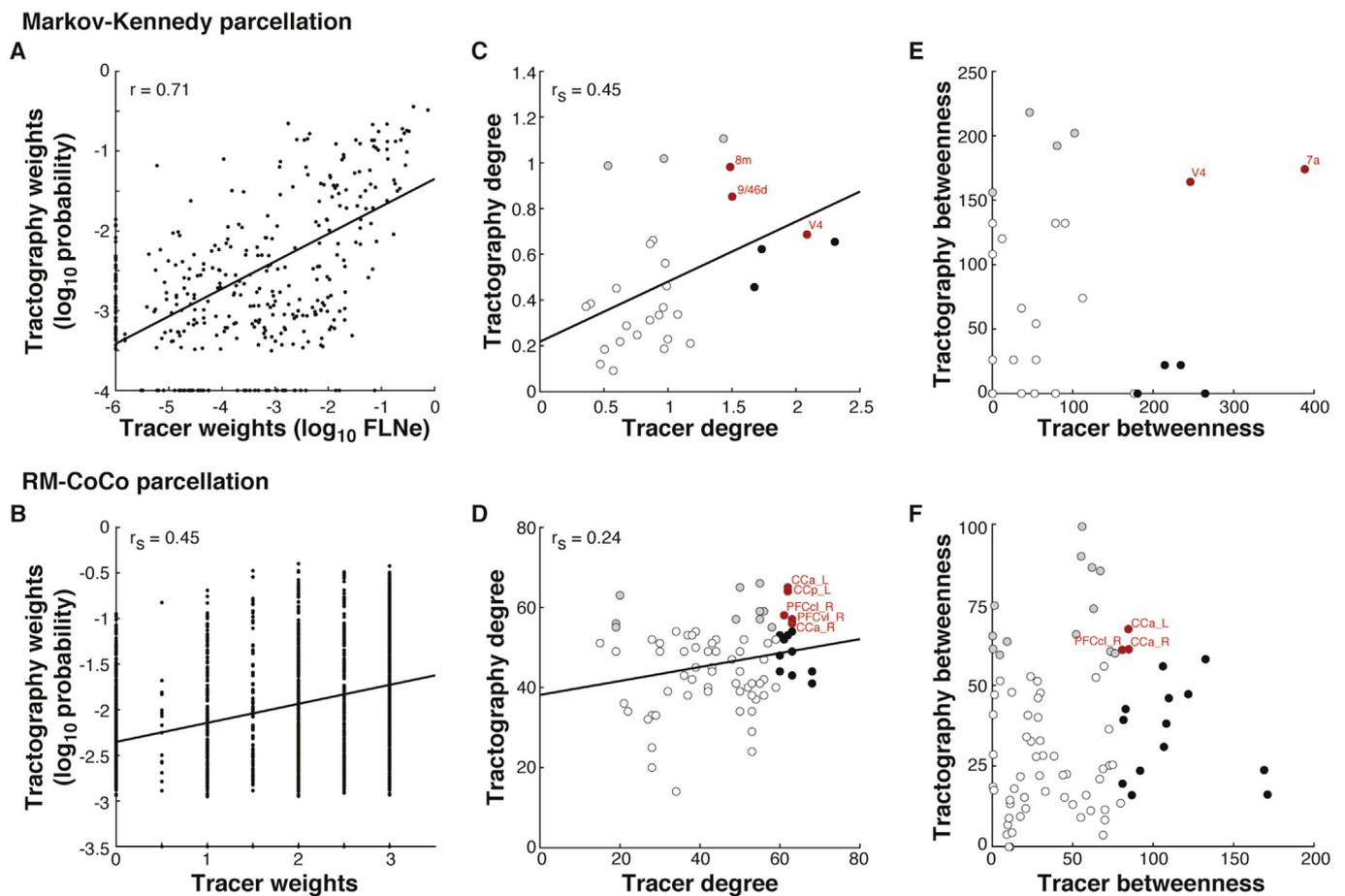


Fig. 3. Correspondence of network topology metrics of *in vivo* tractography and tract-tracing connectomes. (A–B) Connectome weight estimates from DWI tractography were well correlated with those from tracer studies. (C–F) Centrality estimates from DWI-based networks are correlated with those from tracer studies for degree but not betweenness centrality estimates. Only some hubs, identified as those with centrality >80th percentile, in the DWI-based networks correspond with hubs in tracer-based networks (red data points). These included cortical areas 7a, 8 m, 9/46d, V4 in the Markov-Kennedy parcellation (C,E), and anterior and posterior cingulate cortex (CCa and CCp) and centrolateral and ventrolateral prefrontal cortex (PFCcl and PFCvl) in the RM-CoCo parcellation (D, F). Hubs in tracer-based networks not identified as hubs in DWI-based networks denoted in black; misidentified hubs in DWI-based networks that are not hubs in tracer-based networks denoted in grey. Correctly identified hubs denoted in red. *r*: Pearson correlation coefficient; *r_s*: Spearman correlation coefficient.

versa (grey data points). Hypergeometric tests indicated no significant overlap between tracer and DWI hubs for either centrality measure (Markov-Kennedy: degree, $p = 0.07$; betweenness, $p = 0.21$; RM-CoCo: degree, $p = 0.63$; betweenness: $p = 0.42$; but see Figs. S8C–D). These findings suggest that tractography-based estimates of node centrality may not accurately reflect actual topologically central cortical regions.

2.6. Network architecture

2.6.1. Modularity

One common way to describe brain network architecture has been to decompose brain networks into smaller communities or modules that are responsible for more specialized functions, and the connections between communities as serving the potential to integrate across these functions (Meunier et al., 2010; Sporns and Betzel, 2016). We examined whether the modular organization of tractography-based networks accurately reflected those obtained from tract tracing. Tractography-based networks showed a remarkably similar organization of subnetworks as compared to the tracer-based networks (Fig. 4A). For the Markov-Kennedy parcellation, the modular organization differed only in its assignment of three nodes (F2, F5, ProM) to the prefrontal module in the tractography-based network (Fig. 4A top right, dark blue) rather than a more fronto-parietal one in the tracer-based network (Fig. 4A top left, light green). However,

even in the tracer-based network, these three nodes exhibit extensive and strong connectivity with the prefrontal module (Fig. 4A top left, grey edges). For the RM-CoCo parcellation, the decomposition of the tractography-based network resulted in a fourth module (Fig. 4A, bottom right, dark blue) and the assignment of some prefrontal areas (PFCdl, FEF, PMCm, PFCvl, PMCvl) to a more prefrontal module (light blue) rather than a more fronto-parietal one as in the tracer-based network (Fig. 4A, bottom left, light green). We determined the distance between the two sets of modules by computing the variation of information (VI) between them (Meilă, 2007). For both parcellations, the VI was significantly lower between the tractography-based partitions and the tracer-based ones as compared to the tracer-based null networks (Markov-Kennedy: 0.13 vs $0.70 \pm 4.2 \times 10^{-15}$; RM-CoCo: 0.27 vs $0.35 \pm 3.0 \times 10^{-15}$), suggesting that the tractography- and tracer-based partitions were more similar to each other than expected by chance.

We also assessed the accuracy of the tractography-based networks community assignments by first imposing the community structure of the tracer-based networks on the tractography-based ones and then computing the participation coefficients for each of the nodes in the tractography-based networks. The participation coefficient describes the extent to which each node is connected to nodes in other modules. If the “true” community assignments and, in particular, the between-module distribution of connections from the tracer-based networks were well

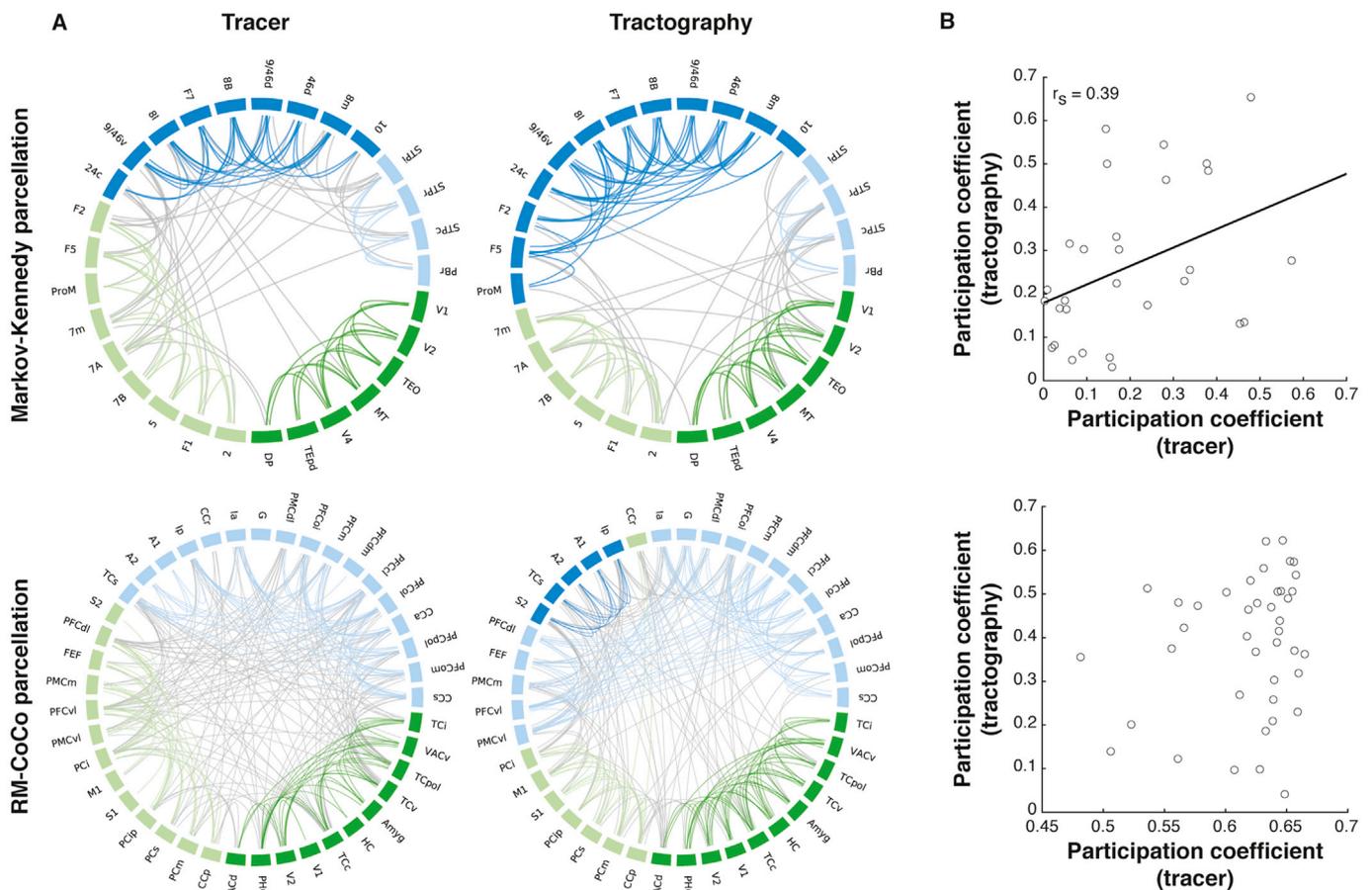


Fig. 4. Modularity partitions of tractography-based networks are well matched with those from tracer-based networks. (A) Connectogram depictions of modular structure of each network type for the Markov-Kennedy (top) and the RM-CoCo (bottom) parcellations. Community assignments are denoted by node color. Within-module edges are denoted with the same color as the module while between-module edges are denoted in gray. Edges were thresholded for ease of visualization. For the Markov-Kennedy parcellation, the visualization threshold was set to 33% for both network types. For RM-CoCo, the visualization threshold was set to keep connections in the strongest weight category (i.e., 3) in the tracer-based network and the tractography-based network was matched for the number of edges. (B) Correlation between tractography- and tracer-based network participation coefficients for the Markov-Kennedy (top) and RM-CoCo (bottom) parcellations. The community assignment of tracer-based networks were imposed on the tractography-based networks to determine participation coefficients.

estimated by the topology of the tractography-based networks, then the participation coefficients of the tractography-based network nodes computed in this manner should match those from the tracer-based networks. There was a moderate match for the Markov-Kennedy parcellation (Fig. 4B; $r_s = 0.39$, $p = 0.02$, Spearman rank correlation) but not for the RM-CoCo parcellation (Fig. 4B; $r_s = 0.03$, $p = 0.42$; also see Fig. S10). Together with the observed low VI between partition lists, these results suggest that nodal community assignments are well represented by DWI-based connectomes and the distribution of connections between and within modules can, to some extent, be estimated as well.

2.6.2. Rich club architecture

Brain networks have also been described as having a so-called “rich club” architecture, whereby a subset of high degree nodes exhibit dense connectivity with each other, often poised to mediate intermodular communication and forming a strong anatomical core (van den Heuvel and Sporns, 2013b). We examined whether a rich club architecture could be detected in the tracer-based networks, and the extent to which the tractography-based networks were able to replicate such findings. In the case of the Markov-Kennedy tracer network, it has been shown how its density restricts the detection of a rich club when using a binarized network for detection (Knoblauch et al., 2016). To determine whether this was also the case for the symmetrized version of the tracer network and whether considering the weights of the network would change rich

club detectability, we computed the normalized rich club coefficient by including network weights in addition to topology when generating the null models (Alstott et al., 2014). Similar to previous findings (Knoblauch et al., 2016), the Markov-Kennedy tracer network approached a rich club architecture at a degree of 24 ($p = 0.08$ for weighted networks, $p = 0.06$ for mixed networks) but rich club architecture was not consistently detected across a range of degrees and by and large not significant for any of the types of network considered even when weights were considered (Fig. 5A, left). Although the DWI-based Markov-Kennedy network showed an increase in the normalized rich club coefficient at high degree levels, none were significantly greater than 1 following FDR correction (Fig. 5A, right). The DWI network, like the tracer one, was unable to capture a rich club architecture for this parcellation.

As we have previously reported (Shen et al., 2015), the RM-CoCo tracer network exhibits a rich club architecture at multiple degree levels (Fig. 5B, left). The DWI-based RM-CoCo network also exhibits a rich club architecture at multiple degree levels for all three types of models considered (Fig. 5B, right). A hypergeometric test of significant rich club levels detected in this network when a mixed model was considered showed significant overlap with the tracer network in 3 of 14 levels (at $k = 55$, $p < 0.01$; at $k = 54$, $p = 0.01$; and at $k = 53$, $p = 0.03$; also see Fig. S11). For level $k = 55$, 10 rich club hubs that included regions of the prefrontal and cingulate cortex were identified in both the tracer- and DWI-based networks (Fig. 5C; red). Some additional regions

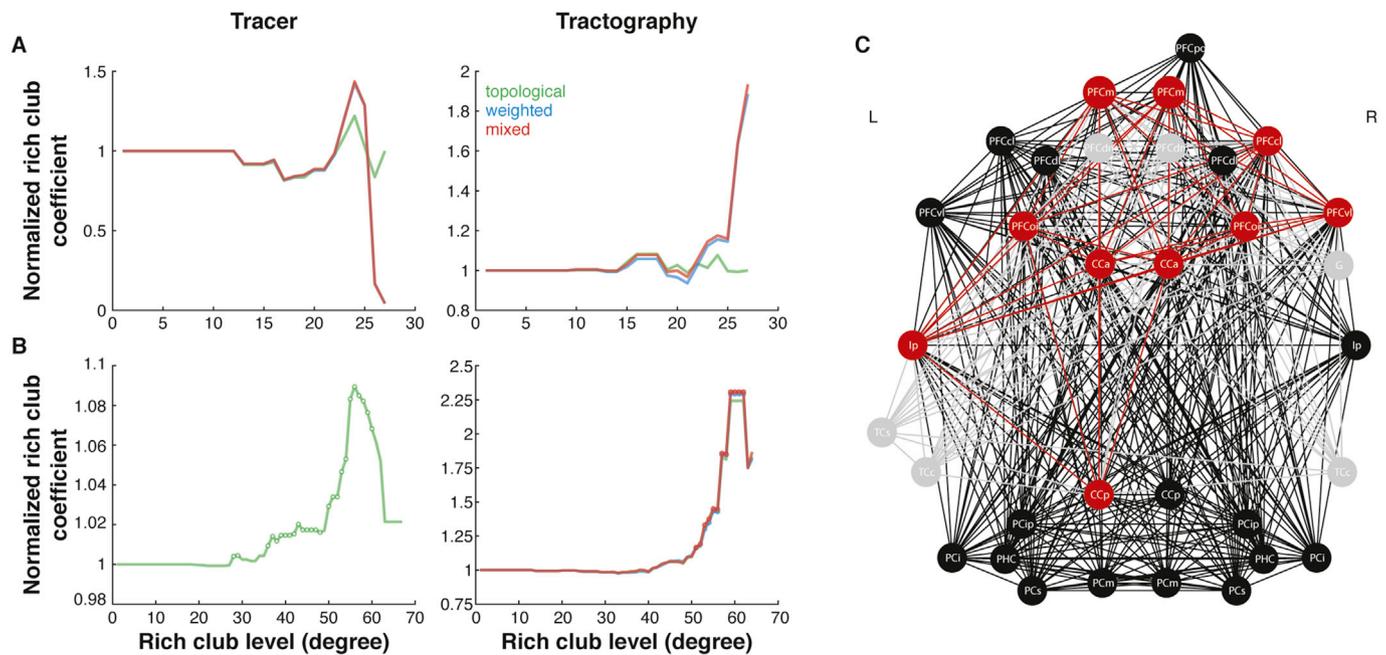


Fig. 5. Rich club (RC) architecture in tracer- and DWI-based networks. Normalized rich club coefficient for tracer- (left) and DWI-based (right) networks for the Markov-Kennedy (A) and RM-CoCo (B) parcellations. RC levels (i.e., normalized rich club coefficients significantly >1) denoted by circles. (C) RCs at degree level 55 for RM-CoCo parcellation. Red nodes and edges depict those that are common to both tracer- and DWI-based networks. RC nodes and edges incorrectly detected by DWI are depicted in grey, and those in tracer-based networks but missed by DWI are depicted in black.

of the temporal cortex along with dorso-medial prefrontal cortex were RC hubs in the DWI-based network but not the tracer-based one (Fig. 5C; grey), while a number of parietal and prefrontal RC hubs of the tracer-based network were notably missing from the DWI-based network (Fig. 5C; black).

2.6.3. Core-periphery architecture

For denser networks, such as the Markov-Kennedy tract-tracer one, a

core-periphery architecture has been described (Ercsey-Ravasz et al., 2013). Here, we first determined whether the core-periphery architecture previously reported in the Markov-Kennedy tracer network is detectable in the symmetrized version of the network and whether this could then be reconstructed by tractography-based connectomes. For the symmetrized tracer-based network, we detected a set of 18 nodes that contributed to the high-density core, with the 11 remaining nodes considered to be in the periphery (Fig. 6A). For the tractography-based

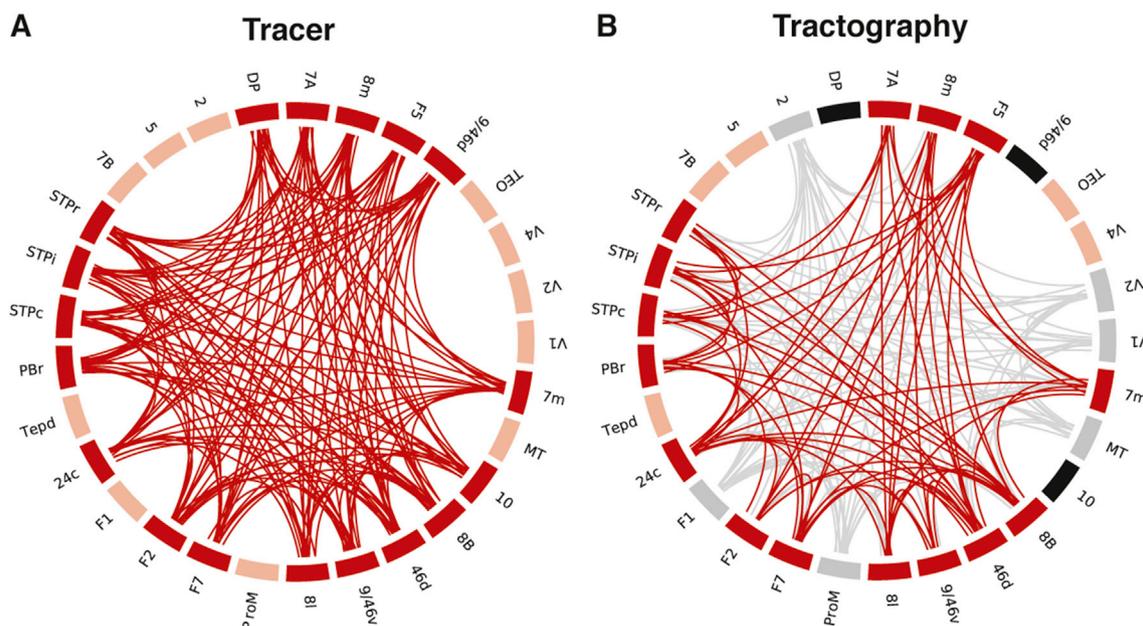


Fig. 6. Core-Periphery architecture in tracer- and DWI-based networks. (A) Core-periphery architecture detected in Markov-Kennedy tract tracing network. Core nodes and edges depicted in red, peripheral nodes depicted in coral. (B) Core-periphery architecture detected in the tractography-based network. Correctly identified core nodes and edges in red, peripheral nodes in coral. Incorrectly identified core nodes and edges depicted in grey, while core nodes that were missed depicted in black.

network, a core-periphery architecture was also detected, with the core consisting of 21 nodes, of which 16 were correctly identified (Fig. 6B). A hypergeometric test showed significant overlap in the core memberships of the tracer and tractography networks ($p < 0.05$).

3. Discussion

We have tested the performance of *in vivo* diffusion and tractography-based connectomes by comparing them to the gold standard connectomes from tracer data in macaque monkeys. We found that the reconstruction of individual connections to be moderately accurate, with a steep tradeoff between sensitivity and specificity that replicates previous *ex vivo* reports. We also found the proportion of streamlines detected between any two given regions can serve as a robust estimate of the number of axons that run between them, and that this relationship was dependent on the distance between regions. We then performed a series of validation studies on network topology metrics and demonstrated how the assignment of nodes into communities in tractography-based connectomes is fairly accurate and that a high-density rich club or core-periphery organization can be detected, just as they can be in the corresponding tracer-based networks. However, the proper identification of hubs within modules, and membership in rich club or core-periphery type architectures was less accurate, likely owing to the great number of false positive connections generated by probabilistic tractography. As network analysis has quickly become a popular approach for analyzing cortical connectomes, leading to the influential and expanding fields of connectomics and network neuroscience (Bassett and Sporns, 2017; Sporns, 2013), our findings are critical for the interpretation of network topology results based on *in vivo* measurements of structural connectivity.

3.1. Reconstruction of interareal cortico-cortical connections *in vivo*

Mapping the cortical connectome and uncovering its topological layout is a major ongoing research endeavour, involving many large-scale efforts like the Human Connectome Project (Van Essen et al., 2013). Structural connectomes *in vivo* can only be constructed with diffusion weighted imaging and tractography at present. The majority of such approaches aim to reconstruct the large-scale cortico-cortical connectivity matrix and subsequently analyze it using network metrics (Bassett et al., 2008; Gong et al., 2009; Hagmann et al., 2008). Here, we found the reconstruction of connections between the cortical areas to be above chance, but not highly accurate. Our obtained quantitative measures, such as AUC, are comparable to recent reports investigating intrahemispheric connections (Azadbakht et al., 2015; Donahue et al., 2016; Thomas et al., 2014; van den Heuvel et al., 2015). Our results provide additional quantitative evidence on the feasibility to correctly uncover the correct pairs of interconnected areas with diffusion imaging and tractography, by using two different benchmark datasets obtained by tract tracing. Despite large differences in how these two benchmark datasets were derived, how their connectivity was expressed and how they have differing network topologies, intrahemispheric tractography results for these two parcellations were remarkably similar. We add to the current understanding of connectome reconstruction accuracy by also examining interhemispheric tractography and show that it exhibits worse reconstruction quality than intrahemispheric tractography. This is consistent with our observation that tractography accuracy decreases with increasing distance between regions (Donahue et al., 2016; also see Li et al., 2012), which is only compounded when tracking across hemispheres. However, when we limited our analysis to only those interhemispheric connections that were explicitly defined in CoCoMac, the accuracy of DWI tractography approached that of intrahemispheric tracking. Precision, in particular, was considerably higher for this subset analysis, suggesting that the false positive rate decreased dramatically when we only considered explicitly defined connections. This suggests that, to some degree, missing information in CoCoMac about interhemispheric connections may be lowering our estimates of DWI

tractography accuracy.

We must note that the DWI and tracer datasets reported here were obtained from different subjects and, within each dataset, different ratios of macaque species (*Macaca mulatta* vs. *Macaca fascicularis*). The inaccuracies in tracking that we report may be attributed to the variability across different subjects and samples. A comparison of the results between the DWI tractography obtained from rhesus monkeys and the single cynomolgous yielded no qualitative differences but it is difficult to draw any substantive conclusions from such a small sample. Moreover, both cortical parcellations were defined on the F99 template (Van Essen, 2002), which is from a single rhesus macaque and adds yet another source of variability to our data. The extent to which each of these sources of variability affects connectome accuracy requires further examination in future studies. Of note, we found cross-subject correlations of DWI matrices to be relatively high. Together with a recent study of multiple large datasets in humans that reported relatively low variability across individual connectomes (Zimmermann et al., 2018), this suggests that individual differences in neuroanatomical pathways may not be well captured in large-scale DWI connectomes.

Tracer data has often been considered the “gold standard” for studying neuroanatomical pathways but, as with any experimental technique, suffers from its own limitations. These include variability across individual injections, the uptake of tracers by passing axonal fibers, as well as the distance travelled by particular tracers (for discussion, see Köbbert et al., 2000; Lanciego and Wouterlood, 2011). Some connections in the tracer matrices may be derived from only a few injections in a few animals or, in some cases, a single injection in a single animal. This is less likely for data from the CoCoMac database where multiple studies/injections may contribute to a single connection but it is still the case that certain regions, like those of the limbic system, are underrepresented (Bezgin et al., 2012). False negatives are also possible in tracer studies, especially for weak connections or injection sites with many connections to other areas (Markov et al., 2014). Combined with the fact that some areas can show widely variable connectivity depending on the subregion at the injection site (e.g., area F5: Gerbella et al., 2011; area F2: Markov et al., 2014), variability within the tracer data exists and has not been accounted for here.

Earlier validation studies have highlighted the inability of tractography to resolve long-range connections. Both probabilistic (Li et al., 2012) and deterministic (Dauguet et al., 2007; Zalesky and Fornito, 2009) tractography algorithms suffer from false negatives associated with long-range fibers due to the increasing uncertainty of tractography with distance (Jbabdi et al., 2015). Simply biasing connection weights towards long-distance connections using the distance correction option did little to resolve this problem as accuracy measures relied on the binary classification of the existence or non-existence of connections (but see Azadbakht et al., 2015). Instead, implementing the distance correction option affected the ability of tractography to estimate the strength of connections, decreasing it substantially when distance was estimated with the more realistic measure of geodesic distance rather than Euclidean distance.

Connectomes constructed using deterministic tractography are generally more sparse than those constructed using probabilistic tractography (Zalesky et al., 2016), and can suffer from a large number of false negative connections (Bastiani et al., 2012; Gong et al., 2009). Recent reports of probabilistic tractography, including the results presented here, have additionally indicated that while the majority of ‘true connections’ are successfully reconstructed, they instead come at the price of a large number of false positive connections (e.g., Thomas et al., 2014). The choice between deterministic and probabilistic tractography then, can be considered as a choice between constructing low-sensitivity/high-specificity connectomes versus high-sensitivity/low-specificity ones. Relevant for the probabilistic tractography results presented here, excessive false positive connections have been reported as a major drawback of diffusion imaging and tractography in a validation study with simulated human brain data

(Maier-Hein et al., 2017). These findings, along with the observation that false positives have a much larger impact on estimates of network topology as compared to false negatives (Bastiani et al., 2012; Zalesky et al., 2016), should be explicitly taken into account as important limitations when interpreting results from diffusion imaging tractography. Our results indicate that thresholding the weakest weights in the tractography-based networks on the order of 20–30% did not affect the percentage of correctly detected connections. Moreover, thresholding on the order of 55–85% did not affect AUC as it decreased sensitivity while dramatically increasing specificity. This is consistent with previous estimates for optimizing the tradeoff between sensitivity and specificity (de Reus and van den Heuvel, 2013b; Donahue et al., 2016). Choosing to threshold by discarding the weakest weights, however, may result in also discarding weak true positives. Weak connections are known to be important in determining the brain's functional organization (Gallos et al., 2012; Goulas et al., 2015) and may be better represented in networks that have been constructed using methods that take the consistency of edge reconstruction across subjects into account (Roberts et al., 2017). When we used this consistency-based thresholding method, we found qualitatively similar accuracy results to our weight-based thresholding technique.

We also found that lower curvature thresholds, at least for macaque data, result in fewer false positives and greater specificity without greatly affecting other accuracy measures (Azadbakht et al., 2015; also see Dauguet et al., 2007). Whether this is a result of less cortical folding and, therefore, less convoluted white matter pathways in the macaque brain (Herculano-Houzel et al., 2010; Zilles et al., 2013) or whether it constitutes an indicative guide for human tractography remains to be seen.

Using the streamline count as a proxy of fiber density has been previously criticized because it is susceptible to differences in tract lengths, curvature and branching (Jones, 2010; Jones et al., 2013). However, we showed how the probability values obtained with tractography were significantly correlated with an explained variance in line with Donahue et al. (2016), and nearly twice that of van den Heuvel et al. (2015). Since it is clear from tracer studies that physical distance plays a large role in the existence and strength of connections (Beul et al., 2017; Markov et al., 2013), a cautionary note is needed when interpreting such validation results. We have shown that for predicting the presence/absence of connections, a model based on physical distance alone was able to achieve comparable and even higher AUC than the diffusion and tractography-based reconstructions. Moreover, the correlation between the strength of connections and the tractography probabilities was diminished when physical distance was taken into account. Physical distance-based models (e.g., Bezgin et al., 2017; Ercsey-Ravasz et al., 2013) may therefore offer a more stringent baseline than using tractography alone while advancements in both imaging and tractography methods are still needed for the accurate reconstruction of cortical connectomes *in vivo*.

3.2. Investigating network topology of the cortex *in vivo*

The success of some network metrics but not others in the tractography-based connectomes was dependent on their resilience to rewirings. We found the partitioning of the cortico-cortical network into modules to be highly similar between the invasive and non-invasive connectivity datasets. These results bestow some confidence in module partitioning results obtained *in vivo*. This is in line with recent work that showed how false negatives, and even false positives, in connectomes affect modularity partitioning minimally (Zalesky et al., 2016). Partitioning brain networks into modules can result in variable communities across iterations (Sporns and Betzel, 2016). To minimize the effects of unstable partitionings, we chose to use the most consistent community structure detected from multiple iterations of partitioning. Additional work is still needed to fully assess how the accuracy of partitioning affects comparisons of modularity between different networks.

The participation coefficient, a higher-order network metric commonly used to identify intra-modular “provincial” and inter-modular “connector” hubs based on their cross-modular edges, was not consistent across the datasets. Our results suggested that for the Markov-Kennedy parcellation, inaccuracies mostly arose from the reconstruction of inter- and intra-modular connections as the participation coefficients were correlated even when we controlled for differences in community structure by keeping the partitioning scheme fixed when computing participation coefficients. For the RM-CoCo parcellation, however, there was an additional cost from the slightly inaccurate classification of nodes into their respective communities. Indeed, two networks can have extremely similar modularity partitions but their underlying connections could be statistically independent.

The susceptibility of centrality measures to rewirings resulted in discrepancies between the results obtained from tracer and tractography data. While we found a significant positive correlation for degree centrality, we found no significant correlations for betweenness centrality in our dataset and only moderate correlations in a second dataset (see Supplemental Material). The top most connected nodes or “hubs” between the two modalities also did not fully overlap when using either centrality metric. Our results indicate that more confidence can be assigned to degree as compared to betweenness centrality when tractography-derived connectomes are used. Along similar lines, global descriptions of structural organization like rich-club or core-periphery architectures, if they existed in the tracer networks, could be obtained from the tractography-based networks. However, the particular identification of nodes as hubs within these architectures was less accurate, owing again to the susceptibility of the identification to rewirings. Taken together, these results suggest that caution must be taken when using DWI-based tractography for identifying hubs, as identification is extremely susceptible to false connections.

4. Materials and Methods

4.1. Data acquisition

Data were collected from 10 male adult macaque monkeys (9 *Macaca mulatta*, 1 *Macaca fascicularis*, age 5.8 ± 1.9 years). A subset of these animals ($N = 3$) had MRI-compatible dental acrylic implants anchored to the skull with ceramic bone screws. All surgical and experimental procedures were approved by the Animal Use Subcommittee of the University of Western Ontario Council on Animal Care and were in accordance with the Canadian Council of Animal Care guidelines.

Surgical preparation and anaesthesia protocols have been previously described (Hutchison et al., 2011). Briefly, animals were anaesthetized before their scanning session and anaesthesia was maintained using 1.0–1.5% isoflurane during image acquisition. Images were acquired using a 7-T Siemens MAGNETOM head scanner with a high performance gradient (Siemens AC84 II; $G_{max} = 80$ mT/m; $SlewRate = 400$ T/m/s and an in-house designed and manufactured 8-channel transmit, 24-channel receive coil optimized for the primate head. For each monkey, two diffusion weighted scans were acquired with opposite phase encoding in the superior-inferior direction at 1 mm isotropic resolution. For seven animals, 2D EPI diffusion data were acquired (Siemens Advanced Diffusion WIP 511) with TE/TR = 48.8 ms/7500 ms, $b = 1000$ s/mm², 64 directions, 104×104 matrix, 24 slices, iPat = 3 and bandwidth of 1923 Hz/px. For the remaining 3 animals, a multiband EPI diffusion sequence (Feinberg et al., 2010; Moeller et al., 2010) was used; TE/TR = 47 ms/6000 ms, multiband = 2, $b = 1000$ s/mm², 64 directions, 128×128 matrix, 24 slices, iPat = 2, Partial fourier = 5/8 and bandwidth of 1502 Hz/px. A 3D T1w structural reference was collected for all animals using an MP2RAGE (Marques et al., 2010) acquisition at 500 μ m isotropic resolution; TE/TR = 3.15 ms/6500 ms, T1/TI2 = 800 ms/2700 ms, flip1/flip2 = 4/5, 256×256 matrix, 128 slices, iPat = 2 and 240 Hz/px bandwidth.

4.2. Image preprocessing

Diffusion-weighted image preprocessing was implemented using the FMRIB Software Library toolbox (FSL v5). This consisted of susceptibility-induced distortion correction using FSL's 'topup' (similar to Andersson et al., 2003) and 'eddy' (Andersson and Sotiropoulos, 2016) functions, and modeling of multiple fiber directions using FSL's 'bedpostx' function (Behrens et al., 2007). ROI parcellations specified in F99 macaque template space were registered using the Advanced Normalization Tools (ANTS) software package (Avants et al., 2011) to each animal's T1w anatomical image using a nonlinear registration and then linearly registered to diffusion space. Seed and target ROI masks were defined as the white matter (WM) voxels adjacent to each gray matter (GM) ROI, referred to as the GM-WM interface. An exclusion mask for each seed mask was also created using the GM voxels adjacent to the seed mask. For intrahemispheric tracking, exclusion masks of the opposite hemisphere were also used.

4.3. Tractography procedures

Two distinct parcellation schemes were chosen to match available tract-tracing data. The first ('Markov-Kennedy') was an intrahemispheric parcellation of 29 ROIs that matched those contributing to the edge-complete connectivity matrix described in Markov et al. (2014) (Fig. 1A, top). This parcellation as defined on the F99 atlas was downloaded from the SumsDB database [<http://brainvis.wustl.edu/wiki/index.php/Sums>About>]. The second ('CoCo-RM') was a whole-cortex parcellation of 82 ROIs matching the connectivity matrix described in Shen et al. (2012) (also see Kötter and Wanke, 2005) (Fig. 1A, bottom). This parcellation was first drawn on the F99 surface (Bezgin et al., 2008) and converted to a labeled volume with a 2 mm extrusion (Shen et al., 2012) using the Caret software package (Van Essen and Dierker, 2007) [<http://brainvis.wustl.edu/wiki/index.php/Caret>About>].

Tractography was performed between all ROIs using both parcellation schemes with FSL's 'probtrackx2' function. Parameters used for tracking were: 5000 seeds, 2000 steps, 0.5 mm step length, termination of paths that loop back on themselves and rejection of paths that pass through an exclusion mask. Curvature threshold was varied (0.2, 0.4, 0.6, 0.8) and distance correction was toggled on and off. The distance correction option produces a connectivity distribution that is the expected length of the streamlines that cross the voxel multiplied by the number of samples that cross it. In effect, the distance correction option serves to increase the weights of long-distance connections. The "default" parameter combination was considered as that commonly used in human studies (curvature threshold: 0.2; distance correction: off).

A structural connectivity matrix for each parameter set in each parcellation in each animal was generated by taking the number of streamlines detected between each ROI pair and dividing it by the total number of streamlines that were successfully sent from the seed mask (i.e., those that were not rejected or excluded). Each connectivity matrix was subsequently symmetrized.

For distance-based analyses, the distance between ROIs in the Markov-Kennedy parcellation was determined as the geodesic distance of their barycenters, that is, the shortest distance passing through the white matter that connects the barycenters of a pair of areas (as available from the Core-Nets database: core-nets.org). The distance between ROIs in the RM-CoCo parcellation was computed as the Euclidean distance between the centers of all ROIs. For both parcellations, connections were binned according to eight distance quantiles for analysis.

4.4. Data analysis

4.4.1. Assessment of interareal connection accuracy

For determining the accuracy of each subject's DWI-based structural connectivity matrices (i.e., analyses illustrated in Fig. 2), the corresponding tracer matrices were symmetrized (Fig. 1A) and all matrices

were binarized. Accuracy measures were computed for each subject's connectivity matrix corresponding to each combination of tractography parameters and thresholding and are reported as group means \pm SD. % correct was computed as: $\frac{TP+TN}{TP+FP+TN+FN}$, where *TP* are true positives, *TN* are true negatives, *FP* are false positives, and *FN* are false negatives (after Azadbakht et al., 2015). We additionally computed the area under the ROC curve (AUC), sensitivity, specificity, and precision of the DWI-derived matrices. Comparisons were only performed on the upper triangles of matrices. Repeated measures one-way ANOVAs were performed to assess the effect of curvature threshold on accuracy, treating the curvature thresholds as four manipulations on the same group of subjects.

To account for the effects of density on the accuracy measures, we generated a null distribution for each tractography parameter and threshold combination by averaging the observed DWI-derived networks across subjects and then producing a set of 1000 randomly connected undirected networks of the same size and density using the Brain Connectivity Toolbox (BCT; <https://sites.google.com/site/bctnet>) function 'makerandCLJ_und.m'. Accuracy measures were then computed for each set of 1000 networks and the 95% confidence interval of the null accuracy distributions was determined. Statistically significant accuracy measures were considered as those greater than the null mean + 95% CI.

To quantify the extent to which physical distance alone can predict the existence of connections between cortical areas, a logistic regression model was used in combination with the Markov-Kennedy tracer dataset. The existence of connections served as the binary dependent variable while the physical geodesic distance between the barycenters of cortical areas served as the independent variable. The model parameters were estimated in a training set consisting of 70% of the total number of area pairs and the rest of the pairs were used to estimate the ROC curve. The procedure was repeated 1000 times and each time the training set was assembled by sampling with replacement. The reported ROC curves and AUC values correspond to these 1000 iterations.

4.4.2. Assessment of reconstructed structural topology

For network analyses, average DWI-based structural connectivity matrices for both the Markov-Kennedy and RM-CoCo parcellations were generated by selecting, for each subject, the matrix having maximum AUC when compared to the tract-tracing data. The combination of tractography parameters and thresholding that yielded the maximum AUC for each subject in each parcellation is provided in Table S2. In an alternative approach, a fixed combination of tractography parameter settings and thresholds was applied to all animals based on the averaged AUC values. This alternative method for constructing the DWI matrix did not lead to substantive differences in results (data not shown). To enable direct comparison of tracer- and tractography-based networks, tracer matrices were symmetrized. Consequently, network analytic results presented here on the tracer matrices differ slightly from previous studies (e.g., Ercsey-Ravasz et al., 2013; Knoblauch et al., 2016; Shen et al., 2015, 2012). Tractography-based networks were then thresholded to match the density of the corresponding tracer-derived networks. In the case of the RM-CoCo parcellation, intra- and interhemispheric quadrants of the tractography-based network were treated with different thresholds due to their vast differences in density in the tract-tracing network.

The edge weights of the averaged tractography networks were correlated with those from the tracer networks to determine the extent to which tractography could capture the strength of a white matter fiber tract. For the Markov-Kennedy parcellation, the tractography edge weights followed a lognormal distribution, consistent with a previous report for the corresponding tracer weights (Markov et al., 2014). As such, a Pearson correlation was used for the normally-distributed logarithmized weights from the Markov-Kennedy parcellation. This also allowed for direct comparison with previous studies that have used Pearson correlations to report the relationship between tracer and tractography derived weights. The tracer weights for the RM-CoCo

parcellation, however, are categorical and a Spearman rank correlation was instead used for relating the two sets of edge weights for the RM-CoCo parcellation. Statistical significance of all correlations was assessed using permutation tests by resampling data pairs without replacement 10,000 times.

For the remaining network analyses, the Markov-Kennedy parcellation edges in both tracer- and tractography-based networks were treated as weighted. For the RM-CoCo parcellation, because of the categorical nature of the weighted information in the CoCoMac database, edges were binarized for both tracer and tractography networks when centrality measures (degree and betweenness) were computed. Measures of centrality, modularity partitioning and participation coefficients were obtained using functions from the BCT. For weighted graphs, the degree of each node was computed as the sum of its edge weights while for binarized graphs, node degree was taken as the total number of its edges. For the calculation of betweenness centrality, the edge weights were inverted so that larger weights corresponded with shorter paths. Community detection was performed using the Louvain algorithm (Blondel et al., 2008). As we did not know how the small network of 29 nodes from the Markov-Kennedy parcellation should be partitioned, we first varied the resolution parameter (γ) between 0 and 2 in increments of 0.05 and determined the most commonly detected number of partitions >1 in that range. The minimum value of γ that produced that number of partitions was selected. For the RM-CoCo parcellation, we similarly varied the resolution parameter but selected a γ value that gave a reasonable number of partitions based on previous studies of whole-brain modularity in the macaque (Goulas et al., 2015; Harriger et al., 2012). Partitioning for both parcellations was then repeated 100 times using the selected γ value, and the most consistent partitioning was chosen for analysis. This was done independently for both tracer- and tractography-based networks. For the Markov-Kennedy parcellation partitions, both tracer ($\gamma = 0.65$) and tractography ($\gamma = 0.65$) networks were consistently partitioned 100/100 times. For the RM-CoCo parcellation, the most common partition of the tracer network ($\gamma = 1$) occurred 17/100 times while that of the tractography network ($\gamma = 0.95$) occurred 44/100 times. Spearman rank correlation coefficients were computed to assess the correspondence of network measures between the two modalities.

Rich club detection was performed following the procedures described by Alstott et al. (2014) for computing null networks that are topological, weighted, and of mixed topo-weighted form. Core-periphery detection was performed as described in Ercsey-Ravasz et al. (2013). Briefly, the cortico-cortical network was subject to a modified Bron-Kerbosch algorithm with both pivoting and degeneracy ordering (Eppstein et al., 2010). The algorithm detects all cliques up to the maximum size. A clique is a subset of the nodes of the network among which the maximum possible amount of connections exists. The core was defined as the union of all the nodes participating in the cliques of maximum size.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.neuroimage.2019.02.018>.

References

- Alstott, J., Panzarasa, P., Rubinov, M., Bullmore, E.T., Vértes, P.E., 2014. A unifying framework for measuring weighted rich clubs. *Sci. Rep.* 4, 7258. <https://doi.org/10.1038/srep07258>.
- Amaral, D.G., Behnia, H., Kelly, J.L., 2003. Topographic organization of projections from the amygdala to the visual cortex in the macaque monkey. *Neuroscience* 118 (4), 1099–1120. [https://doi.org/10.1016/S0306-4522\(02\)01001-1](https://doi.org/10.1016/S0306-4522(02)01001-1).
- Andersson, J.L.R., Skare, S., Ashburner, J., 2003. How to correct susceptibility distortions in spin-echo echo-planar images: application to diffusion tensor imaging. *Neuroimage* 20, 870–888. [https://doi.org/10.1016/S1053-8119\(03\)00336-7](https://doi.org/10.1016/S1053-8119(03)00336-7).
- Andersson, J.L.R., Sotiropoulos, S.N., 2016. An integrated approach to correction for off-resonance effects and subject movement in diffusion MR imaging. *Neuroimage* 125, 1063–1078. <https://doi.org/10.1016/j.neuroimage.2015.10.019>.
- Avants, B.B., Tustison, N.J., Song, G., Cook, P.A., Klein, A., Gee, J.C., 2011. A reproducible evaluation of ANTs similarity metric performance in brain image registration. *Neuroimage* 54, 2033–2044. <https://doi.org/10.1016/j.neuroimage.2010.09.025>.
- Azadbakht, H., Parkes, L.M., Haroon, H.A., Augath, M., Logothetis, N.K., De Crespigny, A., D'Arceuil, H.E., Parker, G.J.M., 2015. Validation of high-resolution tractography against in vivo tracing in the macaque visual cortex. *Cerebr. Cortex* 25, 4299–4309. <https://doi.org/10.1093/cercor/bhu326>.
- Bassett, D.S., Bullmore, E., Verchinski, B.A., Mattay, V.S., Weinberger, D.R., Meyer-Lindenberg, A., 2008. Hierarchical Organization of Human Cortical Networks in Health and Schizophrenia. *J. Neurosci.* 28.
- Bassett, D.S., Sporns, O., 2017. Network neuroscience. *Nat. Neurosci.* 20, 353–364. <https://doi.org/10.1038/nn.4502>.
- Bastiani, M., Shah, N.J., Goebel, R., Roebroeck, A., 2012. Human cortical connectome reconstruction from diffusion weighted MRI: The effect of tractography algorithm. *Neuroimage* 62, 1732–1749. <https://doi.org/10.1016/j.neuroimage.2012.06.002>.
- Baum, G.L., Ciric, R., Roalf, D.R., Betzel, R.F., Moore, T.M., Shinohara, R.T., Kahn, A.E., Vandekar, S.N., Rupert, P.E., Quarmley, M., Cook, P.A., Elliott, M.A., Ruparel, K., Gur, R.E., Gur, R.C., Bassett, D.S., Satterthwaite, T.D., 2017. Modular Segregation of Structural Brain Networks Supports the Development of Executive Function in Youth. *Curr. Biol.* 27, 1561–1572 e8. <https://doi.org/10.1016/j.cub.2017.04.051>.
- Behrens, T.E.J.E.J., Berg, H.J., Jbabdi, S., Rushworth, M.F.S.F.S., Woolrich, M.W.W., 2007. Probabilistic diffusion tractography with multiple fiber orientations: What can we gain? *Neuroimage* 34, 144–155. <https://doi.org/10.1016/j.neuroimage.2006.09.018>.
- Beul, S.F., Barbas, H., Hilgetag, C.C., 2017. A Predictive Structural Model of the Primate Connectome. *Sci. Rep.* 7, 43176. <https://doi.org/10.1038/srep43176>.
- Bezzin, G., Solodkin, A., Bakker, R., Ritter, P., McIntosh, A.R., 2017. Mapping complementary features of cross-species structural connectivity to construct realistic “Virtual Brains. *Hum. Brain Mapp.* 38, 2080–2093. <https://doi.org/10.1002/hbm.23506>.
- Bezzin, G., Vakorin, V.A., van Opstal, A.J., McIntosh, A.R., Bakker, R., 2012. Hundreds of brain maps in one atlas: registering coordinate-independent primate neuro-anatomical data to a standard brain. *Neuroimage* 62, 67–76. <https://doi.org/10.1016/j.neuroimage.2012.04.013>.
- Bezzin, G., Wanke, E., Krumnack, A., Kötter, R., 2008. Deducing logical relationships between spatially registered cortical parcellations under conditions of uncertainty. *Neural Network* 21, 1132–1145. <https://doi.org/10.1016/j.neunet.2008.05.010>.
- Blondel, V.D., Guillaume, J.-L., Lambiotte, R., Lefebvre, E., 2008. Fast unfolding of communities in large networks. *J. Stat. Mech. Theor. Exp.* 2008, P10008. <https://doi.org/10.1088/1742-5468/2008/10/P10008>.
- Crossley, N.A., Mechelli, A., Scott, J., Carletti, F., Fox, P.T., McGuire, P., Bullmore, E.T., 2014. The hubs of the human connectome are generally implicated in the anatomy of brain disorders. *Brain* 137, 2382–2395. <https://doi.org/10.1093/brain/awu132>.
- Dauguet, J., Peled, S., Berezovskii, V., Delzescaux, T., Warfield, S.K., Born, R., Westin, C.F., 2007. Comparison of fiber tracts derived from in-vivo DTI tractography with 3D histological neural tract reconstruction on a macaque brain. *Neuroimage* 37, 530–538. <https://doi.org/10.1016/j.neuroimage.2007.04.067>.
- de Reus, M.A., van den Heuvel, M.P., 2013a. Rich Club Organization and Intermodule Communication in the Cat Connectome. *J. Neurosci.* 33, 12929–12939. <https://doi.org/10.1523/JNEUROSCI.1448-13.2013>.
- de Reus, M.A., van den Heuvel, M.P., 2013b. Estimating false positives and negatives in brain networks. *Neuroimage* 70, 402–409. <https://doi.org/10.1016/j.neuroimage.2012.12.066>.
- Donahue, C.J., Sotiropoulos, S.N., Jbabdi, S., Hernandez-Fernandez, M., Behrens, T.E., Dyrby, T.B., Coalson, T., Kennedy, H., Knoblauch, K., Van Essen, D.C., Glasser, M.F., 2016. Using Diffusion Tractography to Predict Cortical Connection Strength and Distance: A Quantitative Comparison with Tracers in the Monkey. *J. Neurosci.* 36, 6758–6770. <https://doi.org/10.1523/JNEUROSCI.0493-16.2016>.
- Eppstein, D., Löffler, M., Strash, D., 2010. Listing All Maximal Cliques in Sparse Graphs in Near-optimal Time.
- Ercsey-Ravasz, M., Markov, N.T., Lamy, C., VanEssen, D.C., Knoblauch, K., Toroczkai, Z., Kennedy, H., 2013. A Predictive Network Model of Cerebral Cortical Connectivity Based on a Distance Rule. *Neuron* 80, 184–197. <https://doi.org/10.1016/j.neuron.2013.07.036>.
- Feinberg, D.A., Moeller, S., Smith, S.M., Auerbach, E., Ramanna, S., Glasser, M.F., Miller, K.L., Ugurbil, K., Yacoub, E., 2010. Multiplexed echo planar imaging for sub-second whole brain fMRI and fast diffusion imaging. *PLoS One* 5. <https://doi.org/10.1371/journal.pone.0015710>.
- Formito, A., Zalesky, A., Breakspear, M., 2015. The connectomics of brain disorders. *Nat. Rev. Neurosci.* 16, 159–172. <https://doi.org/10.1038/nrn3901>.

- Gallos, L.K., Makse, H.A., Sigman, M., 2012. A small world of weak ties provides optimal global integration of self-similar modules in functional brain networks. *Proc. Natl. Acad. Sci. U. S. A.* 109, 2825–2830. <https://doi.org/10.1073/pnas.1106612109>.
- Gerbella, M., Belmalih, A., Borra, E., Rozzi, S., Luppino, G., 2011. Cortical connections of the anterior (F5a) subdivision of the macaque ventral premotor area F5. *Brain Struct. Funct.* 216 (1), 43–65. <https://doi.org/10.1007/s00429-010-0293-6>.
- Gong, G., He, Y., Concha, L., Lebel, C., Gross, D.W., Evans, A.C., Beaulieu, C., 2009. Mapping Anatomical Connectivity Patterns of Human Cerebral Cortex Using In Vivo Diffusion Tensor Imaging Tractography. *Cerebr. Cortex* 19, 524–536. <https://doi.org/10.1093/cercor/bhn102>.
- Goulas, A., Schaefer, A., Margulies, D.S., 2015. The strength of weak connections in the macaque cortico-cortical network. *Brain Struct. Funct.* 220, 2939–2951. <https://doi.org/10.1007/s00429-014-0836-3>.
- Grayson, D.S., Bliss-Moreau, E., Bennett, J., Lavenex, P., Amaral, D.G., 2017. Neural Reorganization Due to Neonatal Amygdala Lesions in the Rhesus Monkey: Changes in Morphology and Network Structure. *Cerebr. Cortex* 53, 1–14. <https://doi.org/10.1093/cercor/bhx080>.
- Griffa, A., Baumann, P.S., Thiran, J.P., Hagmann, P., 2013. Structural connectomics in brain diseases. *Neuroimage* 80, 515–526. <https://doi.org/10.1016/j.neuroimage.2013.04.056>.
- Hagmann, P., Cammoun, L., Gigandet, X., Meuli, R., Honey, C.J., Wedeen, V.J., Sporns, O., 2008. Mapping the structural core of human cerebral cortex. *PLoS Biol.* 6, e159. <https://doi.org/10.1371/journal.pbio.0060159>.
- Harriger, L., van den Heuvel, M.P., Sporns, O., 2012. Rich club organization of macaque cerebral cortex and its role in network communication. *PLoS One* 7, e46497. <https://doi.org/10.1371/journal.pone.0046497>.
- Herculano-Houzel, S., Mota, B., Wong, P., Kaas, J.H., 2010. Connectivity-driven white matter scaling and folding in primate cerebral cortex. *Proc. Natl. Acad. Sci. U. S. A.* 107, 19008–19013. <https://doi.org/10.1073/pnas.1012590107>.
- Hutchison, R.M., Leung, S.L., Mirsattari, S.M., Gati, J.S., Menon, R.S., Everling, S., 2011. Resting-state networks in the macaque at 7T. *Neuroimage* 56. <https://doi.org/10.1016/j.neuroimage.2011.02.063>.
- Jbabdi, S., Sotiropoulos, S.N., Haber, S.N., Van Essen, D.C., Behrens, T.E., 2015. Measuring macroscopic brain connections in vivo. *Nat. Neurosci.* 18, 1546–1555. <https://doi.org/10.1038/nn.4134>.
- Jones, D., 2010. Challenges and limitations of quantifying brain connectivity in vivo with diffusion MRI. *Imaging Med.* 2, 341–355. <https://doi.org/10.2217/IM.10.21>.
- Jones, D.K., Knösche, T.R., Turner, R., 2013. White matter integrity, fiber count, and other fallacies: The do's and don'ts of diffusion MRI. *Neuroimage* 73. <https://doi.org/10.1016/j.neuroimage.2012.06.081>.
- Knoblauch, K., Ercsey-ravasz, M., Kennedy, H., Toroczkai, Z., 2016. The Brain in Space. In: Kennedy, H., Van Essen, D.C., Christen, Y. (Eds.), *Micro-, Meso- and Macro-Connectomics of the Brain*, pp. 45–74. <https://doi.org/10.1007/978-3-319-27777-6>.
- Köbber, C., Apps, R., Bechmann, I., Lanciego, J.L., Mey, J., Thanos, S., 2000. Current concepts in neuroanatomical tracing. *Prog. Neurobiol.* 62 (4), 327–351. [https://doi.org/10.1016/S0301-0082\(00\)00019-8](https://doi.org/10.1016/S0301-0082(00)00019-8).
- Kötter, R., Wanke, E., 2005. Mapping brains without coordinates. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 360, 751–766. <https://doi.org/10.1098/rstb.2005.1625>.
- Lanciego, J.L., Wouterlood, F.G., 2011. A half century of experimental neuroanatomical tracing. *J. Chem. Neuroanat.* 42 (3), 157–183. <https://doi.org/10.1016/j.jchemneu.2011.07.001>.
- Li, L., Rilling, J.K., Preuss, T.M., Glasser, M.F., Hu, X., 2012. The effects of connection reconstruction method on the interregional connectivity of brain networks via diffusion tractography. *Hum. Brain Mapp.* 33, 1894–1913. <https://doi.org/10.1002/hbm.21332>.
- Maier-Hein, K.H., Neher, P.F., Houde, J.-C., Côté, M.-A., Garyfallidis, E., Zhong, J., Chamberland, M., Yeh, F.-C., Lin, Y.-C., Ji, Q., Reddick, W.E., Glass, J.O., Chen, D.Q., Feng, Y., Gao, C., Wu, Y., Ma, J., Renjie, H., Li, Q., Westin, C.-F., Deslauriers-Gauthier, S., González, J.O.O., Paquette, M., St-Jean, S., Girard, G., Rheault, F., Sidhu, J., Tax, C.M.W., Guo, F., Mesri, H.Y., Dávid, S., Froeling, M., Heemskerk, A.M., Leemans, A., Boré, A., Pinsard, B., Bedetti, C., Desrosiers, M., Brambati, S., Doyon, J., Sarica, A., Vasta, R., Cerasa, A., Quattrone, A., Yeatman, J., Khan, A.R., Hodges, W., Alexander, S., Romascano, D., Barakovic, M., Auria, A., Esteban, O., Lemkaddem, A., Thiran, J.-P., Cetingul, H.E., Odry, B.L., Mailhe, B., Nadar, M.S., Pizzagalli, F., Prasad, G., Villalon-Reina, J.E., Galvis, J., Thompson, P.M., Requejo, F.D.S., Laguna, P.L., Lacerda, L.M., Barrett, R., Dell'Acqua, F., Catani, M., Petit, L., Caruyer, E., Daducci, A., Dyrby, T.B., Holland-Letz, T., Hilgetag, C.C., Stieltjes, B., Descoteaux, M., 2017. The challenge of mapping the human connectome based on diffusion tractography. *Nat. Commun.* 8, 1349. <https://doi.org/10.1038/s41467-017-01285-x>.
- Markov, N.T., Ercsey-Ravasz, M., Lamy, C., Ribeiro Gomes, A.R., Magrou, L., Misery, P., Giroud, P., Barone, P., Dehay, C., Toroczkai, Z., Knoblauch, K., Van Essen, D.C., Kennedy, H., 2013. The role of long-range connections on the specificity of the macaque interareal cortical network. *Proc. Natl. Acad. Sci. U. S. A.* 110, 5187–5192. <https://doi.org/10.1073/pnas.1218972110>.
- Markov, N.T., Ercsey-Ravasz, M.M., Ribeiro Gomes, A.R., Lamy, C., Magrou, L., Vezoli, J., Misery, P., Falchier, A., Quilodran, R., Gariel, M.A., Sallet, J., Gamanut, R., Huissoud, C., Clavagnier, S., Giroud, P., Sappey-Marinié, D., Barone, P., Dehay, C., Toroczkai, Z., Knoblauch, K., Van Essen, D.C., Kennedy, H., 2014. A weighted and directed interareal connectivity matrix for macaque cerebral cortex. *Cerebr. Cortex* 24, 17–36. <https://doi.org/10.1093/cercor/bhs270>.
- Marques, J.P., Kober, T., Krueger, G., van der Zwaag, W., Van de Moortele, P.F., Gruetter, R., 2010. MP2RAGE, a self bias-field corrected sequence for improved segmentation and T1-mapping at high field. *Neuroimage* 49, 1271–1281. <https://doi.org/10.1016/j.neuroimage.2009.10.002>.
- Meilá, M., 2007. Comparing clusterings-an information based distance. *J. Multivariate Anal.* 98, 873–895. <https://doi.org/10.1016/j.jmva.2006.11.013>.
- Meunier, D., Lambiotte, R., Bullmore, E.T., 2010. Modular and hierarchically modular organization of brain networks. *Front. Neurosci.* 4, 1–11. <https://doi.org/10.3389/fnins.2010.00200>.
- Moeller, S., Yacoub, E., Olman, C.A., Auerbach, E., Strupp, J., Harel, N., Ugurbil, K., 2010. Multiband multislice GE-EPI at 7 tesla, with 16-fold acceleration using partial parallel imaging with application to high spatial and temporal whole-brain fMRI. *Magn. Reson. Med.* 63, 1144–1153. <https://doi.org/10.1002/mrm.22361>.
- Park, H.-J., Friston, K., 2013. Structural and functional brain networks: from connections to cognition. *Science* 342, 1238411. <https://doi.org/10.1126/science.1238411>.
- Perry, A., Wen, W., Lord, A., Thalamuthu, A., Roberts, G., Mitchell, P.B., Sachdev, P.S., Breakspear, M., 2015. The organisation of the elderly connectome. *Neuroimage* 114, 414–426. <https://doi.org/10.1016/j.neuroimage.2015.04.009>.
- Roberts, J.A., Perry, A., Roberts, G., Mitchell, P.B., Breakspear, M., 2017. Consistency-based thresholding of the human connectome. *Neuroimage* 145, 118–129. <https://doi.org/10.1016/j.neuroimage.2016.09.053>.
- Shen, K., Bezgin, G., Hutchison, R.M., Gati, J.S., Menon, R.S., Everling, S., McIntosh, A.R., 2012. Information processing architecture of functionally defined clusters in the macaque cortex. *J. Neurosci.* 32, 17465–17476. <https://doi.org/10.1523/JNEUROSCI.2709-12.2012>.
- Shen, K., Hutchison, R.M., Bezgin, G., Everling, S., McIntosh, A.R., 2015. Network structure shapes spontaneous functional connectivity dynamics. *J. Neurosci.* 35, 5579–5588. <https://doi.org/10.1523/JNEUROSCI.4903-14.2015>.
- Sporns, O., 2014. Contributions and challenges for network models in cognitive neuroscience. *Nat. Neurosci.* 17, 652–660. <https://doi.org/10.1038/nn.3690>.
- Sporns, O., 2013. The human connectome: Origins and challenges. *Neuroimage* 80, 53–61. <https://doi.org/10.1016/j.neuroimage.2013.03.023>.
- Sporns, O., Betzel, R.F., 2016. Modular Brain Networks. *Annu. Rev. Psychol.* 67, 613–640. <https://doi.org/10.1146/annurev-psych-122414-033634>.
- Stephan K.E., Kamper, L., Bozkurt, A., Burns, G., Young, M., Kötter, R., 2001. Advanced database methodology for the Collation of Connectivity data on the Macaque brain (CoCoMac). *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 356, 1159–1186. <https://doi.org/10.1098/rstb.2001.0908>.
- Thomas, C., Ye, F.Q., Irfanoglu, M.O., Modi, P., Saleem, K.S., Leopold, D.A., Pierpaoli, C., 2014. Anatomical accuracy of brain connections derived from diffusion MRI tractography is inherently limited. *Proc. Natl. Acad. Sci. U. S. A.* 111, 16574–16579. <https://doi.org/10.1073/pnas.1405672111>.
- van den Heuvel, M.P., de Reus, M.A., Feldman Barrett, L., Scholtens, L.H., Coopmans, F.M.T., Schmidt, R., Preuss, T.M., Rilling, J.K., Li, L., 2015. Comparison of diffusion tractography and tract-tracing measures of connectivity strength in rhesus macaque connectome. *Hum. Brain Mapp.* 36, 3064–3075. <https://doi.org/10.1002/hbm.22828>.
- van den Heuvel, M.P., Mandl, R.C.W., Stam, C.J., Kahn, R.S., Hulshoff Pol, H.E., 2010. Aberrant Frontal and Temporal Complex Network Structure in Schizophrenia: A Graph Theoretical Analysis. *J. Neurosci.* 30.
- van den Heuvel, M.P., Sporns, O., 2013a. Network hubs in the human brain. *Trends Cognit. Sci.* 17, 683–696. <https://doi.org/10.1016/j.tics.2013.09.012>.
- van den Heuvel, M.P., Sporns, O., 2013b. An anatomical substrate for integration among functional networks in human cortex. *J. Neurosci.* 33, 14489–14500. <https://doi.org/10.1523/JNEUROSCI.2128-13.2013>.
- van den Heuvel, M.P., Sporns, O., 2011. Rich-Club Organization of the Human Connectome. *J. Neurosci.* 31, 15775–15786. <https://doi.org/10.1523/JNEUROSCI.3539-11.2011>.
- Van Essen, D.C., 2002. Surface-based atlases of cerebellar cortex in the human, macaque, and mouse. In: *Annals of the New York Academy of Sciences*. <https://doi.org/10.1111/j.1749-6632.2002.tb07588.x>.
- Van Essen, D.C., Dierker, D.L., 2007. Surface-based and probabilistic atlases of primate cerebral cortex. *Neuron* 56 (2), 209–225. <https://doi.org/10.1016/j.neuron.2007.10.015>.
- Van Essen, D.C., Smith, S.M., Barch, D.M., Behrens, T.E.J., Yacoub, E., Ugurbil, K., WU-Minn HCP Consortium, 2013. The WU-Minn Human Connectome Project: An overview. *Neuroimage* 80, 62–79. <https://doi.org/10.1016/j.neuroimage.2013.05.041>.
- Zalesky, A., Fornito, A., 2009. A DTI-Derived Measure of Cortico-Cortical Connectivity 28, pp. 1023–1036. <https://doi.org/10.1109/TMI.2008.2012113>.
- Zalesky, A., Fornito, A., Cocchi, L., Gollo, L.L., van den Heuvel, M.P., Breakspear, M., 2016. Connectome sensitivity or specificity: which is more important? *Neuroimage* 142, 407–420. <https://doi.org/10.1016/j.neuroimage.2016.06.035>.
- Zalesky, A., Fornito, A., Seal, M.L., Cocchi, L., Westin, C.-F., Bullmore, E.T., Egan, G.F., Pantelis, C., 2011. Disrupted Axonal Fiber Connectivity in Schizophrenia. *Biol. Psychiatry* 69, 80–89. <https://doi.org/10.1016/j.biopsych.2010.08.022>.
- Zilles, K., Palomero-Gallagher, N., Amunts, K., 2013. Development of cortical folding during evolution and ontogeny. *Trends Neurosci.* 36 (5), 275–284. <https://doi.org/10.1016/j.tics.2013.01.006>.
- Zimmermann, J., Griffiths, J., Schirner, M., Ritter, P., McIntosh, A.R., 2018. Subject-Specificity of the Correlation Between Large-Scale Structural and Functional Connectivity. *Netw. Neurosci.* 3 (1), 90–106. https://doi.org/10.1162/NETN_a.00055.
- Zuo, X.-N., He, Y., Betzel, R.F., Colcombe, S., Sporns, O., Milham, M.P., 2016. Human Connectomics across the Life Span. *Trends Cognit. Sci.* 11, 1–14. <https://doi.org/10.1016/j.tics.2016.10.005>.